



# STANDARD VALUES *in* BLOOD

*Being the first fascicle of a Handbook of  
Biological Data*

EDITED BY  
ERRETT C. ALBRITTON, A.B., M.D.  
*Ery Professor of Physiology  
The George Washington University*



Prepared under the Direction of the Committee  
on the Handbook of Biological Data  
AMERICAN INSTITUTE OF BIOLOGICAL SCIENCES  
THE NATIONAL RESEARCH COUNCIL

Philadelphia and London  
W. B. SAUNDERS COMPANY

COPYRIGHT, 1952, BY W. B. SAUNDERS COMPANY  
COPYRIGHT UNDER THE INTERNATIONAL COPYRIGHT UNION  
*All rights reserved. This book is protected by copyright. No part of  
it may be reproduced in any manner without written permission from  
the publisher, except for any purpose of the United States Government  
Made in the United States of America.*

*Reprinted March, 1953, and April, 1955*  
Library of Congress Catalog Card Number 52-12867

# Foreword

On January 31, 1949, the National Academy of Sciences - National Research Council contracted with the Wright Air Development Center, United States Air Force, to gather and compile for publication the more basic established data on the composition and reactions of blood. A compact volume was desired that would contain authoritative tabular data of most pressing need to scientists working in the various fields of biology, including the medical sciences. The present work, first issued as Air Force Technical Report No. 6039, is the result.

The direction of the work was entrusted to the Committee on the Handbook of Biological Data, an organ of the American Institute of Biological Sciences. The Institute is affiliated with the National Research Council as a unit in the Council's Division of Biology and Agriculture. The membership of the Committee is representative of the major fields in plant and animal biology.

Seeking the highest degree of authoritativeness for the work, the Committee recognized that the specialist in a field from which a table is drawn can best exercise the critical judgment necessary to evaluate and select data for an authoritative table. The specialist can best identify those data born of the most acceptable methods of measurement and those having the greatest likelihood, or actual history, of reproducibility in competent hands. The Committee accordingly prescribed that in the selection and review of data the broadest collaboration be sought among investigators in hematology and related fields.

To the Editor has fallen the responsibility of determining the table of contents and the format and composition of each table as it appears on the page, and of enlisting the aid of contributors to supply the necessary data and of reviewers to give the data independent evaluation. The editorial office has also been the focus of advice and counsel sought and received in generous measure from all fields of biology \*

Acknowledgment is made, on behalf of the Committee, to the Wright Air Development Center, United States Air Force, for the foresight and scientific judgment inherent in the commission to prepare this tabular monograph on blood, to the biologists of this and other countries whose generous devotion of time as contributors and reviewers has made possible the completion of the work as it stands, and to the many others, unlisted, who have given the Committee solicited advice.

Acknowledgment is made to Dr. Cloyd Heck Marvin, President of The George Washington University, for generously making it possible for a member of his faculty to assume the editorship; to Dr. J. W. Heim, member of the Committee and project director for the Wright Air Development Center, for his unflagging effort toward bringing to realization the idea of a handbook of tabular biological data, and to Dr. Ulrich K. Henschke for giving the manuscript meticulous and critical study.

Finally the Editor acknowledges with gratitude the loyalty and labors of his administrative assistant, Mrs. Dorothy C. Stafford, and of his staff of research analysts, Dr. Hameed Bacchus, Mr. Harold J. Berman, Mr. Albert Einheber, Mr. Howard L. Gordon, Mr. Louis P. Munan, Mr. William S. Spector and Dr. Wasley D. Yushok.

ERRETT C. ALBRITTON

---

\*Two other comparable collections of tabular data are nearing completion as the present work goes to press, one drawn from the field of nutrition and metabolism and the other from that of growth, reproduction and life history. Both plant and animal forms are included. Other collections are planned.

THE COMMITTEE ON THE HANDBOOK  
OF BIOLOGICAL DATA

THEODORE C. BYERLY, *Chairman*  
ERRETT C. ALBRITTON, *Executive Secretary*

DETLEV W. BRONK\*  
RALPH E. CLELAND\*  
HAROLD J. CONN  
FRANK P. CULLINAN\*  
A. P. GAGGE  
DAVID R. GODDARD

\*ex officio

CHARLES M. GOSS  
WILLIAM W. GREULICH  
FRANK G. HALL  
J. W. HEIM  
PAUL E. HOWE  
CLARENCE J. HYLANDER\*

E. MORTON JELLINEK  
MILTON O. LEE\*  
JOHN S. NICHOLAS\*  
ADOLPH H. SCHULTZ  
PAUL A. WEISS  
RAYMUND L. ZWEMER\*



EDITORIAL STAFF

ERRETT C. ALBRITTON, *Editor*  
DOROTHY C. STAFFORD, *Adm. Asst.*

HABEEB BACCIUS  
HAROLD J. BERMAN

ALBERT EINHEBER  
HOWARD L. GORDON  
LOUIS P. MUNAN

WILLIAM S. SPECTOR  
WASLEY D. YUSHOK

# Contributors and Reviewers

- Abelson, Neva M.  
 Abramson, Harold A.  
 Adler, Harry F.  
 Aebersold, Paul C.  
 Aggeler, Paul M.  
 Albert, A.  
 Alexander, Benjamin  
 Allen, Fred H.  
 Alles, Gordon A.  
 Allison, James Boyd  
 Almquist, Herman J.  
 Alsever, John B.  
 Alt, Howard L.  
 Altland, Paul Daniel  
 Altschule, Mark D.  
 Amberson, William R.  
 Andresen, Marjory I.  
 Andresen, P. H.  
 Armstrong, Arthur Riley  
 Armstrong, S. Howard, Jr.  
 Asdell, S. A.  
 Ashby, Winifred M.  
 Augustinsson, Klas-Bertil
- Baker, Carl  
 Baker, Gladys Lucille  
 Barrett, William C.  
 Barron, Donald H.  
 Baumann, C. A.  
 Bazett, H. C.  
 Beall, Desmond  
 Beard, Joseph W.  
 Beatty, Clarissa H.  
 Beckman, Harry  
 Bedson, Samuel P.  
 Beerstecher, Ernest, Jr.  
 Begg, Charles F.  
 Bell, J. Milton  
 Benotti, Joseph  
 Bensley, Edward H.  
 Bergeim, Olaf  
 Berliner, Robert W.  
 Berman, Lawrence  
 Bessey, Otto A.  
 Bessman, Samuel P.  
 Best, Charles H.  
 Bethell, Frank H.  
 Beyer, K. H.  
 Bing, Franklin C.  
 Bing, Richard C.  
 Bird, Robert M.  
 Bischoff, Fritz E.  
 Black, Edgar C.  
 Black, R. M.  
 Blincoe, Clifton R.  
 Block, Matthew Harold  
 Block, Richard J.
- Bloom, Marvin L.  
 Bloom, William  
 Boeri, Enzo  
 Bollman, Jesse L.  
 Bonnycastle, Desmond D.  
 Bowen, William J.  
 Boyd, William C.  
 Brain, Reginald T.  
 Brambel, Charles E.  
 Brand, Erwin  
 Braun, Werner  
 Brecher, George  
 Briggs, David R.  
 Brinkhous, Kenneth M.  
 Britton, C. J. C.  
 Brody, Samuel  
 Broun, Goronwy O.  
 Brown, Robert K.  
 Brown, William E.  
 Bruner, H. D.  
 Brunetti, Rosemary O.  
 Burch, Helen B.  
 Burnett, Lee  
 Bussabarger, Robert A.  
 Butler, Allan M.  
 Butt, Hugh R.  
 Byers, Sanford Oscar  
 Byerly, T. C.
- Cantarow, Abraham  
 Cantril, Simeon T.  
 Carr, C. Jelleff  
 Cartwright, George E.  
 Casey, Albert E.  
 Catchpole, H. R.  
 Caton, William F.  
 Cayer, David  
 Chambers, William H.  
 Chance, Britton  
 Chanutlin, Alfred  
 Charipper, Harry A.  
 Christensen, Halvor N.  
 Clark, Paul F.  
 Cohen, Philip P.  
 Cohn, Edwin J.  
 Cole, Harold Harrison  
 Cole, Warren H.  
 Collier, H. Bruce  
 Comroe, Julius H., Jr.  
 Conley, C. Lockard  
 Conn, Harold J.  
 Connor, Arthur  
 Consolazio, William V.  
 Copenhaver, W. M.  
 Corcoran, Arthur C.  
 Cotlove, Ernest  
 Cournand, Andre  
 Cox, Gertrude M.
- Crafts, Roger C.  
 Craige, A. H., Jr.  
 Creskoff, Adolph J.  
 Cronkite, Eugene P.  
 Crookshank, H. Robert  
 Csonka, Frank A.  
 Cutter, Richard D.  
 Cutting, Windsor C.
- Daft, Floyd S.  
 Dalldorf, Gilbert J.  
 Dameshek, William  
 D'Angelo, Savino A.  
 Danowski, T. S.  
 Darrow, Daniel C.  
 Davenport, Horace W.  
 Davidsohn, Israel  
 Davidson, Charles S.  
 Davidson, Leonard T.  
 Dawson, Alden B.  
 Day, Paul L.  
 Deamer, William C.  
 DeBeer, Edwin J.  
 DeBoer, Benjamin  
 Delfs, Eleanor  
 DeMarsh, Q. B.  
 Dempsey, Edward W.  
 Denstedt, Orville F.  
 deRobertis, Eduardo  
 deTakats, Geza  
 Diamond, Louis K.  
 Dickstein, Benjamin  
 Dieckmann, William Joseph  
 Diggs, Lemuel Whitley  
 Dill, D. B.  
 Dille, Roger S.  
 Dische, Zacharias  
 Doan, Charles A.  
 Doelle, George B.  
 Dole, Vincent P.  
 Domm, L. V.  
 Dorfman, Albert  
 Dorfman, Ralph I.  
 Dougherty, Thomas F.  
 Dounce, Alexander L.  
 Dowdy, Andrew H.  
 Drabkin, David L.  
 Drinker, Cecil K.  
 Dublin, I. N.  
 Dukes, H. H.  
 Dunn, Max S.  
 duVigneaud, Vincent
- Ecker, Enrique E.  
 Edelmann, Abraham  
 Edsall, John T.  
 Eggers, Harold E.

Ehrich, William E.  
Eichelberger, Lillian  
Eisenmenger, William J.  
Elman, Robert  
Elvehjem, C. A.  
Endicott, Kenneth M.  
Ernstene, Arthur C.  
Essig, E. O.  
Evans, Robley D.  
Evans, Titus C.  
Everett, John W.  
Everett, Mark R.

Fabian, Abraham A.  
Fairhall, Lawrence T.  
Fantl, Paul  
Farmer, Chester J.  
Farris, Edmund J.  
Fazekas, Joseph F.  
Fegler, George  
Feldman, Harry A.  
Ferguson, John H.  
Ferguson, J. K. W.  
Ferry, Ronald M.  
Fiddes, John  
Field, John B.  
Firminger, Harlan E.  
Fishman, William H.  
Flock, Eunice V.  
Florio, Lloyd J.  
Fonio, Anton  
Forbes, Thomas R.  
Fowler, Willis M.  
Fraps, Richard M.  
Freeman, Smith  
Friedell, Hymen L.  
Friedman, David L.  
Friedemann, Theodore E.  
Frommeyer, Walter B., Jr.  
Frontali, Gino  
Fry, Fred  
Furth, Jacob

Gaebler, Oliver H.  
Gamble, James L.  
Garber, Edward E.  
Gardner, Mary V.  
Garfield, Morton M.  
Gendel, Benjamin R.  
Gersh, Isidore  
Gibbs, Owen Stanley  
Gibson, John G.  
Glaser, Kurt  
Glasser, Otto  
Gomori, George  
Goodhart, Robert S.  
Goss, Charles M.  
Graber, Edward E.  
Gram, H. C.  
Granick, S.  
Gray, John S.  
Gray, M. Geneva  
Green, David E.  
Green, Harold D.  
Greenberg, David M.

Griffith, John Q.  
Guarino, Armand J.  
Guest, George M.  
Guest, M. Mason  
Gurd, Frank R. N.  
Gutman, Alexander B.  
Gutmann, George E.  
Gyorgy, Paul

Haag, Harvey B.  
Haberman, Sol  
Haden, Russell H.  
Halberg, Franz  
Hald, Pauline M.  
Hall, F. G.  
Ham, Thomas Hale  
Hamilton, Paul B.  
Hamilton, William F.  
Hammarsten, Greta  
Hammon, William McD.  
Hamre, Christopher J.  
Handler, Philip  
Handley, Charles O.  
Hanks, John H.  
Hansel, W.  
Hardin, Robert Calvin  
Harman, John W.  
Harrel, George T.  
Hart, J. Sanford  
Hartman, Carl G.  
Hastings, A. Baird  
Haugaard, Niels  
Hawkins, Rosemary D.  
Hawkins, W. W.  
Heath, Clark W.  
Heck, Frank J.  
Helm, J. W.  
Heller, Victor G.  
Helm, John D., Jr.  
Henderson, L. M.  
Hendry, Edward B.  
Henning, Norbert  
Henschke, Ulrich K.  
Hepler, Opal E.  
Herrin, Raymond C.  
Hess, Walter C.  
Hetherington, Duncan C.  
Hickam, John B.  
Hier, Stanley W.  
Hiestand, W. A.  
Higgins, George M.  
Hill, Robert M.  
Hiller, Alma E.  
Hilmer, Paul E.  
Hirschboeck, John S.  
Hisaw, Frederick L.  
Hitchcock, David I.  
Hoffman, George R.  
Hogan, Albert G.  
Hollaender, Alexander  
Holland, Albert H., Jr.  
Holman, H. H.  
Homburger, Freddy  
Hooker, Charles W.  
Horan, Margaret

Horecker, B. L.  
Horn, T. H.  
Howard, A. H.  
Howe, Paul E.  
Huggett, A. St. G.  
Huggins, Charles B.  
Hughes, Josiah S.  
Hughes, Walter L., Jr.  
Hunter, F. R.  
Hutchinson, G. Evelyn

Ingram, Marylou  
Israel, Martin C. G.  
Ivy, Andrew Conway

Jacobs, M. H.  
Jacobson, L. O.  
Jandorf, Bernard J.  
Jaques, L. B.  
Johnson, Robert E.  
Jones, Oliver P.  
Jordan, Harvey E.  
Junqueira, L. C. U.

Kagan, E. M.  
Kaplan, Nathan O.  
Kark, Robert M.  
Katzenelbogen, Solomon  
Kay, Herbert D.  
Kerr, Stanley E.  
Keutmann, E. Henry  
Keys, Ansel  
Kindred, James E.  
King, Earl Judson  
Kirk, John Esben  
Kirsner, Joseph B.  
Kisch, Bruno Z.  
Kochakian, Charles D.  
Koelle, George B.  
Kohn, Henry I.  
Koppanyi, Theodore  
Kornhauser, S. I.  
Kough, R. H.  
Kracke, Roy R.  
Krebs, Adolph T.  
Krebs, H. A.  
Krop, Stephen  
Krumhaar, Edward B.

Lait, H.  
Laki, Koloman  
Lalich, Joseph J.  
Lambertsen, Christian J.  
Lamport, Harold  
Langston, William C.  
Lardy, Henry A.  
Largent, Edward J.  
Larson, Edward  
Lasker, Gabriel Ward  
Latimer, Homer B.  
Latta, John S.  
Laughlin, William S.  
Lawrence, John S.  
Lawson, Hampden C.  
Leatham, James H.

Leavell, Byrd S.  
Leblond, C. P.  
Lee, Milton O.  
Lehninger, A. L.  
LePage, G. A.  
LeRoy, George V.  
Lesnick, Gerson J.  
Levey, Stanley  
Levine, Philip  
Levine, Victor E.  
Lewis, Howard B.  
Lewis, Jessica H.  
Lillenthal, Joseph L., Jr.  
Lillie, R. D.  
Limarzi, Louis R.  
Lindeboom, G. A.  
Link, Karl Paul  
Lipmann, Fritz  
Lippman, Richard W.  
Lisco, Hermann  
Little, J. Maxwell  
Logan, Milan A.  
London, Irving M.  
Looney, Joseph M.  
Lowry, Oliver H.  
Lucas, Alfred M.  
Lucia, Salvatore P.  
Luft, Ulrich C.  
Lukens, F. D. W.  
Lynch, Eleanor L.

McCall, Keith B.  
McCay, Clive M.  
McCutcheon, F. Harold  
McGehee, Edward H.  
McGilvery, R. W.  
McHenry, E. W.  
McLean, Franklin C.  
McManus, J. F. A.  
McQuarrie, Irvine  
McShan, W. H.

MacCardle, Ross C.  
Machella, Thomas E.  
MacNamee, Paul D.  
Maengwyn-Davies, Gertrude D.  
Magath, Thomas B.  
Manery, Jeanne F.  
Mann, Evelyn B.  
Mann, Frank C.  
Manning, Isaac H.  
Markson, David E.  
Mason, Harold L.  
Matson, G. Albin  
Mayerson, H. S.  
Meister, Alton  
Mendel, B.  
Mendell, Theodore H.  
Mendlowitz, Milton  
Menkin, Valy  
Merriman, John R.  
Merritt, Katherine K.  
Mertens, Elizabeth  
Metcalf, Robert L.  
Meyer, L. M.  
Meyer, Ovid O.

Michels, Nicholas A.  
Milbe, F. R.  
Miller, A. T., Jr.  
Miller, Franklin R.  
Miller, Morell W.  
Milne, John  
Milstone, J. Haskell  
Minor, Allen H.  
Mirsky, I. Arthur  
Montagna, William  
Moore, Carl V.  
Morgan, Karl Z.  
Morrison, Ira R.  
Morrison, Peter R.  
Mugrage, Edward R.  
Muir, A. H.  
Mulligan, Richard M.  
Munson, Paul L.

Neal, Paul A.  
Newell, R. R.  
Nice, Leonard B.  
Noback, Charles R.

Oglesby, William T.  
Olwin, John H.  
Orten, James M.  
Osborne, Stafford L.  
Osgood, Edwin E.  
Otis, Arthur B.  
Overman, Richard R.  
Owen, Charles A.  
Owen, Philip S.  
Owren, Paul A.

Page, Karl E.  
Palm, Eric  
Pappenheimer, John R.  
Parrott, Ernest M.  
Paschkis, Karl E.  
Pasquier, M. A.  
Patek, Arthur J., Jr.  
Pearson, Paul B.  
Penrod, Kenneth E.  
Persike, Edward C.  
Peters, John H.  
Peters, John P.  
Pett, Lionel Bradley  
Pincus, Gregory  
Plass, E. D.  
Platner, Wesley S.  
Plaut, G. W. E.  
Plum, Claus Munk  
Pomerat, C. M.  
Ponder, Eric  
Power, Marschelle H.

Quastel, J. H.  
Quick, Armand J.

Ragsdale, Arthur C.  
Rahn, Hermann  
Rahn, Otto  
Rapoport, Samuel  
Rasmussen, A. F., Jr.  
Rasmussen, K.  
Rebuck, John W.

Reich, Carl  
Reisner, Edward H.  
Rekers, Paul E.  
Richter, Kenneth M.  
Riddle, Oscar  
Riley, Richard L.  
Riser, William H., Jr.  
Ritchey, Mary G.  
Roberts, Sidney  
Robson, J. M.  
Roofe, Paul G.  
Root, Raymond W.  
Rosahn, Paul D.  
Rosenfield, Richard E.  
Rosner, Lawrence  
Ross, Joseph F.  
Ross, Morris H.  
Rossiter, R. J.  
Rundles, R. Wayne  
Russell, Jane Anne

Sabine, Jean Captain  
Salter, William T.  
Samter, Max  
Samuels, Leo T.  
Saubertlich, Howerde E.  
Saunders, Francis J.  
Sawitsky, A.  
Sawyer, Charles H.  
Sayers, George  
Scatchard, George  
Schrek, Robert  
Schwarte, Louis H.  
Schweigert, B. S.  
Seegers, Walter H.  
Seibert, Florence B.  
Seligman, Arnold M.  
Selle, W. A.  
Selye, Hans  
Sendroy, Julius, Jr.  
Shemin, David  
Sherwin, Carl P.  
Shettlar, Marvin R.  
Shorb, Mary S.  
Shukers, Carroll F.  
Sibley, John A.  
Silver, S. D.  
Simonsen, Daisy G.  
Singer, Richard B.  
Sizer, Irwin W.  
Smith, Carl H.  
Smith, Christianna  
Smith, Clement A.  
Smith, Emil L.  
Smith, H. P.  
Smith, Kenneth  
Smith, Olive Watkins  
Smith, Paul K.  
Smyth, Henry F., Jr.  
Snyder, Laurence H.  
Somers, G. Fred  
Spangler, S.  
Sperry, Warren M.  
Spicer, Samuel S.  
Stadie, William C.  
Stahmann, Mark A.



Stasney, Joseph  
Stearns, Genevieve  
Steele, Betty F.  
Stefanini, Mario  
Stekol, Jakob A.  
Stone, Robert S.  
Stowell, Robert E.  
Strauss, Maurice B.  
Strong, L. E.  
Strumia, Max. M  
Sturgeon, Philip  
Sundberg, R. Dorothy  
Swann, H. G.  
Sydenstricker, Virgil P.  
Sykes, J. F.  
Szego, Clara M.

Taffel, Max  
Tagnon, Henry  
Talbot, Nathan B.  
Talbot, John H.  
Tatum, Edward L.  
Tauber, Henry  
Taylor, F. H. Laskey  
Taylor, Henry Longstreet  
Thannhauser, S. J.  
Thewlis, Ethel  
Thieme, Frederick  
Tocantins, Leandro M.  
Tompkins, Edna H.  
Tuft, H. S.  
Turner, Abby H.  
Turner, Charles W.  
Tyslow, R.

Ulrich, Albert  
Unger, Lester J.  
  
Valentine, William N.  
Vallee, Bert L.  
Van Liere, Edward J.  
Van Slyke, Donald Dexter  
van Wagtendonk, W. J.  
Vaughan, Stuart L.  
Venning, Eleanor H.  
Vestling, Carl S.  
Villela, Gilberto G.  
Vogt, Marthe  
von Euler, U. S.

Waelsch, Heinrich B.  
Wagner, Richard  
Waisman, Harry A.  
Walford, Lionel A.  
Walker, Burnham S.  
Wallace, William M.  
Walters, Orville S.  
Wang, Chi Che  
Ware, Arnold G.  
Warner, E. D.  
Warren, Shields  
Warren, Stafford L.  
Washburn, Alfred H.  
Watkins, C. H.  
Watson, C. J.  
Watson, R. Janet  
Waugh, Theodore R.  
Weil, Leopold  
Wells, Benjamin B.

West, G. B.  
Whitby, Sir Lionel E. H  
White, Abraham  
White, L. B.  
White, Paul Dudley  
Whitehead, William H.  
Whiting, P. W.  
Wicks, Lester F.  
Wiener, Alexander S.  
Williams, Harold H.  
Williams, J. N., Jr.  
Wilson, Sloan J.  
Windle, William F.  
Wintrobe, Maxwell M.  
Wirtschaftler, Zolton T.  
Wislocki, George B.  
Witebsky, Ernest  
Wojcik, L. D.  
Wolf, Stewart  
Wolff, James Alexander  
Wolff, René  
Wood, Earl H.  
Wood, John L.  
Woodard, Helen Quincey  
Wright, Claude-Starr

Young, I. Maureen

Zarrow, M. X.  
Ziegler, Mildred R.  
Ziffren, Sidney E.  
Zimmerman, Henry A.  
Zirkle, Raymond E.  
Zwemer, Raymond L.

# Contents

<u>Table</u>	<u>Subject</u>	<u>Page</u>
	Introduction	1
	PHYSICAL PROPERTIES	
1	Blood Specific Gravity	5
2	Relative Viscosities, Blood Plasma, Serum	6
3	Blood Miscellaneous Physical Properties	7
4	Blood Colloids, Osmotic Pressure	8
5	Erythrocyte Sedimentation Rate, Time	9
6	Erythrocyte Fragility Man	10
7	Erythrocyte Fragility Vertebrates Other Than Man	10
	COAGULATION PHENOMENA	
8	Blood Coagulation Theory of P. A. Owren (1951)	11
9	Blood Coagulation Theory of A. J. Quick (1951)	12
10	Blood Coagulation Theory of W. H. Seegers (1951)	13
11	Blood Coagulation Theory of M. Stefanni (1951)	14
12	Blood Coagulation Theory of L. M. Tocantins (1951)	15
13	Blood Coagulation Time Man	16
14	Heparin-Retarded Blood Coagulation Time Man	17
15	Heparin Tolerance Man	17
16	Skin Bleeding Time Man	18
17	Blood Clot Retraction Volume, Time Man	19
18	Plasma Prothrombin Time Man	20
19	Plasma Prothrombin, Relative Levels Vertebrates	21
20	Plasma Accelerator Globulin, Relative Levels Vertebrates	21
21	Blood Platelet Count Man	22
22	Blood Platelet Count Laboratory, Farm Animals	23
23	Capillary Fragility Man	24
	BLOOD GROUPS	
24	The ABO System of Blood Factors and Their Frequencies	25
25	The Distribution of The ABO Blood Groups in Various Populations	26
26	Heredity of The ABO Blood Groups	27
27	Heredity of The ABO Subgroups	28
28	The Rh-Hr (CDE-cde) System of Blood Factors and Their Frequencies	29
29	The Distribution of the Rh-Hr Blood Types in Various Populations	30
30	The MNS System of Blood Factors and Their Frequencies	31
31	The Distribution of the MN Blood Factors in Various Populations	32
32	Heredity of the MN Blood Factors	32
33	The Kell-Cellano System of Blood Factors and Their Frequencies	33
34	The Lewis System of Blood Factors and Their Frequencies	34
35	The Lutheran System of Blood Factors and Their Frequencies	35
36	The P System of Blood Factors and Their Frequencies	36
	ERYTHROCYTES AND HEMOGLOBIN	
37	Blood Erythrocyte Values Man	37
38	Blood Erythrocyte, Hematocrit, Hemoglobin Platelet Values Man	38
39	Blood Erythrocyte and Hemoglobin Values Sea Level Man	39
40	Blood Erythrocyte and Hemoglobin Values at Altitude Man	40
41	Blood Erythrocyte Values in Pregnancy and Postpartum Man	41
42	Blood Erythrocyte Values Mammals Laboratory	42
43	Blood Hematocrit and Hemoglobin Values Vertebrates	43
44	Blood Erythrocyte Values Fish	44
45	Erythrocyte Diameters Mammals	45
46	Blood Erythrocyte Values, Fetus Newborn, Adult Female Mammals	46
47	Erythrocyte Oxygen Consumption	48
48	Erythrocyte Carbohydrate Metabolism	49
	LEUKOCYTES	
49	" " " " " " " " " "	50
50	" " " " " " " " " "	52
51	" " " " " " " " " "	52
52	" " " " " " " " " "	53

53	Blood Leukocyte Ranges - Farm Animals	54
54	Morphology of Blood Leukocytes and Platelets - Man	55
55	Properties and Functions of White Blood Cells and Related Forms	57
<b>BLOOD AND BONE MARROW CELLS</b>		
56	Genealogy of the Formed Elements of Blood, Theory of William Bloom	67
57	Genealogy of the Formed Elements of Blood, Downey, via Oliver P. Jones	68
58	Nomenclature Recommended For Cells of Blood, Charles A. Doan	69
59	Bone Marrow (Sternal) Differential Cell Count, Summary - Man	70
60	Bone Marrow (Sternal) Differential Cell Count in Pregnancy - Man	71
61	Bone Marrow (Sternal) Differential Cell Count: Dog	72
62	Bone Marrow (Rib) Differential Cell Count, Summary - Dog	73
63	Bone Marrow (Rib) Differential Cell Count, Summary - Man	74
64	Morphology of Formed Elements of Bone Marrow - Man	75
65	Histochemical Properties of Blood and Bone Marrow Cells - Man	76
66		86
<b>WATER, CARBOHYDRATE, LIPID, PROTEIN</b>		
67	Blood Water and Solids	87
68	Blood Carbohydrates and Related Substances	88
69	Blood Lipids	92
70	Blood Proteins - Man	94
71	Plasma Proteins - Animals Other Than Man	95
72	Plasma Proteins, Certain Properties and Reactions - Man	96
73	Plasma Proteins, Physical Properties - Man	97
74		98
<b>AMINO ACIDS, NON-PROTEIN NITROGEN</b>		
75	Blood Free Amino Acids	99
76	Blood Non-Protein Nitrogen Compounds	101
77	Nitrogen Content of Blood Nitrogen Compounds	102
<b>PHOSPHORUS, SULFUR</b>		
78	Blood Phosphorus, Inorganic	103
79	Blood Phosphorus, Organic Acid-Soluble	104
80	Blood Adenosine Triphosphate Phosphorus	105
81	Blood Diphosphoglycerate Phosphorus	106
82	Blood Nucleotide Phosphorus	107
83	Blood Lipid Phosphorus	107
84	Blood Hexose Phosphate Phosphorus	108
85	Blood Phytic Acid Phosphorus	108
86	Blood Sulfur	
87	Blood Sulfur Compounds	
<b>VITAMINS, HORMONES, ENZYMES</b>		
88	Blood Vitamins	109
89	Blood Hormones	112
90	Blood Enzyme Activity	113
91	Blood Coenzymes	116
<b>ELECTROLYTES, MINERALS, GASES, ACID-BASE</b>		
92	Blood Electrolytes	117
93	Blood Minor Minerals	119
94	Arterial Blood Gases, $O_2$ and $CO_2$ , at Altitude - Man	120
95	Ionic Balance and Buffer Base, Arterial Blood, Cells - Man	121
96	Acid-Base Values, Blood and Plasma - Man	123
97		124
<b>EFFECTS OF RADIATION, STORAGE</b>		
98	Effects of Radiation on Peripheral Blood	125
99	Effects of Radiation on Hematopoietic Tissues	129
100	Changes in Stored Preserved Blood	136
<b>THERAPEUTIC AGENTS</b>		
101	Effective Blood Levels of Therapeutic Agents	138
	Radiography	145
	Index	193

# Introduction

The tables in this unusually complete collection of data on blood are unique in the high degree of reliability sought for the data. Data have been supplied and authenticated by over six hundred leading investigators in biology and clinical medicine. The tables have been exhaustively reviewed, some by as many as twenty experts in the field.

The tables are also unique in their treatment of the well known phenomenon of biological variability. In addition to giving a single representative value for each item covered in a table (the value is known in statistical terms as the estimated "universe mean") the tables present information on variability in a most easily comprehensible and directly usable form. The reader who seeks to know the range of variation of a quantity does not need to perform a calculation, as he must do when only the "standard deviation" is reported, but may read directly from the table the estimated upper and lower limits of the "95% range." The method of estimate, in each case, is identified by a simple code letter, which at the same time rates the individual range as to its trustworthiness. Range data as commonly encountered represent a mixture of variability as it exists between individuals and variability as it exists within individuals.

The significance of the code letters, used as superscripts attached to the ranges in the tables, follows:

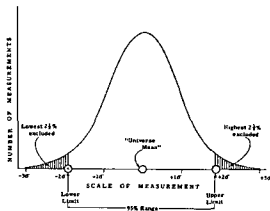
(a) By the method of greatest accuracy, the 95% range is obtained by fitting a recognized type of frequency curve to a group of measured values and excluding the extreme 2.5% of area under the curve at each end. (See sketch to the right.)

(b) By a less accurate method, the 95% range is estimated by a simple statistical calculation, assuming a "normal distribution" and using the "standard deviation." This estimate is used when the group of values is too small for curve fitting, as is usually the case.

(c) A third and still less accurate procedure for estimate of the 95% range is simply to give the highest value and lowest value of the reported sample group of measurements. It underestimates the 95% range for small samples and overestimates for larger sample sizes, but may be used in preference to the preceding method when the sample shows convincing evidence that the variable is asymmetrical in distribution.

(d) The upper and lower limits of the range of variation, as commonly encountered by an investigator experienced in measuring the quantity in question, constitute still another estimate of the 95% range. The trustworthiness of limits so placed should not be underestimated.

In many instances, where information as to the manner of estimate has been lacking at the time of going to press, it has been necessary to report an estimate of the 95% range without an identifying superscript. Effort to assemble the missing information is continuing.



Although the data in each table are the best available at the time the table was prepared, it is recognized that all data are subject to revision as investigators improve techniques and make more measurements. The reader is warned against attributing significance to small differences from species to species. He is invited to submit any values or ranges that he feels should be given consideration, and is particularly invited to add to the coverage of animal forms.



# TABLES



# 1. BLOOD SPECIFIC GRAVITY

Blood (B), RBC (C), Plasma (P)

Animal		Value		Range	Temperature (°C)	Method
(A)		(B)		(C)	(D)	(E)
1	Man	B	♂ 1.056	1.052-1.061 <sup>b</sup>	25/4	Copper Sulfate
2		C	♂ 1.093 <sup>2</sup>	1.089-1.097 <sup>b</sup>		
3		P	♂ 1.024	1.022-1.026 <sup>b</sup>		
4		C	1.099	1.094-1.107 <sup>c</sup>		Gravimetric
5		P	1.028	1.026-1.031 <sup>b</sup>		
6	Cat	B	♂ 1.050	1.046-1.054 <sup>b</sup>	T/T	Falling Drop
7		B	♀ 1.051	1.045-1.057 <sup>b</sup>		
8	Cattle	B	♂ 1.052	1.046-1.058 <sup>b</sup>	20/4	Gravimetric
9		B	♀ 1.053	1.046-1.061 <sup>b</sup>		
10		C	♂ 1.084	1.079-1.090 <sup>b</sup>		Gravimetric
11		P	♂ 1.029	1.026-1.033 <sup>b</sup>		
12	Dog	B	♂ 1.052		22-26/4	Falling Drop
13		B	1.045			
14	Goat <sup>8</sup>	B	♂ 1.042	1.036-1.048 <sup>b</sup>	25/4	Copper Sulfate
15		B	♀ 1.044	1.036-1.051 <sup>b</sup>		
16		B	♂♀ 1.042	1.035-1.049 <sup>b</sup>		
17		P	♂ 1.023	1.019-1.026 <sup>b</sup>		
18		P	♀ 1.021	1.018-1.024 <sup>b</sup>		
19		P	♂♀ 1.022	1.019-1.025 <sup>b</sup>		
20	Horse	B	♂♀ 1.053	1.046-1.059 <sup>c</sup>	20/4	Benzene-Chloroform
21	Mouse <sup>10</sup>	B	1.057	1.052-1.062 <sup>b</sup>	25/4	Falling Drop
22	Pig, young <sup>11</sup>	B	♂ 1.047	1.038-1.055 <sup>b</sup>	25/4	Copper Sulfate
23		B	♀ 1.043	1.035-1.052 <sup>b</sup>		
24		B	♂♀ 1.046	1.039-1.054 <sup>b</sup>		
25		P	♂ 1.022	1.021-1.025 <sup>b</sup>		
26		P	♀ 1.023	1.020-1.027 <sup>b</sup>		
27		P	♂♀ 1.022	1.019-1.025 <sup>b</sup>		
28	Rabbit	B	1.050	1.048-1.052 <sup>b</sup>	25/4	Falling Drop
29		C	1.098	1.093-1.104 <sup>c</sup>		Gravimetric
30		P	1.025	1.018-1.031 <sup>b</sup>		
31	Rat <sup>13</sup>	B	1.056 <sup>12</sup>	1.054-1.058 <sup>b</sup>	25/4	Falling Drop
32		B	♂ 1.054	1.046-1.061 <sup>b</sup>	25/4	Copper Sulfate
33		B	♀ 1.054	1.046-1.061 <sup>b</sup>		
34		P	♂ 1.023	1.017-1.028 <sup>b</sup>		
35		P	♀ 1.022	1.018-1.027 <sup>b</sup>		
36		P	♂♀ 1.023	1.018-1.028 <sup>b</sup>		
37	Sheep	B	♂♀ 1.051 <sup>14</sup>	1.041-1.061 <sup>b</sup>	20/4	Benzene-Chloroform
38		C	1.084	1.080-1.087 <sup>c</sup>		Gravimetric
39		P	1.028	1.025-1.029 <sup>c</sup>		

/1/ Referred to water at 4°C or to water at temperature of measurement (T). /2/ Of "packed" cells, not corrected for an estimated 7% of trapped plasma, corrected = 1.098 (1.095-1.101).

/3/ Mostly Holstein-Friesian strain /4/ 2.5-18 yrs /5/ 18-30 months. /6/ Mongrels.

/7/ 1-3 days. /8/ Angora and Toggenburg strains, 1-2 yrs /9/ Castrated. /10/ CBA strain, 60-80 days. /11/ Duroc-Jersey, Poland-China, Chester White strains. /12/ Sprague-Dawley, 200-250g /13/ Mixed strains, 100-300g. /14/ Cheviot, blackface, grayface strains.



## 2. RELATIVE VISCOSITIES, BLOOD, PLASMA, SERUM

	Animal	Temperature of Meas. (°C)	Relative Viscosity <sup>1</sup>		
			Blood <sup>2</sup>	Plasma <sup>3</sup>	Serum <sup>3</sup>
	(A)	(B)	(C)	(D)	(E)
1	Man	38	4.7	1.8	1.5
2	Cat	38	4.2		
3	Dog	38	4.7		
4		37	5.5		
5	Goat	20	4.0		
6	Horse	20	4.1	1.9	1.7
7	Ox	20	4.6		
8	Pig	20	5.9		1.6
9	Rabbit	37	3.4		1.4
10	Sheep	20	4.3	1.6	1.5
11	Frog	15	2.8 <sup>4</sup>		1.5
12	Turtle	20	2.2 <sup>4</sup>		

/1/ Relative to water at the temperature of measurement. Absolute viscosity = relative viscosity x absolute viscosity of water. Absolute viscosity of water in poises (=dyne-seconds/cm<sup>2</sup>), accurate to within 2%, is 0.00680 at 38°C, 0.00692 at 37°C, 0.0101 at 20°C, and 0.0114 at 15°C. /2/ Apparent viscosity as measured in capillary tube viscosimeters with radii greater than 0.05cm, length greater than 150 radii, and at apparent Reynolds numbers in the range 50-800 (apparent Reynolds number =  $ur\rho/v'$  where  $u$  = mean velocity,  $r$  = tube radius,  $\rho$  = density, and  $v'$  = apparent viscosity, all expressed in c. g. s. units). Apparent viscosity of blood varies inversely with the temperature of measurement, directly with the protein content, and with the hematocrit. Hematocrits for above values not available in original reports. Hematocrit differences may be the chief reason for variations among species and between the sexes. /3/ Calculated from protein concentration in g/100ml plasma or serum (=c) and fluidity lowering constant (=k) where apparent viscosity =  $100/(100-kc)$ ;  $k$  = 5.8 for coordinates 1D, 4.8 for 1E, 7.0 for 6D, 6.1 for 6E, 5.7 for 8E, 5.3 for 9E, 6.5 for 10D, 5.8 for 10E. For values of c, see table on Plasma Proteins, Laboratory and Farm Animals. /4/ Corrected for defibrination (=10% less viscous than whole blood).

### 3. BLOOD MISCELLANEOUS PHYSICAL PROPERTIES

RBC (C); Plasma (P); Serum (S)

Variable and Animal		Value	Range	Variable and Animal		Value	Range
(A)		(B)	(C)	(A)		(B)	(C)
1 2 3 4 5 6 7 8	Electrical charge (millivolts)	C -16.8		19	Man	-0.562	-0.555 to -0.570 <sup>c</sup>
		C -17.8		20	Dog	-0.571	
		C -21.1		21	Horse	-0.564	
		C -14.2		22	Ox	-0.585	
		C -17.0		23	Pig	-0.615	
		C -17.9		24	Rabbit	-0.592	
		C -7.0		25	Sheep	-0.619	
		C -18.6		26	Fish, pike	-0.514	
9 10 11 12 13 14 15 16	Electrophoretic mobility <sup>1</sup> (cm <sup>2</sup> /volt sec. x 10 <sup>-4</sup> )	C 1.31		27	Fish, plaice	-0.732	
		C 1.39		28	Fish, tench	-0.523	
		C 1.65		Refractive index			
		C 1.11		29	Man <sup>2</sup>	1.34215	1.34920-1.35110 <sup>c</sup>
		C 1.33		30	Fish, cod <sup>3</sup>	1.34165	1.34182-1.34249 <sup>c</sup>
		C 1.40		31	Fish, plaice <sup>3</sup>	1.34052-1.34237 <sup>c</sup>	
		C 0.55		32	Fish, tench <sup>3</sup>	1.34089	1.34082-1.34094 <sup>c</sup>
		C 1.45		Specific heat (gram calories)			
17 18	Electrical conductivity (Mhos x 10,000)	S 120	117-124 <sup>c</sup>	33	Man	P 0.94	
		S 106	105-111 <sup>c</sup>	34	Man	C 0.77	
				35	Man	P+Q 0.87	

/1/ M/15 phosphate buffer at pH 7.4. /2/ At 17.5°C with Abbe refractometer, refractive index of water being 1.3320. /3/ At 17.5°C with Zeiss Pulfrich refractometer calibrated with distilled water and standard salt solutions at the same temperature.

# 4. BLOOD<sup>1</sup> COLLOIDS, OSMOTIC PRESSURE

Class		Animal	mm of water <sup>2</sup>	
			Value	Range
(A)		(B)	(C)	(D)
1	Amphibia	Frog ( <i>Rana catesbiana</i> )	103	96-115°C
2		Frog ( <i>Rana temporaria</i> )	70	-140°C
3		Toad ( <i>Bufo vulgaris</i> )	133	83-242°C
4	Aves	Hen	150	140-160°C
5		Dove	110	80-120°C
6	Elasmobranchs	Dogfish, lesser spotted ( <i>Scyliorhinus caniculus</i> ) <sup>3</sup>		31-36°C
7		Dogfish, smooth ( <i>Mustelus mustelus</i> ) <sup>3</sup>		57-64°C
8		Dogfish, spiny ( <i>Squalus acanthias</i> ) <sup>3</sup>		42-43°C
9		Ray, electric ( <i>Torpedo marmorata</i> )		42-52°C
10	Mammalia	Man	330	280-480°C
11		Cat	300	240-330°C
12		Dog	310	230-470°C
13		Goat	300	300-310°C
14		Guinea pig	250	230-280°C
15		Horse	280	230-350°C
16		Ox	280	260-300°C
17		Pig	330	300-350°C
18		Rabbit	290	230-350°C
19		Rat	260	220-290°C
20		Sheep	300	290-340°C
21				
22	Pisces	Fresh Water		
23		Carp, common ( <i>Cyprinus carpio</i> )	225	100-113°C
24		Eel ( <i>Anguilla anguilla</i> ) <sup>3</sup>	101	112-146°C
25		Pike ( <i>Esox lucius</i> )		93-109°C
26		Tench ( <i>Tinca vulgaris</i> )		174-250°C
27		Sea Water		112-114°C
28		Bass ( <i>Labrax lupus</i> )	113	146-173°C
29		Cod ( <i>Gadus morrhua</i> )		195-213°C
30		Eel, conger ( <i>Conger conger</i> ) <sup>3</sup>		196-198°C
31		Gurnard ( <i>Trigla lucerna</i> )	115	107-126°C
32		Mackerel ( <i>Scomber scombrus</i> )		181-186°C
33		Plaice ( <i>Pleuronectes platessa</i> )	174	
34	Reptilia	Scorpion fish ( <i>Scorpaena scrofa</i> )		
35		Turbot ( <i>Rhombus maximus</i> )		
		Turtle ( <i>Malacoclemmys</i> )	spring <sup>4</sup> fall <sup>5</sup>	58 96
				48-68°C 81-112°C

/1/ Serum. Variations in osmotic pressure due to differences in temperature of measurement (in the range of 0°-37°C) are generally less than those due to error of measurement. /2/ mm of water/13.546 = mm of Hg. /3/ Original report in literature for 7B = *Scyllium canicula*, for 8B = *Mustelus hinmulus*, for 9B = *Acanthias vulgaris*, for 23B = *Anguilla vulgaris*, and for 28B = *Conger vulgaris*. /4/ Fasting. /5/ Fed.

# 5. ERYTHROCYTE SEDIMENTATION RATE, TIME<sup>1</sup>

Animal	mm/hr		Method of:		Animal	mm/hr		Method of:
	Value <sup>2</sup>	Range <sup>2</sup>	(B)	(C)		Value <sup>2</sup>	Range <sup>2</sup>	
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
Man					23	Goat	0.50	Reichel
1		♂ 0-8			24	Guinea pig	1.67	Reichel
2		♀ 0-10	Cutler		25		1.06	Westergren
3		♂ 0-6	Landau		26		1.5	Linzenmeier-Raunert
4		♀ 0-9			27		18	Cutler
5		♂ 0-10	Smith		28	Horse	127	Reichel
6		♀ 0-10			29		69	Linzenmeier-Raunert
7		♂ 0-8	Walton		30	Pig	5.35	Reichel
8		♀ 0-8			31		5	Linzenmeier-Raunert
9		♂ 0-15	Westergren		32	Rabbit	1.05	Wintrobe
10		♀ 0-20			33		2	Reichel
11		♂ 0-9	Wintrobe		34		2	Linzenmeier-Raunert
12		♀ 0-15			35	Rat	♂ 0.7	Cutler
13	8.6	4-13	Cutler		36		♀ 1.8	
14	9.13	3-13	Smith		37	Sheep	0.55	Reichel
15	7.30	4-13	Reichel		38		0.5	Linzenmeier-Raunert
16			Linzenmeier-Raunert		39	Fowl	3.75	Reichel
17	1.17	1-1.8	Reichel		40		4	Linzenmeier-Raunert
18	1		Linzenmeier-Raunert		41	Goose	3.42	Reichel
19		♂ 1-4	Wintrobe		42	Man	6-10 hrs. <sup>6</sup>	
20		♀ 1-52			43	Guinea pig	60 hrs. <sup>6</sup>	Linzenmeier
21	2		Linzenmeier-Raunert		44	Rabbit	17-42 hrs. <sup>6</sup>	
22	4	2, 5-5	Reichel					

/1/ Increased during infections, diseases associated with inflammation and tissue injury, with hemodilution; decreased with hemoconcentration. /2/ Measured at the end of the first hour; rate is usually unchanged within the first hour; rates in succeeding hours decrease asymptotically. /3/ 12 days-14 yrs; capillary blood. /4/ 11-56 days pregnant. /5/ No change after thyroidectomy. /6/ Sedimentation time.

## 6. ERYTHROCYTE FRAGILITY: MAN

Venous Blood (Bv), Oxygenated Blood (Bo), Cutaneous Blood (Bc)

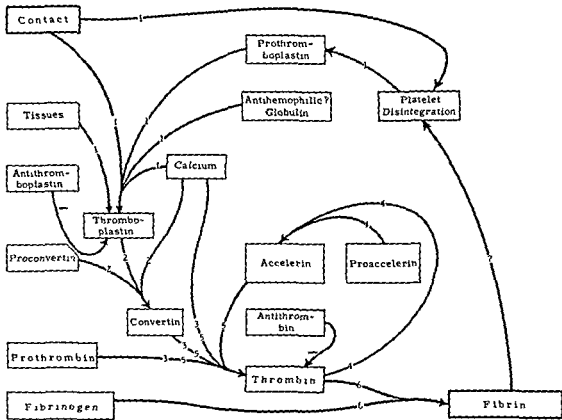
Anticoagulant	Temperature (°C)	Dilution of Blood <sup>1</sup>	Duration of Hemolysis	% NaCl Solution (g NaCl/100 ml)			Method of Determining Degree of Hemolysis
				for 10% Hemolysis	for 50% Hemolysis	for 90% Hemolysis	
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
1 Waxed tube		1 in 26	20 min	Bo 0.43 <sup>2</sup>	0.38 <sup>2</sup>	0.34 <sup>2</sup>	Hemoglobin liberated
2 No anticoagulant		1 in 200	30 min	Bc 0.47 <sup>2</sup>	0.43 <sup>2</sup>	0.40 <sup>2</sup>	RBC count
3 Oxalate	Rm temp	1 in 201	30 min	Bv 0.46 <sup>2</sup>	0.41 <sup>2</sup>	0.37 <sup>2</sup>	Hemoglobin liberated
4 Oxalate	37	1 in 41 <sup>3</sup>	30 min	Bv 0.46 <sup>2</sup>		0.22 <sup>2</sup>	Hemoglobin liberated

/1/ Volume of blood in volume of total mixture of blood and hemolyzing solution. /2/ Estimated from graphs. /3/ Washed cells. /4/ Hemolysis "starts" at. /5/ Hemolysis "complete" at.

## 7. ERYTHROCYTE FRAGILITY: VERTEBRATES OTHER THAN MAN

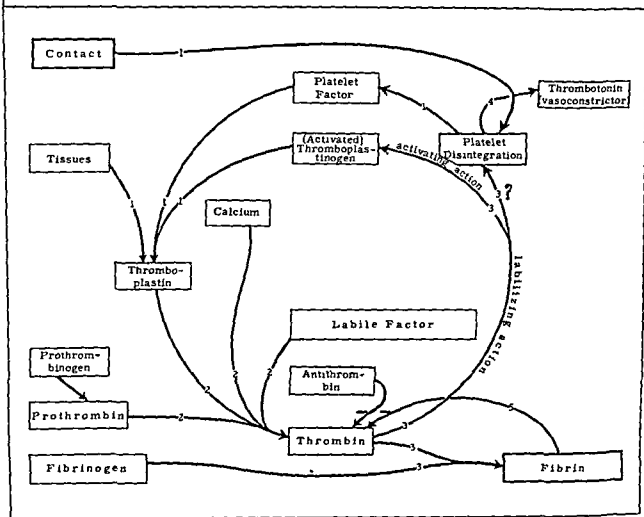
Animal	%NaCl Solution (g/100ml)		Animal	%NaCl Solution (g/100ml)	
	for Initial Hemolysis	for Complete Hemolysis		for Initial Hemolysis	for Complete Hemolysis
(A)	(B)	(C)	(A)	(B)	(C)
1 Baboon		0.18	12 Llama	0.17	0.03
2 Camel		0.26	13 Marmoset		0.40
3 Cat	0.69	0.50	14 Mouse	0.54	0.33
4 Kitten	0.58	0.47	15 Ox	0.59	0.42
5 Chimpanzee		0.28	16 Pig	0.74	0.45
6 Dog	0.45	0.36	17 Rabbit	0.57	0.45
7 Dromedary	0.17	0.07	18 Rat	0.48	0.38
8 Goat	0.62	0.48	19 Sheep	0.60	0.45
9 Guinea pig	0.45	0.33	20 Lamb	0.69	0.48
10 Horse	0.59	0.39	21 Chicken	♂ 0.40	0.32
11 Lemur		0.42	22	♀ 0.41	0.28

### 8. BLOOD COAGULATION: Theory of P.A. Owren (1952)



[1] Tissue injury yields thromboplastin directly, while contact causes disintegration of platelets and release of prothromboplastin, which is activated by contact. "antihemophilic globulin" and calcium to give thromboplastin. [2] Thromboplastin and proconvertin in the presence of calcium form convertin. An anticonvertin probably exists which opposes the activity of convertin. [3] Convertin together with calcium brings about a minimal conversion of prothrombin to thrombin. [4] This initially formed thrombin starts the accelerator system, i.e., the conversion of proaccelerin to accelerin. [5] Accelerin accelerates the conversion of prothrombin to thrombin in the presence of convertin and calcium. [6] Thrombin is now in sufficient quantity to convert fibrinogen to fibrin. [7] Fibrin provokes the disintegration of the platelets with further release of thromboplastic substances already mentioned.

## 9. BLOOD COAGULATION: Theory of A.J. Quick (1952)

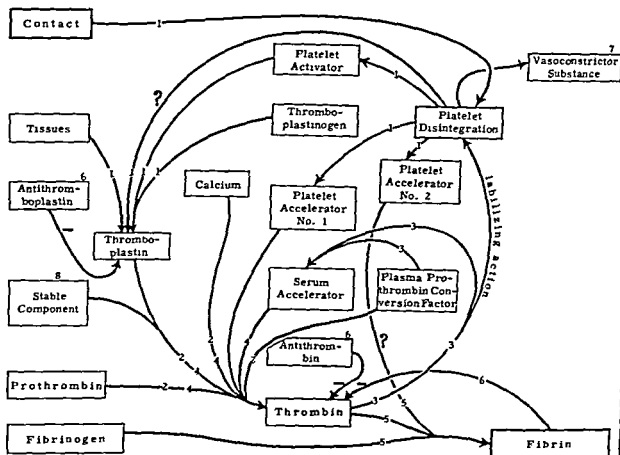


/1/ Thromboplastin is (a) released directly by injured tissue and is also (b) formed by the interaction of plasma thromboplastinogen and a platelet factor, the latter released from disintegrating platelets. /2/ Thromboplastin, prothrombin, calcium, and a labile factor, interact stoichiometrically to form thrombin. In human blood part of the prothrombin is in an inactive form, prothrombinogen. On contact with a rough surface it becomes activated. /3/ The thrombin formed not only converts fibrinogen to fibrin, but activates thromboplastinogen and thereby probably brings about the lysis of platelets and initiates the chain reaction. /4/ The platelets also release a vasoconstrictor, thrombotonin, under the labilizing action of thrombin. The resulting local vasoconstriction aids in the hemostatic process. /5/ The prompt removal of thrombin by fibrin holds in check the autocatalytic reaction mediated through the action of thrombin on platelets and thromboplastinogen.



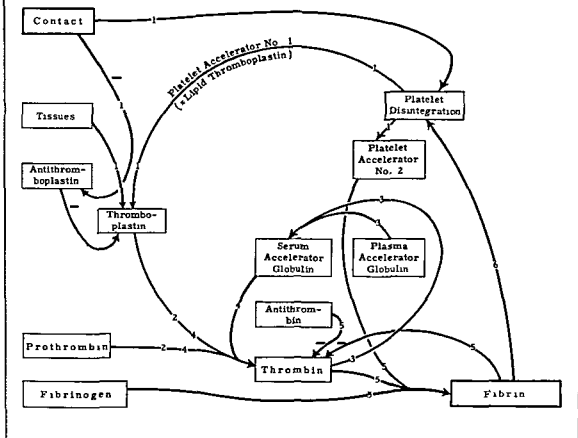


# 11. BLOOD COAGULATION: Theory of M. Stefanini (1952)



/1/ Injured tissue yields thromboplastin directly, while platelet disintegration yields, (a) platelet activator, which interacts with plasma thromboplastinogen (and possibly another plasma component) to form thromboplastin, (b) possibly, thromboplastin directly and (c) two accelerators principles, Nos. 1 and 2. /2/ Thromboplastin, calcium, plasma prothrombin conversion factor and stable component interact to bring about a minimal conversion of prothrombin to thrombin. /3/ This initially formed thrombin initiates the autocatalytic phase of the process, i. e., (a) starts the accelerator system--the conversion of plasma prothrombin conversion factor into a more active form, i. e., serum accelerator, and (b) "labilizes" the platelets to release more platelet activator and accelerators. /4/ Thromboplastin and calcium, together with serum accelerator, platelet accelerator No. 1 and, possibly, stable component cause accelerated conversion of prothrombin to thrombin. /5/ A now sufficient quantity of thrombin, aided by the action of platelet accelerator No. 2, converts fibrinogen to fibrin. /6/ Antithromboplastin and antithrombin act to decrease the speed of coagulation reactions in which they are involved. The fibrin clot, by adsorbing thrombin, also effectively checks the autocatalytic mechanism of blood coagulation. /7/ The platelet disintegration which takes place at the site of vascular injury also liberates a vasoconstrictor agent which aids in the arrest of the hemorrhage. /8/ Stable component is a factor which appears decreased during dicumarol therapy and is sharply decreased in the hypoprothrombinemia of the newborn. Its properties are very similar to those of prothrombin.

## 12. BLOOD COAGULATION: Theory of L. M. Tocantins (1952)



/1/ Contact of the blood with certain surfaces (damaged blood vessel endothelium, glass) initiates the first changes

15. the conversion of inactive plasma accelerator globulin to active serum accelerator globulin. /4/ Thromboplastin together with serum accelerator globulin causes acceleration of the conversion of prothrombin to thrombin. /5/ Some of the thrombin may be inactivated by antithrombin. The thrombin that escapes such inactivation acts, with the aid of platelet accelerator No. 2, to cause the conversion of fibrinogen to fibrin. Some of the excess thrombin is removed from the plasma by adsorption on fibrin. /6/ Fibrin probably causes further disintegration of platelets.

# 13. BLOOD COAGULATION TIME: MAN

Venous Blood (Bv); Cutaneous Blood (Bc); Venous Plasma (Pv)

Method	Tempera- ture (°C)	Clotting Time <sup>1</sup> Minutes			Method of:
		Value		Range	
		(C)	(D)		
(A)	(B)				(E)
1 ml blood in tube	Rm. Temp.	Bv	6.5	5-8	Lee and White
		Bc	3.0	2-4	
		37	Bv	11.5	7-15 <sup>b</sup>
2 ml blood in each of 2 tubes	Rm. Temp.	Bv	8.9	3.5-14.3 <sup>b</sup>	Aggeler and Lucia
	37	Bv	7.5	4.7-10.4 <sup>b</sup>	
	25	Bv		10.5-19.5	Lewis
	29	Bv		6.5-14.5	
	37	Bv		5.5-12.5	
	45	Bv		5-9.5	
1 ml blood in each of 4 tubes	Rm. Temp.	Bv	11.0	5-20	Copley and Houlhan
1 ml blood in paraffined tube	37	Bv	29.3	16-43 <sup>b</sup>	Tocantins
1 ml blood in siliconed tube		Bv		22-61	Lewis
2 ml blood in each of 2 lusteroid tubes	37	Bv		25-50	Diggs
Bead of blood on platinum wire	37	Bc		1.5-2	Gibbs
Blood in 5 fine capillary tubes <sup>4</sup>	37	Bc		3-7	
0.1 ml oxalated plasma + CaCl <sub>2</sub> in each of 2 tubes	37	Pv		1.5-2.3 <sup>2,3</sup>	Quick
0.2 ml oxalated plasma + CaCl <sub>2</sub> in each of 2 tubes	23-26	Pv	5.3 <sup>2</sup>	3-8 <sup>c</sup>	Cheney
0.6 ml oxalated plasma + CaCl <sub>2</sub> in tube	37	Pv	2.7 <sup>2,5</sup>	2-3.5 <sup>c</sup>	Nygaard

/1/ Time varies with method; shortened by agitation, high temperature, uncoated glass, rapid vs slow centrifugation, etc. /2/ "Plasma recalcification time," "coagulation time of recalcified plasma."

/3/ Tube centrifuged at 1000 rpm should not clot more than 15 sec sooner than tube centrifuged at 3000 rpm. /4/ Tubes broken to read end point. /5/ Photoelectric end point reading.

# 14. HEPARIN-RETARDED BLOOD COAGULATION TIME: MAN

## Venous Blood (Bv)

Method		Temperature	Heparin added $\mu$ g	Clotting Time Minutes		Method of:
				Value	Range	
(A)		(B)	(C)	(D)	(E)	(F)
1	1 ml blood + heparin in tube	Rm. Temp.	1	Bv	8-16	de Takats
2			4	Bv 25	20-28 <sup>c</sup>	
3	1 ml blood + heparin in tube	Rm. Temp.	0.8 <sup>1</sup>	Bv 18	10-25	Waugh and Ruddick
4			1.5 <sup>1</sup>	Bv 28	19-38	
5			2.3 <sup>1</sup>	Bv 38	29-52	
6			3.1 <sup>1</sup>	Bv 48	35-60	
7			3.8 <sup>1</sup>	Bv 57	43-70	
8			4.6 <sup>1</sup>	Bv 64	52-83	
9			5.4 <sup>1</sup>	Bv 71	64-88	
10	1 ml blood + heparin in tube	Rm. Temp.	10	Bv	40-60	Tuft and Rosenfield

/1/ Originally reported in Toronto units. 1  $\mu$ g = 0.13 units.

# 15. HEPARIN TOLERANCE: MAN

## Venous Blood (Bv)

Method		Temperature ( $^{\circ}$ C)	Clotting Time Minutes		Method of:
			Value	Range	
(A)		(B)	(C)	(D)	(E)
1	10 min. after 10 mg heparin intravenously, 1 ml blood in 1 capillary tube <sup>2</sup>	37 <sup>1</sup>	Bv	5-7 <sup>d3</sup>	de Takats
2	10 min. after 25 mg heparin intravenously, 1 ml blood in 1 tube	37	Bv	40-90 <sup>3,4</sup>	Hagedorn and Barker

/1/ Held in palm of hand. /2/ Tube broken to read end point. /3/ Range excludes hypo- and hyper-reactors. /4/ Approx. 88% range.

16. SKIN BLEEDING TIME <sup>1</sup> : MAN						
Finger (F); Earlobe (E); Forearm, volar surface (Fv)						
Method	Wound Depth mm	Bleeding Time Seconds		Method of:		
		Value (C)	Range (D)			
(A)	(B)	(C)	(D)	(E)		
1 Puncture skin; blood absorbed	3	F;E	60-180	Duke		
2 every 15-30 seconds	3-4	Fv	147			
3 Puncture skin; immerse wound in physiological saline, 37°C	6	F	17-340 <sup>c,3</sup>	Copley and Lalich		
4 Sphygmomanometer cuff	2	Fv	192	Ivy		
5 maintained at 40 mm Hg	3-4	Fv	248			
6 pressure; skin puncture;	3	Fv	240 <sup>5</sup>			
7 blood absorbed every:	3	Fv	0-240 <sup>6</sup>			

/1/ Depends upon skin temperature, skin circulation, area punctured, thickness of skin, depth of puncture, psychic influences, etc. /2/ In 95% of persons, less than 374 seconds. /3/ In 95% of persons, less than 180 seconds. /4/ Frank bleeding should stop by 360 seconds; blood tinged oozing may last another 360 seconds. /5/ Average of two tests. /6/ Includes 99.54% of persons.

# 17. BLOOD CLOT RETRACTION VOLUME, TIME: MAN

Variable				Value	Range	Method of:
(A)				(B)	(C)	(D)
1 2 3 4 5	Clot Retraction	Volume %	Extracorporeal clot volume <sup>1</sup> as % of volume of blood specimen <sup>2</sup>	7.9	-4.1 <sup>3</sup> to 19.9 <sup>b</sup>	Aggeler and Lucia
			Expressed serum volume as % of volume of blood specimen <sup>4</sup>		40-55	Diggs
			Expressed serum volume as % of total serum present in blood specimen <sup>5</sup>	78.1	62-94 <sup>b</sup>	Tocantins
	Time Minutes		Venous blood in test tube at room temp.		30-60	Diggs
			Drop of blood in castor oil	33	20-45	Hirschboeck

/1/ = "Fluid volume of clot." /2/ The clot exclusive of RBC, WBC, and platelets, after clot retraction at 1 hr at 37°C. /3/ Negative values may occur due to discrepancies between methods of measurement of formed elements vs. total clot volume. /4/ After maximal clot retraction. /5/ Venous blood in paraffin tube, 13 mm diam., at 37°C, 2 hr. Amount of serum initially present calculated from the hematocrit.

# 18. PLASMA PROTHROMBIN TIME<sup>1</sup>: MAN

Venous Blood (Bv); Cutaneous Blood (Bc); Venous Plasma (Pv)

Method	Temperature (°C)	Thrombo- plastin added	Clotting Time Seconds		Method of:
			Value	Range	
(A)	(B)	(C)	(D)	(E)	(F)
1	37	Rabbit brain <sup>3</sup>	Pv	11-12	Quick
2	37	Rabbit brain <sup>4</sup>	Pv	17-19	Magath and Hurst
3	38.5	Russell viper <sup>5</sup> venom + lecithin	Pv	10 8-12 <sup>b</sup>	Witts and Hobson
4	37	Human brain <sup>3</sup>	Pv	11.5 10-13 <sup>b</sup>	Aggeler and Lucia
5	37	Rabbit brain <sup>3</sup>	Pv	24 19-29 <sup>b,8</sup>	Shapiro et al
6	Rm. Temp.	Russell viper <sup>5</sup> venom	Pv	23 16-30 <sup>c</sup>	Page and Russell
7	37	Rabbit brain <sup>3</sup>	Pv <sup>7</sup>	41 34-48 <sup>b,8</sup>	Shapiro et al
8	37	Human brain <sup>3</sup>	Pv <sup>7</sup>	28 21-35 <sup>b</sup>	Aggeler and Lucia
9	37	Human brain <sup>3</sup>	Pv <sup>7</sup>	16 13-19 <sup>b</sup>	
10	Rm. Temp.	Rabbit brain <sup>3</sup>	Bc	20 18-22 <sup>d</sup>	Kato and Poncher
11	Rm. Temp.	Beef lung or rabbit brain <sup>6</sup>	Bc	20-30	Karabin and Anderson
12	Rm. Temp.	Beef lung or rabbit brain <sup>3</sup>	Bv	25-50	Ziffren et al

/1/ Results of different methods are not comparable due to differences in concentration of reacting substances. Departures from normal can be adjudged as such only by comparison with normal controls run in the same laboratory. /2/ See also Table 19. /3/ Acetone dehydrated. /4/ Dried at 37°C. /5/ Daboia. /6/ Fresh. /7/ Oxalated plasma diluted with physiological saline. /8/ The authors draw conclusions from the difference between the values for diluted and undiluted plasma.

# 19. PLASMA PROTHROMBIN, RELATIVE LEVELS<sup>1,2</sup>: VERTEBRATES

Animal		Two-Stage Test <sup>3</sup> Dog = 100%	Animal		Two-Stage Test <sup>3</sup> Dog = 100%
(A)		(B)	(A)		(B)
1	Man	84 <sup>4</sup>	6	Rat	95
2	Cat	91	7	Chicken	50
3	Dog	100	8	Turtle	42
4	Guinea pig	53	9	Dogfish	8
5	Rabbit, albino	89	10	Sea bass	31
			11	Stingray	27

/1/ Data inadequate for stable estimate of universe mean. /2/ Dog plasma normally contains approximately 350 units of prothrombin/ml of plasma. A unit of prothrombin = amount required to form 1 unit of thrombin which is the quantity that will cause clotting of 1 ml of standard fibrinogen solution in 15 seconds, under standard conditions. /3/ Method of Warner, Brinkhous and Smith. Oxalated, and diluted venous plasma is defibrinated by precipitation with ammonium sulfate. The prothrombin is converted to thrombin by use of thromboplastins. The thrombin is assayed against a standard fibrinogen solution. Somewhat different relative values are obtained by the Quick one-stage method. /4/ Mean = 294 units/ml; range = 273-325 units/ml.

# 20. PLASMA ACCELERATOR GLOBULIN, RELATIVE LEVELS<sup>1,2</sup>: VERTEBRATES

Animal		One-Stage Method <sup>3</sup> Rabbit = 100%	Two-Stage Method <sup>4</sup> Dog = 100% <sup>5</sup>	Animal		One-Stage Method <sup>3</sup> Rabbit = 100%	Two-Stage Method <sup>4</sup> Dog = 100% <sup>5</sup>
(A)		(B)	(C)	(A)		(B)	(C)
1	Man	2	85	6	Rabbit, albino	100	94
2	Cat		80	7	Rat, albino		36
3	Cow	15	74	8	Chicken	5	2
4	Dog	20	100	9	Turtle		2
5	Guinea pig	10	21				

/1/ Data inadequate for stable estimate of universe mean. /2/ =Labile Factor, Proaccelerin (Factor V), or plasma prothrombin conversion factor. /3/ Method of Quick and Stefanini. /4/ Method of Ware and Seegers. /5/ =176 units/ml plasma. A unit of plasma Ac globulin=1000 times the amount that when present in 1 ml of a reacting mixture of prothrombin, thromboplastin, and calcium will reproduce a standard curve of thrombin production.



# 21. BLOOD PLATELET COUNT: MAN

Arterial blood (Ba); Venous blood (Bv); Cutaneous blood (Bc)

Platelet Count thousands/mm <sup>3</sup>				Method of:				
Value		Range						
(A)		(B)		(C)				
1	Ba	♂	350	Direct	Tocantins			
2	Bv	♂	310			150-690		
3	Bc	♂	250				180-358	
4	Bv	♂	294					204-395
5	Bv	♀	291		214-360			
6	Bc	♂	273			170-313		
7	Bc	♂♀	409				273-545 <sup>b</sup>	
8	Bv	♂	257					140-440 <sup>b</sup>
9	Bc		716	500-900 <sup>1</sup>	Dameshek			
10	Bc		234			130-350		
11	Bc		514				437-586	

/1/ 84% range.

# 22. BLOOD PLATELET COUNT: LABORATORY, FARM ANIMALS

Animal	Platelet Count thousands/mm <sup>3</sup> blood		Investigator
	Value	Range	
(A)	(B)	(C)	(D)
1 Cat	345	164-500	Field, 1930
2	422		Lawrence & Valentine, 1947
3	493	368-712	Arndt, 1925
4	519	356-760	Weiser, 1922
5 Cattle	684	542-975	Arndt, 1925
6 Calf	490		Mariconda, 1933
7	461 <sup>1,2</sup>	188-960 <sup>c</sup>	Tocantins, 1936
8 Dog	467 <sup>3</sup>	353-535	Aynaud, 1909
9	492	298-793	Arndt, 1925
10	638 <sup>4</sup>	584-856	Ledingham, 1915
11 Guinea pig	719 <sup>1</sup>	550-880	Watabiki, 1917
12	783 <sup>1,4</sup>	525-900	Tocantins, 1938
13 Horse	335	249-461	Weiser, 1922
14	352	254-560	Hikmet, 1927
15 Monkey <sup>1,6</sup>	267	155-424	Krumbhaar & Musser, 1920
16 Mouse	278	246-339	Copley & Robb, 1942
17	987		Jacobson, 1944
18 Pig	403	296-616	Hikmet, 1925
19 Rabbit	533 <sup>1</sup>	170-1,120 <sup>c,5</sup>	Casey & Rosahn, 1932
20	536 <sup>7</sup>	424-586	Aynaud, 1909
21	454 <sup>1</sup>	190-760	Yamamoto, 1933
22 Rat	477 <sup>1</sup>	232-641	Shechet et al., 1935
23	754	702-796	Olson, 1939
24	795	620-950	Ma, 1932
25	800	500-1,000	Creskoff et al., 1942
26	823		Machella & Higgins, 1939
27 Sheep	441	284-659	Ercegovac, 1936

/1/ Direct method. /2/ Venous blood (ear). /3/ Arterial blood (femoral).

/4/ Ear blood. /5/ Estimate "b" of 95% range = 288,000-788,000.

/6/ Finger blood. /7/ Arterial and venous blood.

### 23. CAPILLARY FRAGILITY<sup>1</sup>: MAN Suction Tests and Tourniquet Test

Forearm, outer surface (Fo); Forearm, antecubital space (Fa)

Age			Diameter Skin Area Observed		Pressure Duration	Petechiae first appear at negative pressure, mm Hg.		Method of:
			cm		Minutes	Value	Range	
(A)			(B)		(C)	(D)	(E)	(F)
1	Suction Test <sup>2</sup> (Decompression)	All ages (3-90 years)	Fo	1	1	350	100->500	Dalldorf
2		1 month	Fa	1	1	425 <sup>3</sup>		Lindquist
3		6 months	Fa	1	1	188 <sup>3</sup>		
4		1 year	Fa	1	1	156 <sup>3</sup>		
5		5 years	Fa	1	1	100 <sup>3</sup>		
6		10 years	Fa	1	1	96 <sup>3</sup>		
7		Adults	Fa	1	1	141	100-200 <sup>4</sup>	
8		Newborn	Fa	1	1		250-	Frontal
9		2-10 years	Fa	1	1		200-250	
10		22-36 years	Fa	1	1		150-200	

Applied Pressure mm Hg			Diameter Skin Area Observed		Pressure Duration	Petechiae Count		Method of:
			cm		Minutes	Value	Range	
(A)			(B)		(C)	(D)	(E)	(F)
11	Tourniquet Test <sup>5</sup> (Compression)	Negative 200	Fo	1	1		0-10	Aggeler & Lucia
12		Positive 40	Fo		5	0		Diggs
13		50	Fo	5	15		0-4	
14		35; 50 <sup>6</sup>	Fa	6	15			Gothlin
15		$\left[ \frac{\text{Syst.} + \text{Diast.}}{2} \right]^7$	Fo	5	8		0-10 <sup>8</sup>	Wright & Lillienfeld

/1/ Results vary with thickness, texture, and color of skin and other variables apart from the capillary resistance per se. /2/ Negative pressure applied by suction cup. /3/ Author's "Medianwerte" (derivation of these values obscure). Some hospital patients included. /4/ 94% range. /5/ Sphygmomanometer cuff placed on arm and inflated. /6/ Petechial index = 2 x (count after 35 mm Hg pressure, for 15 min.) plus, after 1 hr. 1 x (count after 50 mm Hg pressure, for 15 min); petechial index = 0-12. /7/ Pressure maintained at the midpoint between systolic and diastolic pressures of the individual. /8/ In 92% of persons, 10 or fewer petechiae; in 60%, none.

## 24. THE ABO SYSTEM<sup>1</sup> OF BLOOD FACTORS AND THEIR FREQUENCIES<sup>2</sup>

Blood Groups <sup>3</sup> (Phenotypes <sup>4</sup> )		Genetic Determinants (Genotypes <sup>5</sup> )	RBC Agglutino- gens <sup>6</sup>	Plasma Agglutin- ins <sup>8</sup>	Reactions between RBC and Antisera		
General	Differ- entiated				Anti-A	Anti-B	Anti-A <sub>1</sub>
O 45%	O 45.00%	O/O 45.00%	O <sup>7</sup>	Anti-A, Anti-A <sub>1</sub> & Anti-B	Neg. <sup>9</sup>	Neg	Neg.
A 41%	A <sub>1</sub> 31.39%	A <sub>1</sub> /A <sub>1</sub> 3.53%	A <sub>1</sub>	Anti-B	Pos. <sup>9</sup>	Neg.	Pos.
		A <sub>1</sub> /A <sub>2</sub> 2.56%					
		A <sub>1</sub> /O 25.30%					
	A <sub>2</sub> 9.61%	A <sub>2</sub> /A <sub>2</sub> 0.46%	A <sub>2</sub>		Pos.	Neg.	Neg.
		A <sub>2</sub> /O 9.15%					
B 10%	B 10.00%	B/B 0.72%	B	Anti-A and Anti-A <sub>1</sub>	Neg.	Pos.	Neg.
		B/O 9.28%					
AB 4%	A <sub>1</sub> B 2.94%	A <sub>1</sub> /B 2.94%	A <sub>1</sub> & B	None	Pos.	Pos	Pos.
	A <sub>2</sub> B 1.06%	A <sub>2</sub> /B 1.06%	A <sub>2</sub> & B		Pos.	Pos.	Neg.

/1/ Equivalents in obsolete terminologies of Moss and Jansky are: O = Moss IV, Jansky I; A = Moss and Jansky II; B = Moss and Jansky III; AB = Moss I, Jansky IV. /2/ Expected frequency, observed and calculated for U.S.A. whites.

/3/ Blood groups, named after agglutinogens. Group determined by testing red cells vs. natural antiserum B from group A, and natural antiserum A and A<sub>1</sub> from group B. /4/ Phenotype: the identifiable antigenic make-up of the red cells, dependent on, but not necessarily indicating the complete gene pattern.

/5/ Genotype: make-up of individual in terms of genes derived from parents. Diagonal (/) separates gene contributed by one parent from gene contributed by other parent. /6/ Agglutinin (antigen): Structure on surface (?) of RBC responsible for reaction with corresponding agglutinin in antiserum. /7/ Group O (first characterized as O due to failure of RBC to clump in either anti-A or anti-B sera) has own agglutinin, designated O. /8/ Agglutinin (antibody): Specific substance occurring in plasma, reacts with corresponding agglutinin, to cause agglutination, possibly hemolysis. Anti-O very rare agglutinin, regularly agglutinates RBC of group O and subgroup A<sub>2</sub>. /9/ Pos. = agglutination; Neg. = absence of agglutination.

# 25. THE DISTRIBUTION OF THE ABO BLOOD GROUPS IN VARIOUS POPULATIONS

Population Tested	Number Tested	Group O %	Group A %	Group B %	Group AB %
American Indians	194	98.5	1.5	0	0
Toba (Argentina)	115	23.5	76.5	0	0
Blackfeet (Montana)	310	78.4	20.0	1.6	6.2
Pueblo (New Mexico)	500	34.1	30.8	28.9	0.0
Arabs, Baghdad	805	53.1	44.7	2.1	10.0
Australian Aborigines	1,000	30.0	25.0	35.0	5.1
Chinese, Peiping	1,853	40.0	42.5	12.4	10.5
Denmark	516	26.6	35.7	27.1	7.0
Egypt, Cairo	1,600	46.0	30.0	17.0	3.7
English, Liverpool	1,063	36.3	54.6	5.3	6.9
Eskimos, S.W. Greenland	23,200	34.1	41.0	18.0	6.5
Finns	39,174	36.5	42.5	14.5	0.5
Germans, Berlin	413	36.5	60.8	2.2	8.5
Hawaii	1,000	31.3	19.0	41.2	5.4
Hindus, North	17,157	41.2	39.4	14.0	8.6
Italians	33,834	31.2	38.4	21.8	5.0
Japanese, Tokyo	500	43.2	22.4	29.2	7.1
Negroes, Senegal	763	33.7	38.4	20.8	5.1
Russia	1,200	37.9	46.7	10.3	6.6
Swedes	500	36.8	38.0	18.6	3.7
Turkey, Macedonia	1,003	49.3	26.0	21.0	4.0
U.S.A., Negroes	20,000	45.0	41.0	10.0	
U.S.A., Whites					

**26. HEREDITY OF THE ABO BLOOD GROUPS**  
Application in Disputed Paternity

If Parents' Groups are--	The Children's Groups	
	Can be--	Cannot be--
O and O	O	A, B, AB
O and A	A, O	B, AB
O and B	B, O	A, AB
A and A	A, O <sup>1</sup>	B, AB
A and B	AB, O <sup>3</sup> , A <sup>2</sup> , B <sup>1</sup>	None
B and B	B, O <sup>2</sup>	A, AB
O and AB	A, B	O, AB
A and AB	A, AB, B <sup>1</sup>	O
B and AB	B, AB, A <sup>2</sup>	O
AB and AB	A, B, AB	O

/1/ This type not possible if either parent is genetically A/A. /2/ This type not possible if one parent is genetically B/B. /3/ If one parent was genetically A/O and the other B/O, an O can result.

**27. HEREDITY OF THE ABO SUBGROUPS**  
Application in Disputed Paternity

If Parents' Blood Groups and Subgroups are--	The Children's Blood Groups and Subgroups	
	Can be--	Cannot be--
A <sub>1</sub> and O	O <sup>1</sup> , A <sub>1</sub> , A <sub>2</sub> <sup>2</sup>	B, A <sub>1</sub> B, A <sub>2</sub> B
A <sub>1</sub> and A <sub>1</sub>	O <sup>1</sup> , A <sub>1</sub> , A <sub>2</sub> <sup>3</sup>	B, A <sub>1</sub> B, A <sub>2</sub> B
A <sub>1</sub> and A <sub>2</sub>	O, A <sub>1</sub> , A <sub>2</sub>	B, A <sub>1</sub> B, A <sub>2</sub> B
A <sub>1</sub> and B	O, A <sub>1</sub> , A <sub>2</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B <sup>2</sup>	(None)
A <sub>1</sub> and A <sub>2</sub> B	A <sub>1</sub> , A <sub>2</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B	O
A <sub>1</sub> B and O	A <sub>1</sub> , B	O, A <sub>2</sub> , A <sub>1</sub> B, A <sub>2</sub> B
A <sub>1</sub> B and A <sub>1</sub>	A <sub>1</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B <sup>2</sup>	O, A <sub>2</sub>
A <sub>1</sub> B and A <sub>2</sub>	A <sub>1</sub> , B, A <sub>2</sub> B	O, A <sub>2</sub> , A <sub>1</sub> B
A <sub>1</sub> B and A <sub>2</sub> B	A <sub>1</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B	O, A <sub>2</sub>
A <sub>1</sub> B and B	A <sub>1</sub> , B, A <sub>1</sub> B	O, A <sub>2</sub> , A <sub>2</sub> B
A <sub>1</sub> B and A <sub>1</sub> B	A <sub>1</sub> , B, A <sub>1</sub> B	O, A <sub>2</sub> , A <sub>2</sub> B
A <sub>2</sub> and O	O, A <sub>2</sub>	A <sub>1</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B
A <sub>2</sub> and A <sub>2</sub>	O, A <sub>2</sub>	A <sub>1</sub> , B, A <sub>1</sub> B, A <sub>2</sub> B
A <sub>2</sub> and B	O, A <sub>2</sub> , B, A <sub>2</sub> B	A <sub>1</sub> , A <sub>1</sub> B
A <sub>2</sub> B and O	A <sub>2</sub> , B	O, A <sub>1</sub> , A <sub>1</sub> B, A <sub>2</sub> B
A <sub>2</sub> B and A <sub>2</sub>	A <sub>2</sub> , B, A <sub>2</sub> B	O, A <sub>1</sub> , A <sub>1</sub> B
A <sub>2</sub> B and B	A <sub>2</sub> , B, A <sub>2</sub> B	O, A <sub>1</sub> , A <sub>1</sub> B
A <sub>2</sub> B and A <sub>2</sub> B	A <sub>2</sub> , B, A <sub>2</sub> B	O, A <sub>1</sub> , A <sub>1</sub> B

/1/ O possible here if Blood Group A<sub>1</sub> (in 1st column) is of genotype A<sub>1</sub>O. /2/ A<sub>2</sub> possible if the parent of Blood Group A<sub>1</sub> is of the genotype A<sub>1</sub>/A<sub>2</sub>. /3/ A<sub>2</sub> possible if one parent is of genotype A<sub>1</sub>/A<sub>2</sub> and the other parent is A<sub>1</sub>/A<sub>2</sub> or A<sub>1</sub>O.

# 28. THE Rh-Hr (CDE-cde) SYSTEM OF BLOOD FACTORS<sup>1</sup> AND THEIR FREQUENCIES

Phenotypes and their Frequencies, <sup>3</sup> Classified by Agglutination Reactions by Antisera			Genotypes		
1 Anti-Serum (Anti-D)	3 Antisera (Anti-C, D, E)	4 Antisera (Anti-C, D, E, c, d, e)	Wieners <sup>2</sup> Terminology	Fisher-Race <sup>2</sup> Terminology	Calculated Frequencies (%)
Rh <sub>0</sub> (D) Negative Rh <sub>0</sub> 0, 4-14 %	rh <sup>+</sup> (CDE) 13.4-15.1 %	rr <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	LI	cde/cde	11.4 --- 15.1
	rh <sup>+</sup> (CDE) 1.1-0.8 %	r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	Cde/Cde	0.02 --- 0.01
	rh <sup>+</sup> (CDE) 0.4-0.9 %	r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	Cde/cde	1.2 --- 0.8
		r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	cDE/cDE	0.003 --- 0.014
	rh <sup>+</sup> rh <sup>+</sup> (CDE) (rh <sub>0</sub> ) 0.04-0.05 %	r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	cDE/cde	0.4 --- 0.9
		r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	Cde/cDE	0.02 --- 0.02
		r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	CDE/Cde	0.0001 --- 0.0001
		r <sup>+</sup> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + \\ - - \end{smallmatrix}$	r <sup>+</sup> r <sup>+</sup>	CDE/CDE	0.0001 --- 0.0001
Rh <sub>0</sub> (D) Positive Rh <sub>0</sub> 1-45 %	Rh <sub>0</sub> (CDE) 2.5-2.1 %	R <sub>0</sub> R <sub>0</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>0</sub> R <sub>0</sub>	cDe/cDe	0.1 --- 0.07
	Rh <sub>1</sub> (CDE) 51.2-52.4 %	R <sub>0</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>0</sub> r <sup>+</sup>	cDe/cde	2.4 --- 2.0
		R <sub>1</sub> R <sub>1</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>1</sub> R <sub>1</sub>	CDe/CDe	16.9 --- 17.7
		R <sub>1</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>1</sub> r <sup>+</sup>	CDe/Cde	1.3 --- 0.8
		R <sub>1</sub> R <sub>0</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>1</sub> R <sub>0</sub>	CDe/cDe	2.4 --- 2.1
	Rh <sub>2</sub> (CDE) 16.5-16.1 %	R <sub>1</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>1</sub> r <sup>+</sup>	CDe/cde	30.6 --- 32.7
		R <sub>2</sub> R <sub>2</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>2</sub> R <sub>2</sub>	cDE/cDE	2.7 --- 2.0
		R <sub>2</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>2</sub> r <sup>+</sup>	cDE/cDe	0.2 --- 0.3
		R <sub>2</sub> R <sub>0</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>2</sub> R <sub>0</sub>	cDE/cDe	1.2 --- 0.7
		R <sub>2</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>2</sub> r <sup>+</sup>	cDE/cde	12.5 --- 13.0
		R <sub>2</sub> R <sub>0</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>2</sub> R <sub>0</sub>	CDe/cDe	0.01 --- 0.01
	Rh <sub>3</sub> Rh <sub>4</sub> (CDE) (Rh <sub>0</sub> ) 14.7-13.6 %	R <sub>3</sub> R <sub>1</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> R <sub>1</sub>	CDE/CDe	0.05 --- 0.20
		R <sub>3</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> r <sup>+</sup>	CDE/Cde	0.001 --- 0.005
		R <sub>3</sub> R <sub>2</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> R <sub>2</sub>	CDE/CDE	0.01 --- 0.07
		R <sub>3</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> r <sup>+</sup>	CDE/cDE	0.0004 --- 0.0014
		R <sub>3</sub> R <sub>0</sub> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> R <sub>0</sub>	CDE/CDe	0.0001 --- 0.0006
		R <sub>3</sub> r <sup>+</sup> (CDEcde) $\begin{smallmatrix} + + + \\ - - - \end{smallmatrix}$	R <sub>3</sub> r <sup>+</sup>	CDE/CDE	0.0002 --- 0.0002

/1/ Note Plus (+) indicates agglutination by antisera. Thus C = agglutination of red cells by anti-C (anti-rh<sub>0</sub> and anti-rh<sub>1</sub>).



29. THE DISTRIBUTION OF THE Rh-Hr BLOOD TYPES IN VARIOUS POPULATIONS

Population Tested	No. of Persons Tested	Rh Positive						Rh Negative			
		Rh1		Rh2		Rh1Rh2	Rh1Rh2	rh		rh'	
		Rh1Rh1 %	Rh1rh %	Rh1Rh %	Rh2 %	Rh1Rh2 %	Rh1Rh2 %	rh %	rh' %	rh'rh'' %	rh'rh'' %
American Indians	95										
Mexico	105	1.1	40.7	7.4	9.5	38.1	3.1	0.0	0.0	0.0	0.0
Oklahoma	104	2.9	34.3	5.7	17.1	36.2	2.9	0.0	0.9	0.0	0.0
Ute	156	0.0	33.7	28.8	5.1	12.8	-	0.0	0.0	0.0	0.0
Asiatic Indians	100	1.9	70.5	21.0	15.0	6.0	-	7.1	2.6	0.0	0.0
Australian, Aborigines	350	4.0	39.0	14.0	12.6	16.6	-	0.0	1.0	0.0	0.0
Australian, Whites	132	0.6	54.0	3.0	34.1	0.1	-	14.9	0.9	0.0	0.0
Australian, Whites	927	0.9	60.6	12.2	3.6	-	-	1.5	0.0	0.0	0.0
Chinese	315	2.5	19.7	35.2	19.7	44.4	-	14.8	0.7	1.3	0.0
England, Whites	315	1.0	34.9	2.0	17.7	0.0	-	0.0	0.0	0.0	0.0
Eskimos	100	0.0	87.0	12.3	2.5	22.5	-	15.4	1.5	0.0	0.5
Eskimos	200	1.5	51.5	2.5	47.3	3.0	-	0.0	0.0	0.0	0.0
Filipinos	200	0.5	74.0	13.3	4.0	4.4	-	1.3	0.0	0.0	0.0
Holland, Whites	200	0.0	37.4	0.0	19.6	14.0	-	0.0	0.0	0.0	0.0
Holland, Whites	150	0.0	89.0	4.0	15.0	12.9	-	10.1	1.7	0.5	0.01
Indonesians	100	0.0	39.1	5.1	13.9	5.4	-	14.7	1.1	0.6	0.0
Japanese	100	15.1	53.5	33.8	22.4	4.4	-	12.5	0.9	0.5	0.0
Papuan	179	2.2	20.9	20.2	16.3	0.0	-	8.1	2.7	0.0	0.0
Puerto Ricans	7,317	2.2	20.9	20.2	16.3	0.0	-	7.4	1.5	0.7	0.0
U.S.A., Whites	766	41.2	0.9	22.8							
U.S.A., Whites	223	45.9									
U.S.A., Negroes	135										
U.S.A., Negroes											

### 30. THE MNS SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES<sup>1</sup>

Blood Types <sup>2</sup> (Phenotypes)		Genetic Determinants (Genotypes) <sup>3</sup>	RBC Agglutino- gens <sup>4</sup>	Reactions between RBC and Antisera <sup>5</sup>		
General	Differ- entiated			Anti-M	Anti-N	Anti-S
M 28.53%	Ms 7.74%	Ms/Ms 7.74%	M and s	Pos. <sup>6</sup>	Neg. <sup>6</sup>	Neg.
	MS 20.79%	MS/Ms 14.26%	M, S and s	Pos.	Neg.	Pos.
		MS/MS 6.53%	M and S			
N 21.68%	Ns 14.78%	Ns/Ns 14.78%	N and s	Neg.	Pos.	Neg.
	NS 6.90%	NS/Ns 6.24%	N, S and s	Neg.	Pos.	Pos.
		NS/NS 0.66%	N and S			
	MNs 21.45%	Ms/Ns 21.45%	M, N and s	Pos.	Pos.	Neg.
MN 49.80%	MNS 28.35%	MS/Ns 19.68%	M, N, S and s	Pos.	Pos.	Pos.
		Ms/NS 4.52%	M, N, S and s			
		MS/NS 4.15%	M, N and S			

/1/ Expected frequency, calculated for English whites. /2/ MNS system is independent of the ABO system. A person of group O, for example, may belong to type M, or N, or MN. Types take names after agglutinogens. M and N determined by testing red cells against induced antisera prepared by immunizing rabbits against red cells of known type. S types determined by testing blood with selected human sera containing anti-S. /3/ Diagonal (/) separates gene contributed by one parent from gene contributed by other parent. /4/ M, N, and S are relatively weak antigens in man. S is a property believed very closely linked, genetically, to the M-N alleles. s is assumed but has not been demonstrated with a specific antiserum, therefore, the S/s genotype (e.g., MS/MS) can be distinguished from the S/s genotype (e.g., MS/Ms) only by family studies. /5/ Natural anti-M and anti-N occur rarely. Anti-S has not been identified as a natural agglutinin and has not been produced in animals. Anti-M and anti-N typing sera obtained by immunizing rabbits with human type M and type N cells, respectively. /6/ Pos. = agglutination, Neg. = absence of agglutination.

**31. THE DISTRIBUTION OF THE MN BLOOD FACTORS  
IN VARIOUS POPULATIONS**

Population	No. Tested	Frequency of Types, %		
		M	N	MN
Ainu	504	17.9	31.9	50.2
American Indians				
U.S.A. <sup>1</sup>	205	60.0	4.9	35.1
Blackfeet	95	54.7	5.3	40.0
Australian				
Aborigines	730	3.0	67.4	29.6
Chinese	1,029	33.2	18.2	48.6
Danes	2,023	29.1	21.4	49.5
Egyptians, Cairo	613	28.3	23.1	48.6
English	1,522	30.5	21.4	48.2
Eskimos				
S.W. Greenland	377	67.6	2.4	30.0
Finns	6,926	42.3	13.7	44.0
French	1,400	30.1	19.8	50.1
Germans	40,255	30.2	19.7	50.0
Hindus	300	42.7	10.7	46.7
Italians	736	28.9	17.1	53.9
Japanese	7,551	29.0	21.1	49.9
Russians	763	32.2	21.2	46.5
Swedes	1,200	36.1	16.9	47.0
U.S.A. Negroes	278	28.4	21.9	49.6
U.S.A. Whites	6,129	29.2	21.3	49.6

/1/ Information on tribe not available.

**32. HEREDITY OF THE MN BLOOD FACTORS:  
Application in Disputed Paternity**

If Parents' Factors are--	The Children's Factors	
	Can be--	Cannot be--
MN and MN	M, N, MN	None
MN and M	M, MN	N
MN and N	N, MN	M
M and N	MN	M, N
M and M	M	N, MN
N and N	N	M, MN

### 33. THE K ("KELL-CELLANO") SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES<sup>1</sup>

Blood Factors (Phenotypes)			Genetic Determinants (Genotypes) <sup>2</sup>	RBC Agglutin- ogens <sup>3</sup>	Reactions between RBC and Antisera <sup>4</sup>	
					Anti- Kell (Anti-K)	Anti- Cellano (Anti-k)
Cellano Negative 0.2- 0.27%	Kell Positive 8.8-10.17%	KK	K/K 0.2-0.27%	K	Pos. <sup>5</sup>	Neg. <sup>5</sup>
Cellano Positive 99.8-99.73%		Kk	K/k 8.6-9.90%	K and k	Pos.	Pos.
		kk	k/k 91.2-89.83%	k	Neg.	Pos.

/1/ The first % = frequency of occurrence among U.S.A. whites; the second, among English whites. /2/ Diagonal (/) separates gene contributed by one parent from gene contributed by the other parent. /3/ The Kell-Cellano factors named after the patients from whom the agglutinins were first obtained. /4/ No natural agglutinins have been found in man, nor have agglutinins been obtained from animals. Antisera for typing are prepared by absorption of unwanted agglutinins (as anti-A and anti-B) with red cells containing A and/or B, or by use of saliva or other A and B containing substances. Since K/K individuals are rare, anti-k is difficult to obtain. Only one example of anti-k has been found, the result of active immunization of a K/K woman by her K/k infants, causing her to develop k antibodies. Many examples of anti-K are known. /5/ Pos. = agglutination; Neg. = absence of agglutination.

**34. THE Le ("LEWIS") SYSTEM OF BLOOD FACTORS  
AND THEIR FREQUENCIES<sup>1</sup>**

Blood Factors (Phenotypes)		Genetic Determinants (Genotypes <sup>2</sup> )	RBC Agglutino- gens	Reactions between RBC and Antisera <sup>4</sup>	
				Anti-Le <sup>a</sup>	Anti-Le <sup>b</sup>
Lewis positive 20.5%	Le(a+b-)	Le <sup>a</sup> /Le <sup>a</sup>	Le <sup>a</sup>	Pos. <sup>5</sup>	Neg. <sup>5</sup>
Lewis negative 79.5%	Le(a-b+)	Le <sup>a</sup> /Le <sup>b</sup>	Le <sup>b</sup> and Le <sup>a3</sup>	Neg.	Pos.
		Le <sup>b</sup> /Le <sup>b</sup>	Le <sup>b</sup>	Neg.	Pos.
	Le(a-b-)	?	?	Neg.	Neg.

/1/ Frequency observed for U.S.A. whites. /2/ Le<sup>a</sup> = "Lewis" (Mourant) = L<sub>1</sub> (Andresen); Le<sup>b</sup> = L<sub>2</sub> (Andresen). Diagonal (/) separates gene contributed by one parent from gene contributed by other parent. /3/ Le<sup>a</sup> may be present; thus far available antisera do not detect it. Some persons negative to both anti-Le<sup>a</sup> and Le<sup>b</sup>, indicating the possible existence of a third, fairly common allele. /4/ All Anti-Lewis agglutinins are believed to be naturally occurring, but they are not common, and have not been obtained from animals. Typing sera are prepared by absorption of unwanted agglutinins (as anti-A and anti-B) with red cells containing agglutinogens A and/or B. /5/ Pos. = agglutination; Neg. = absence of agglutination.

**35. THE Lu ("LUTHERAN") SYSTEM OF BLOOD FACTORS  
AND THEIR FREQUENCIES<sup>1</sup>**

Blood Factors (Phenotypes)		Genetic Determinants (Genotypes <sup>2</sup> )	RBC Agglutino- gens <sup>3</sup>	Reactions <sup>4</sup> between RBC and Anti-Lu <sup>a</sup>
Lutheran positive 8.0%	Lu (a+)	Lu <sup>a</sup> /Lu <sup>a</sup> 0.2%	Lu <sup>a</sup>	Pos. <sup>5</sup>
		Lu <sup>a</sup> /Lu <sup>b</sup> 7.8%	Lu <sup>a</sup> and Lu <sup>b</sup>	Pos.
Lutheran negative 92.0%	Lu (a-)	Lu <sup>b</sup> /Lu <sup>b</sup> 92.0%	Lu <sup>b</sup>	Neg. <sup>5</sup>

/1/ Expected frequency, observed and calculated for English whites, U.S.A.: Lu-positive, 19.1%; Lu-negative, 80.9%. /2/ Diagonal (/) separates gene contributed by one parent from gene contributed by other parent. /3/ Lu<sup>b</sup> is assumed but has not been demonstrated with specific antiserum. /4/ No natural agglutinins have been found, but have been produced by deliberate transfusion. Anti-Lu<sup>a</sup> has not been obtained from animals. Antisera prepared by absorption of unwanted agglutinins (as anti-A and anti-B) with red cells containing agglutinogens A and/or B. Donors whose sera contain agglutinins due to previous sensitization are very rare. /5/ Pos. = agglutination; Neg. = absence of agglutination.

**36. THE P SYSTEM OF BLOOD FACTORS  
AND THEIR FREQUENCIES<sup>1</sup>**

Blood Factors (Pheno-types) <sup>2</sup>		Genetic Determinants (Geno-types) <sup>3</sup>	R B C Agglutinogens	Plasma Agglutinins <sup>4</sup>	Reactions <sup>5</sup> between R B C and Anti-P
P Positive 71.4%	PP	P/P 21.6%	P	?	Pos. <sup>6</sup>
	Pp	P/p 49.8%	P & p	?	Pos.
P Negative 28.6%	pp	p/p 28.6%	p	Anti-P <sup>4</sup>	Neg. <sup>6</sup>

/1/ Expected frequencies calculated for U. S. A. whites.

/2/ p is assumed, but has not been demonstrated with a specific antiserum. /3/ Diagonal (/) separates gene contributed by one parent from that contributed by other parent.

/4/ Anti-P is found occasionally, in very low titre, in P-negative individuals. It has been assumed to be a naturally occurring agglutinin and has frequently been obtained from normal animals. /5/ Typing sera are prepared by absorption of unwanted agglutinins (as anti-A and anti-B) with red cells containing agglutinogens A and/or B. /6/ Pos. = agglutination; Neg. = absence of agglutination.

# 37. BLOOD ERYTHROCYTE VALUES: MAN

Variable				Variable			
(A)		(B)	(C)	(A)		(B)	(C)
1	RBC Count	$\sigma$ 5.4	4.6-6.2 <sup>b</sup>	36	RBC Sedimentation Rate (mm/hr)	Westergren Method	$\sigma$ 0-15
2	(millions/mm <sup>3</sup> blood)	$\sigma$ 4.8	4.2-5.4 <sup>b</sup>	37			$\sigma$ 0-20
3	RBC Packed Volume <sup>1</sup> (Hematocrit)	$\sigma$ 5.1	4.2-6.2 <sup>b</sup>	38		Wintrobe Method	$\sigma$ 0-9
4		$\sigma$ 47.2, 4	40-54 <sup>b</sup>	39			$\sigma$ 0-15
5	(ml/100 ml blood)	$\sigma$ 42.4	37-47 <sup>b</sup>	40	RBC Fragility (% NaCl Solution)	Cutler Method	$\sigma$ 0-8
6	Blood Hemoglobin Concentration	$\sigma$ 44.5 <sup>2</sup>		41			$\sigma$ 0-10
7	(g/100 ml blood)	$\sigma$ 15.8	14.0-18.0 <sup>b</sup>	42		Daland & Worthley Method Initial Final	$\sigma$ 0.47
8		$\sigma$ 13.9	11.5-16.0 <sup>b</sup>	43			$\sigma$ 0.27
9	RBC <sup>3</sup> Hemoglobin Concentration	$\sigma$ 14.9		44	RBC Fragility (% NaCl Solution)	Giffin & Sanford Method Initial Final	0.48-0.46
10		$\sigma$ 33.5 <sup>2,4</sup>	30-40 <sup>b</sup>	45			$\sigma$ 0.44-0.42
11				46		$\sigma$ 0.34-0.32	
12				47		Parpart Method Mean	$\sigma$ 0.43
13				48		0.54-0.32	
14				49	Erythrocyte Charge <sup>14</sup> (millivolts)	$\sigma$ -16.8	
15	Corpuscle <sup>3</sup> Thickness	$\sigma$ 2.06	1.7-2.2 <sup>b</sup>	50	Production Rate per Day	Electrophoretic Mobility <sup>14</sup> (cm <sup>2</sup> /volt sec.)	$\sigma$ 1.31 x 10 <sup>-4</sup>
16	( $\mu$ )	$\sigma$ 2.4		51			Erythrocyte pH
17	Corpuscle <sup>3</sup> Surface Area	$\sigma$ 135 <sup>7</sup>	129-146 <sup>b</sup>	52		Erythrocyte Life Span <sup>15</sup> (days)	
18	( $\mu^2$ )	$\sigma$ 163		53			$\sigma$ 109
19	Corpuscle <sup>2,3,4</sup> Volume ( $\mu^3$ )	$\sigma$ 87	70-94 <sup>b</sup>	54	Production Rate per Day	RBC (number/mm <sup>3</sup> blood)	$\sigma$ 45,000
20		$\sigma$ 87	74-98 <sup>b</sup>	55			$\sigma$ 40,000
21		$\sigma$ 87		56		$\sigma$ 42,500	
22	Corpuscle <sup>3</sup> Mass <sup>9</sup> ( $\mu$ g)	$\sigma$ 96	77-103 <sup>b</sup>	57		RBC (millions/kg body wt)	$\sigma$ 3,500
23	Corpuscle Specific Gravity <sup>10</sup>	$\sigma$ 1.0983	0.95-1.10 <sup>b</sup>	58	$\sigma$ 2,630		
24	Corpuscle <sup>3</sup> Hemoglobin Content ( $\mu$ g)	$\sigma$ 29	25-34 <sup>b</sup>	59	Production Rate per Day	RBC blood equiv (ml blood/kg body wt)	$\sigma$ 0.65
25		$\sigma$ 29	24-33 <sup>b</sup>	60			$\sigma$ 0.55
26		$\sigma$ 29	23-35 <sup>b</sup>	61		$\sigma$ 0.60	
27	Corpuscle <sup>3</sup> Iron <sup>11</sup> Content ( $\mu$ g)	$\sigma$ 0.10	0.08-0.12 <sup>b</sup>	62		RBC & Hb replaced (% of total)	$\sigma$ 0.83
28	RBC Volume Coefficient <sup>12</sup>	$\sigma$ 43.5		63	Production Rate per Day		Hemoglobin (g/100 ml blood)
29	(ml RBC / 100 ml blood, 5 million count)	$\sigma$ 43.7		64		$\sigma$ 0.11	
30		$\sigma$ 43.6		65		$\sigma$ 0.12	
31	RBC Hemoglobin Coefficient <sup>12</sup>	$\sigma$ 14.6					
32	(g/100 ml blood, 5 million count)	$\sigma$ 14.4					
33		$\sigma$ 14.5					
34	Color, Volume and Saturation Index <sup>12</sup>	$\sigma$ 1.00	0.85-1.15 <sup>b</sup>				
35	Corpuscle Spherocytic Index	$\sigma$ 0.27					

1/1 Centrifuged at 2000 g or over (= 3000 rpm) for 10 minutes after attaining constant packed cell volume,  $F = m \cdot w \cdot r$  (or,  $F = (1.118 \times 10^{-3}) r n^2 m$ ) where  $w$  = angular velocity  $r$  = radius of circle of rotation in cm, and  $n$  = rpm  
2/2 Heparin or other isotonic anticoagulant  
3/3 "Mean corpuscular"  
4/4 When anticoagulant = 2 mg K oxalate/ml blood, mean and 95% range for 4BC = 45 (40-50), 5BC = 41 (36-45), 10BC = 35 (30-40), 12BC = 34 (30-36)  
micromet  
(13B) by  $f$   
 $2\pi r(x+t)$   
9/9 Calculated from water saturated iron is qu  
pH 7.4  
15/15 Use of isotopic nitrogen Other methods in essential agreement



# 38. BLOOD ERYTHROCYTE, HEMATOCRIT, HEMOGLOBIN AND THROMBOCYTE VALUES, BIRTH TO MATURITY: MAN

Age	Erythrocyte Count		Reticulocyte Count		Eryth. Packed Volume (Hematocrit)		Erythrocyte Volume		Hemoglobin Concentration		Eryth. Hemoglobin Concentration <sup>2</sup>		Thrombocyte (Platelet) Count <sup>4</sup>	
	millions/mm <sup>3</sup>		% of total erythrocytes		ml/100ml blood		μ <sup>3</sup>		g/100ml blood		μg		thousands/mm <sup>3</sup>	
	Value <sup>5</sup> (H)	Range <sup>6</sup> (C)	Value <sup>5</sup> (D)	Range <sup>6</sup> (E)	Value <sup>5</sup> (F)	Range <sup>6</sup> (G)	Value <sup>5</sup> (I)	Range <sup>6</sup> (J)	Value <sup>5</sup> (K)	Range <sup>6</sup> (L)	Value <sup>5</sup> (M)	Range <sup>6</sup> (N)	Value <sup>5</sup> (O)	Range <sup>6</sup> (P)
1 At birth <sup>7</sup>	5.7	4.8-7.1 <sup>c</sup>	4.35	2.50-6.50 <sup>d</sup>	56.6	106	21.5	18.0-27.0 <sup>c</sup>	38	38.0	227	140-290 <sup>c</sup>		
2 First Day	5.6	4.7-7.0 <sup>c</sup>			56.1	106	21.2	17.7-26.5 <sup>c</sup>	38	37.8				
3 End 1st wk.	5.3	4.5-6.4 <sup>c</sup>	1.12	0.10-4.50 <sup>d</sup>	52.7	101	19.6	16.2-25.5 <sup>c</sup>	37	37.2	235	150-320 <sup>c</sup>		
4 End 2nd wk.	5.1	4.3-6.0 <sup>c</sup>	0.67	0.20-1.50 <sup>d</sup>	49.6	96	18.0	14.5-24.2 <sup>c</sup>	35	36.3	247	163-340 <sup>c</sup>		
5 End 3rd wk.	4.9	4.1-6.0 <sup>c</sup>	0.63	0.20-1.30 <sup>d</sup>	46.6	93	16.6	13.2-23.0 <sup>c</sup>	34	35.6	267	177-367 <sup>c</sup>		
6 End 4th wk.	4.7	3.9-5.9 <sup>c</sup>	0.73	0.10-1.00 <sup>d</sup>	44.6	91	15.6	12.0-21.8 <sup>c</sup>	33	35.0	280	185-390 <sup>c</sup>		
7 End 2nd mo.	4.5	3.8-5.8 <sup>c</sup>	1.20	0.50-3.10 <sup>d</sup>	38.9	85	13.3	10.8-18.0 <sup>c</sup>	30	34.2	315	200-428 <sup>c</sup>		
8 End 4th mo.	4.5	3.8-5.3 <sup>c</sup>	1.66	0.90-2.94 <sup>d</sup>	36.5	79	12.4	10.2-15.0 <sup>c</sup>	27	34.0	338	205-465 <sup>c</sup>		
9 End 6th mo.	4.6	3.9-5.3 <sup>c</sup>	1.38	0.72-2.30 <sup>d</sup>	36.2	78	12.3	10.0-15.0 <sup>c</sup>	27	34.0	340	205-470 <sup>c</sup>		
10 End 8th mo.	4.6	4.0-5.4 <sup>c</sup>	1.12	0.65-1.90 <sup>d</sup>	35.8	77	12.1	9.8-15.0 <sup>c</sup>	26	33.8	345	210-473 <sup>c</sup>		
11 End 10th mo.	4.6	4.0-5.5 <sup>c</sup>	0.97	0.62-1.80 <sup>d</sup>	35.5	77	11.9	8.4-14.9 <sup>c</sup>	26	33.5	345	212-470 <sup>c</sup>		
12 End 12th mo.	4.6	4.0-5.5 <sup>c</sup>	0.90	0.60-1.70 <sup>d</sup>	35.2	77	11.6	9.0-14.6 <sup>c</sup>	25	33.0	345	218-470 <sup>c</sup>		
13 End 2nd yr.	4.7	3.8-5.4 <sup>c</sup>			35.5	78	11.7	9.2-15.5 <sup>c</sup>	25	33.0				
14 End 4th yr.	4.7	3.8-5.4 <sup>c</sup>			37.1	80	12.6	9.6-15.5 <sup>c</sup>	27	34.0				
15 End 6th yr.	4.7	3.8-5.4 <sup>c</sup>			37.9	80	12.7	10.0-15.5 <sup>c</sup>	27	33.5				
16 End 8th yr.	4.7	3.8-5.4 <sup>c</sup>			38.9	80	12.9	10.3-15.5 <sup>c</sup>	27	33.2				
17 End 10th yr.	4.8	3.8-5.4 <sup>c</sup>			39.0	80	13.0	10.7-15.5 <sup>c</sup>	27	33.3				
18 End 12th yr.	4.8	3.8-5.4 <sup>c</sup>			39.6	81	13.4	11.0-16.5 <sup>c</sup>	28	33.8				
19 14 yrs. & over														
20 Males	5.4	4.6-6.2 <sup>b</sup>			47.9, 51.0	87, 91.0	15.8	14.0-18.0 <sup>b</sup>	29.10	33.5 <sup>9</sup>				
21 Females	4.8	4.2-5.4 <sup>b</sup>			42.9, 51.0	87, 91.0	13.9	11.5-16.0 <sup>b</sup>	29.10	33.5 <sup>9</sup>				
22 M & F Av.	5.1	4.2-6.2 <sup>b</sup>	1.5	0.5-3.8 <sup>c</sup>	44.5, 51.9	87.9	14.9		29.10	33.5 <sup>9, 10</sup>	409 <sup>4</sup>	273-545 <sup>4</sup>		

<sup>1/1</sup> = Mean Corpuscular Volume (M.C.V.). <sup>2/2</sup> = Mean Corpuscular Hemoglobin (M.C.H.). <sup>3/3</sup> = Mean Corpuscular Hemoglobin Concentration (M.C.H.C.) in %. <sup>4/4</sup> Cutaneous blood. Direct method of Wood, Vogel, and Famaletti except 22 LM=direct method of Hees-Ecker. <sup>5/5</sup> Smoothed means from plotted curves. <sup>6/6</sup> Smoothed sample ranges from plotted curves (except as noted). <sup>7/7</sup> If cord clamped after placental separation, average 560,000 more erythrocytes per mm<sup>3</sup> and 2.6 g/100ml more hemoglobin during the first week of life than if cord clamped immediately after birth. Also in newborn, heel blood (capillary) higher in erythrocytes and hemoglobin than blood from superior sagittal sinus. <sup>8/8</sup> Centrifuged at 2000 g (3000 rpm) or over, for 10 min, after attaining constant packed cell volume. <sup>9/9</sup> Heparin or other isotonic anticoagulant. <sup>10/10</sup> Estimate "b" of 95% range: 40-54 for 20F; 37-47 for 21F; 70-94 for 20G; 74-98 for 21G; 25-34 for 20J; 24-33 for 21J; 23-35 for 22J; 30-40 for 22K.

39. BLOOD ERYTHROCYTE AND HEMOGLOBIN VALUES AT OR NEAR SEA LEVEL: MAN <sup>1</sup>				
Country	Place <sup>2</sup>	RBC Count millions/mm <sup>3</sup> blood	Blood Hemoglobin Concentration g/100ml blood	Corpuscle Hb Content <sup>3</sup> µg
(A)	(B)	(C)	(D)	(E)
1 U. S.	Portland, Oregon	5.39	15.8	29.3
2 U. S.	Portland, Oregon	5.42	15.8	29.2
3 U. S.	New Orleans	5.85	15.9	27.2
4 U. S.	New Orleans	5.26	15.6	29.7
5 U. S.	Kansas	4.84	15.1	31.2
6 U. S.	Omaha	4.69	15.0	32.0
7 U. S.	Kansas	5.11	15.0	29.4
8 Argentina	Buenos Aires	5.50	15.4	28.0
9 Argentina	Buenos Aires	5.30	14.8	27.9
10 Denmark	Copenhagen	5.07	15.0	29.6
11 Germany	Jena	5.06	16.0	31.6
12 Germany	Giessen	4.96	16.0	32.3
13 Hawaii	Honolulu	5.08	15.1	29.7
14 India	Calcutta	5.36	14.8	27.6
15 India	Bombay	5.11	15.4	30.1
16 Norway	Oslo	5.52	16.2	29.3
17 Peru	Lima	5.26	15.7	29.8
18 Peru	Lima	5.14	16.0	31.1
19 Summary, U. S. <sup>4</sup>		5.22	15.5	29.7
20 Summary, all countries		5.22	15.5	29.7

/1/ Males. /2/ Altitudes of less than 0.4 Km. /3/ Mean Corpuscular Hemoglobin (M. C. H.).  
/4/ For standards see Table 37.

# 40. BLOOD ERYTHROCYTE AND HEMOGLOBIN VALUES AT ALTITUDE: MAN<sup>1</sup>

Residents (R); Sojourners (S).

Country	Place	Altitude Km	RBC Count millions/mm <sup>3</sup> blood	Blood Hemoglobin Concentration g/100ml blood	Corpuscle Hb Content <sup>2</sup> µg
(A)	(B)	(C)	(D)	(E)	(F)
1 Sea Level <sup>3,4</sup>		<0.4	R 5.22	R 15.5	29.7
2 Argentina	Tucuman	0.4	R 5.31	R 16.1	30.3
3 Canada	Saskatchewan	0.5	R 5.52	R 15.6	28.3
4 Switzerland	Zurich	0.5	R 5.00	R 15.0	30.0
5 U. S.	Denver, Colo.	1.5	R 5.42	R 16.5	30.4
6 So. Africa	Johannesburg	1.8	R 5.99	R 14.7	24.5
7 Mexico	Mexico City	2.3	R 7.35	R 18.4	25.0
8 Mexico	Mexico City	2.3	R 5.38	R 17.7	32.9
9 Chile	Chuquicamata	2.8	S 5.24	S 15.9	30.3
10 Chile	Ollague	3.7	S 5.54	S 17.0	30.7
11 Peru	Oroya	3.7	R 5.67	R 18.8	33.2
12 Argentina	Mina Aguilar	4.5	R 6.46	R 19.4	30.0
13 Peru	Morococha	4.5	R 6.15	R 20.8	33.8
14 Chile	Montt	4.7	S 5.84	S 17.6	30.1
15 Chile	Quilcha	5.3	R 7.37	R 22.6	30.7
16 Chile	Quilcha	5.3	S 5.95	S 18.4	30.9
17 Chile	Punta	6.1	S 5.77	S 18.3	31.7
18 India	Nanga Parbat	7.0	S 8.10	S 24.7	30.5

/1/ Males. /2/ Mean Corpuscular Hemoglobin (M.C.H.). /3/ Altitudes of less than 0.4 Km  
/4/ For standards see Table 37.

41. BLOOD ERYTHROCYTE VALUES IN PREGNANCY AND POSTPARTUM: MAN

Variable	Second Trimester				Third Trimester				During Labor		Postpartum	
	Value and Range				Value and Range				Value and Range	(I)	Value and Range	
	4th Month (B)	5th Month (C)	6th Month (D)	7th Month (E)	8th Month (F)	9th Month (G)	9th Month (G)	9th Month (G)			10 Days (J)	42 Days (J)
1 RBC Count (millions/mm <sup>3</sup> blood)	4.5 3.8-5.2b	4.3 3.7-5.0b	4.0 3.5-4.8b	4.0 3.5-4.8b	4.1 3.5-4.8b	4.2 3.7-5.0b	4.2 3.7-5.0b	4.2 3.7-5.0b	4.4 4.0-5.0b	4.5 4.0-5.0b	4.5 4.2-5.4b	4.8 4.2-5.4b
3 Packed RBC Volume (hematocrit) (ml/100 ml blood)	40 35-45b	39 34-44b	37 32-42b	37 32-42b	37.5 33-43b	37.5 33-43b	37.5 33-43b	37.5 33-43b	39 34-44b	40 35-45b	42.5 37-47b	42.5 37-47b
5 Blood Hemoglobin Concentration (g/100 ml blood)	12.8 11.4-15.0b	12.2 10.8-14.6b	11.4 10.2-14.0b	11.4 10.2-14.0b	11.6 10.4-14.2b	12.0 10.8-14.4b	12.0 10.8-14.4b	12.0 10.8-14.4b	12.6 11.2-15.0b	12.8 11.4-15.4b	13.8 12.0-16.0b	13.8 12.0-16.0b
7 RBC Hemoglobin Concentration (g/100 ml RBC)	32	31	31	31	31	32	32	32	32	32	32.5	32.5
8 Corpuscle Hemoglobin Content (μg)	28.4	28.4	28.5	28.5	28.3	28.5	28.5	28.5	28.6	28.4	28.7	28.7
9 Corpuscle Volume (μ <sup>3</sup> )	89	91	92	92	91	89	89	89	89	89	89	89

42. BLOOD ERYTHROCYTE VALUES: MAMMALS, LABORATORY

Animal	RBC Count millions/mm <sup>3</sup> blood		Reticulocytes % of total RBC		Corpuscle Diameter $\mu$		Corpuscle Volume $\mu^3$	
	Value	Range	Value	Range	Value	Range	Value	Range
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
1 Cat	8.0	6.5-9.5	0.2		6.2	5-7.2	57	51-63
2 Dog	6.3	4.5-8	0.7	0-2.7	7.2	6.2-8.0 <sup>2</sup>	66	59-68
3 Guinea pig	5.6	4.5-7	0.9	0.4-1.8	7.4 <sup>2</sup>	7.0-7.5 <sup>2</sup>	77	71-83
4 Mouse	9.3	7.7-12.5	4.0		6.2		49	48-51
5 Rabbit	5.7	4.5-7	2.2	2-3	7.5 <sup>3</sup>	6.5-7.5 <sup>2</sup>	61	60-68
6 Rat	8.9 <sup>1</sup>	7.2-9.6 <sup>1</sup>	2.9	0.6-4.9	7.5 <sup>3</sup>	6.0-7.5 <sup>2</sup>	61	57-65

Animal	Packed RBC Volume (Hematocrit) ml/100ml blood		Blood Hemo- globin Conc. g/100ml blood		Corpuscle Hemo- globin Conc. g/100ml RBC		Corpuscle Hemo- globin Content $\mu\text{g}$	
	Value	Range	Value	Range	Value	Range	Value	Range
(A)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)
1 Cat	40.0	28-52	11.2	7.0-15.5	28	23-31	14	12-16
2 Dog	45.5	38-53	14.8	11.0-18.0	33	30-35	23	21-25
3 Guinea pig	42.0	37-47	14.4	11-16.5	34	33-35	26	24.5-27.5
4 Mouse	41.5		14.8	10-19	36	33-39	16	15.5-16.5
5 Rabbit	41.5	33-50	11.9	8.0-15.0	29	27-31	21	19-23
6 Rat	46.0 <sup>1</sup>	39-53 <sup>1</sup>	14.8	12-17.5	32	30-35	17	15-19

/1/ More weight given to common strains. /2/ Dry film. /3/ In plasma.

# 43. BLOOD HEMATOCRIT AND HEMOGLOBIN VALUES: VERTEBRATES<sup>1</sup>

Animal		Packed RBC Volume (Hematocrit) <sup>2</sup> ml/100ml blood		Blood Hemoglobin Concentration g/100ml blood		RBC Hemoglobin Concentration g/100ml blood
		Value	Range	Value	Range	Value
(A)		(B)	(C)	(D)	(E)	(F)
1	Man	44.5 <sup>4</sup>		14.9 <sup>5</sup>		33.5
2	Chimpanzee	41.5		13.0		31
3	Cow	40	33-47 <sup>b3</sup>	11.5	8.7-14.5 <sup>b3</sup>	29
4	Goat	34	20-50 <sup>b</sup>	10.6	7.3-14 <sup>b3</sup>	31
5	Hamster	46	36-54 <sup>c</sup>	13.9	11-17.4 <sup>c</sup>	30
6	Horse	33.4	28-42 <sup>b3</sup>	11.1	8-14 <sup>b3</sup>	33
7	thoroughbred	42		13.9		33
8	Monkey, rhesus	42	32-52 <sup>b3</sup>	12.6	10-16	30
9	Pig	41.5	30-53 <sup>b3</sup>	13.3	10-16.5 <sup>b3</sup>	32
10	Sheep	32	22-40 <sup>b3</sup>	12.6	10-15.5 <sup>b3</sup>	39
11	Chicken	32		11.2 <sup>6</sup>		35
12	Duck	39.5		14.8	9-21 <sup>b3</sup>	37
13	Goose	44		12.8		29
14	Pigeon	42.5		15.3		36
15	Turkey	38		11.2		29
	Fish					
16	carp	31.3	21-40 <sup>b</sup>	10.5	9.4-12.4 <sup>b</sup>	34
17	mackerel	51.5		14.5		28
18	trout	27.2	22-36 <sup>b</sup>	8.5	6.2-11.5 <sup>b</sup>	31
	Frog					
19	Rana catesbeiana	30		7.8		26
20	Rana pipiens				10-12	
21	Snake, garter	28		8.6		31
22	Turtle, box	25	21-27 <sup>b</sup>	7.2	6.1-9.1 <sup>b</sup>	29

/1/ See Table No. 42. /2/ Determined by centrifugation under standard conditions. /3/ Standard deviation from which this range is calculated is from a subgroup whose mean is averaged with one or more others to obtain the value as presented. /4/ Average for both sexes. Hematocrit for males 47, range = 40-54<sup>b</sup>; females = 42, range = 37-47<sup>b</sup>. /5/ Average for both sexes. Hb for males = 15.8, range = 14-18<sup>b</sup>; females = 13.9, range = 11.5-16<sup>b</sup>. /6/ Average for both sexes. Hb for cocks = 12.9, hens = 9.4.

# 44. BLOOD ERYTHROCYTE VALUES: FISH

Animal	RBC Count millions/mm <sup>3</sup> blood	Hematocrit ml/100ml blood	Hemoglobin <sup>1</sup> g/100ml blood	RBC Diameters $\mu$	RBC Volume $\mu^3$
(A)	(B)	(C)	(D)	(E)	(F)
1 Bullhead ( <i>Ameiurus nebulosus</i> )	1.8	26	14.7		311
2 Carp ( <i>Cyprinus carpio</i> )	0.84	31	10.5		278-340
3 Range:	0.65-1.13	21-40	9.4-12.4		
4 Dogfish ( <i>Mustelus canis</i> )	0.46	23.3	4.6	19.1 x 13.8	541
5 Dogfish ( <i>Squalus acanthias</i> )	0.24	18.9	3.8	22.7 x 15.2	820
6 Eel ( <i>Anguilla bostoniensis</i> )	2.4	36	9.3	12.3 x 8.1	149
7 Eel pout ( <i>Zoarces anguillar</i> )	2.94	20.8	4.0	10.4 x 8.7	102
8 Goosefish ( <i>Lophius piscatorius</i> )	1.09	16.8	4.3	13.3 x 9.6	241
9 Hagfish ( <i>Myxine glutinosa</i> )	0.15	22.2	4.6	26.4 x 18.3	1530
10 Lamprey eel ( <i>Petromyzon marinus</i> )	0.33	23.5	5.8	14.3	710
11 Mackerel ( <i>Scomber scombrus</i> )	3.9	46	14.2	12.0 x 8.3	118
12 Menhaden ( <i>Brevoortia tyrannus</i> )	3.2		13.4	9.8 x 7.2	148
13 Pike ( <i>Esox lucius</i> )	1.10	27			
14 Range:	0.40-1.37	16-33			
15 Puffer ( <i>Sphoeroides maculatus</i> )	4.2		7.4	8.8 x 6.9	77
16 Sea robin, common ( <i>Priodontus carolinus</i> )	2.5	24	7.1	9.8 x 6.5	95
17 Sea robin, Northern striped ( <i>Priodontus strigatus</i> )	1.93	22.2	6.2	10.4 x 7.3	130
18 Shark sucker ( <i>Echenis naucrates</i> )	3.75	34	10.5	10.9 x 7.0	91
19 Skate, barndoor ( <i>Raja stibuliforis</i> )	0.27	20.0	3.6	21.9 x 15.6	727
20 Skate, clearnose ( <i>Raja eglanteria</i> )	0.30	24	4.5	23.7 x 14.4	823
21 Stingray ( <i>Dasyatis centrourus</i> )	0.30	19	3.0	20.6 x 14.3	612
22 Sturgeon ( <i>Acipenser sturio</i> )	0.43	21.5	5.0	14.0 x 9.7	506
23 Tautog ( <i>Tautoga onitis</i> )	2.05		6.7	10.8 x 7.3	
24 Trout ( <i>Salvelinus fontinalis</i> )	1.01	27	8.5		314
25 Range:	0.74-1.55	22.2-36	6.2-11.5		284-348

<sup>1</sup>/ With Sahli hemoglobinometer except 2-3D and 24-25D where Evelyn photoelectric colorimeter was used.

# 45. ERYTHROCYTE DIAMETERS<sup>1</sup>: MAMMALS

Animal		Value	Animal		Value
(A)	(B)		(A)	(B)	
1 Man	7.5	59	Hog-deer ( <i>Axis porcinus</i> )	4.7	
2 Anteater, giant ( <i>Myrmecophaga tridactyla</i> )	9.2	80	Hog, domestic ( <i>Sus scrofa</i> )	8.0	
3 Anteater, spiny ( <i>Tachyglossus aculeata</i> ) <sup>2</sup>	6.6	81	Horse ( <i>Equus caballus</i> )	5.5	
4 Ape, Barbary ( <i>Macaca sylvana</i> )	7.6	82	Hyena, striped ( <i>Hyena striata</i> ) <sup>2</sup>	6.8	
5 Ape, black, Celebes ( <i>Cynopithecus niger</i> )	7.1	83	Hyena, large-toothed ( <i>Procyon capensis</i> ) <sup>2</sup>	7.7	
6 Armadillo, six-banded ( <i>Euphractus sexcinctus</i> )	7.5	84	Kangaroo, great gray ( <i>Macropus giganteus</i> )	7.6	
7 Ass ( <i>Equus asinus</i> )	6.4	85	Langur ( <i>Pygathrix aurata</i> ) <sup>2</sup>	7.3	
8 Babirusa ( <i>Babirusa babirusa</i> ) <sup>2</sup>	5.8	86	Lemur, ring-tailed ( <i>Lemur catta</i> )	6.5	
9 Baboon ( <i>Papio doguera</i> ) <sup>2</sup>	7.4	87	Leopard, African ( <i>Felis pardus</i> ) <sup>2</sup>	5.9	
10 Badger, honey ( <i>Mellivora capensis</i> )	8.6	88	Lion, African ( <i>Felis leo</i> )	5.6	
11 Badger, European ( <i>Meles meles</i> ) <sup>2</sup>	6.4	89	Loris, slender ( <i>Loris tardigradus</i> ) <sup>2</sup>	7.3	
12 Badger, sand ( <i>Arctonyx collaris</i> ) <sup>2</sup>	7.0	90	Marmoset ( <i>Callithrix jacchus</i> ) <sup>2</sup>	7.0	
13 Bandicoot, rabbit- ( <i>Thylacynus lagotis</i> ) <sup>2</sup>	6.5	91	Marmot, hoary ( <i>Marmota caligata</i> ) <sup>2</sup>	7.3	
14 Bat, big-eared ( <i>Plecotus auritus</i> )	5.7	92	Mole, European ( <i>Talpa europaea</i> )	5.4	
15 Bat, frosted ( <i>Vespertilio murinus</i> ) <sup>2</sup>	6.1	93	Mongoose, gray ( <i>Herpestes ichneumon</i> ) <sup>2</sup>	5.4	
16 Bat, noctule ( <i>Nyctalus noctula</i> ) <sup>2</sup>	5.8	94	Monkey, capuchin ( <i>Cebus capucinus</i> )	7.3	
17 Bat, Roussette ( <i>Roussetus leachi</i> ) <sup>2</sup>	6.5	95	Monkey, leaf ( <i>Presbytis pyrus</i> ) <sup>2</sup>	7.2	
18 Bear, black ( <i>Ursus americanus</i> )	6.9	96	Monkey, red ( <i>Erythrocebus patas</i> ) <sup>2</sup>	7.5	
19 Bear, European ( <i>Ursus arctos</i> )	6.8	97	Monkey, rhesus ( <i>Macaca mulatta</i> ) <sup>2</sup>	7.4	
	7.2	98	Monkey, spider, black ( <i>Ateles paniscus</i> ) <sup>2</sup>	7.1	
	6.6	99	Monkey, squirrel ( <i>Saimiri sciureus</i> ) <sup>2</sup>	6.8	
	7.1	100	Mouse, field, old world ( <i>Apodemus sylvaticus</i> ) <sup>2</sup>	6.6	
	7.5	101	Mouse, house ( <i>Mus musculus</i> )	6.6	
	6.2	102	Nilgai ( <i>Boselaphus tragocamelus</i> ) <sup>2</sup>	5.2	
	6.2	103	Ocelot ( <i>Felis pardalis</i> )	5.5	
37 Civet, s					
38 Coati, s					
39 Cow, dc					
40 Deer, n					
41 Deer, r					
42 Deer, w					
43 Dingo					
44 Dog, do					
45 Elephar					
46 Elephar					
47 Elk (Ce					
48 Fox, bl					
49 Fox, fl					
50 Fox, re					
51 Gayal					
52 Gibbon					
53 Giraffe					
54 Gnu, wt					
55 Goat, d					
56 Guinea					
57 Hare, s					
58 Hippo					



# 46. BLOOD ERYTHROCYTE VALUES IN FETUS, NEWBORN AND ADULT FEMALE: MAMMALS<sup>1</sup>

R B C Count (millions/mm<sup>3</sup> blood)

Stage of Development	Man			Cat			Cow			Goat			Pig			Rabbit			Rat			Sheep		
	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit
1 Newborn	1.1	0.3-2.3		3.9	3.7-4.1		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5	
2 Fetus at term	2.8	2.3-3.5		4.8	4.3-5.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3	
3 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
4 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
5 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
6 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
7 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
8 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
9 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
10 Newborn	4.8	3.8-6.0		5.7	5.1-6.4		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
11 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
12 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	

Hematocrit (ml/100ml blood)

Stage of Development	Man			Cat			Cow			Goat			Pig			Rabbit			Rat			Sheep		
	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit
1 Newborn	1.1	0.3-2.3		3.9	3.7-4.1		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5	
2 Fetus at term	2.8	2.3-3.5		4.8	4.3-5.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3	
3 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
4 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
5 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
6 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
7 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
8 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
9 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
10 Newborn	4.8	3.8-6.0		5.7	5.1-6.4		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
11 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
12 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	

Blood Hemoglobin Concentration (g/100ml blood)

Stage of Development	Man			Cat			Cow			Goat			Pig			Rabbit			Rat			Sheep		
	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit	Value	Range	Unit
1 Newborn	1.1	0.3-2.3		3.9	3.7-4.1		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5		0.25	0.1-0.5	
2 Fetus at term	2.8	2.3-3.5		4.8	4.3-5.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3		2.5	0.2-1.3	
3 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
4 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
5 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
6 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
7 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
8 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
9 Fetus at term	3.5	2.3-4.1		5.3	4.8-5.8		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0		3.0	2.0-4.0	
10 Newborn	4.8	3.8-6.0		5.7	5.1-6.4		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
11 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	
12 Adult female	4.8	4.3-5.1		6.6	6.0-7.0		6.5	6.0-7.0		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4		5.7	5.1-6.4	

1/ Values quoted for adult female are not necessarily those of the mother. Additional values for man may be found in the table on Erythrocyte Values in Frequency and Postpartum. 2/ Gestation period for man in days is 280, cat, 60, goat, 147, pig, 118, rabbit, 31, rat, 21; sheep 147. 3/ Cord or venous blood. 4/ Age, 3-12 days. 5/ Probably by caesarian section. 6/ Caesarian section. 7/ Age, 1-10 days. 8/ Age, 1-10 days. 9/ Age, 2-18 hrs. 10/ Age 24-48 hrs. 11/ Age, 1-3 days. 12/ Age, 8-14 days.

# 46. BLOOD ERYTHROCYTE VALUES IN FETUS, NEWBORN AND ADULT FEMALE MAMMALS (Concluded)

Corpuscle Hemoglobin Content (μg)

Stage of Development	Man		Cat		Cow		Goat		Pig		Rabbit		Rat		Sheep	
	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)
37 Fetus at term <sup>2</sup>	53	47-57C			21.3	20.5-22.0C			56	38-87					40.5	
38 Fetus at term <sup>2</sup>	40	35-45C			21.1	20.6-21.6C			27	19-40					20.5	
39 Fetus at term <sup>2</sup>	42	38-51C			18.6	17.5-20.2C			23	17-29						
40 Fetus at term <sup>2</sup>	40	35-45C	36	24-38C	17.5	17.4-17.6C			27	21-35	44	35-46C				
41 Fetus at term <sup>2</sup>	39	33-45C	28	24-38C	16.6	16.5-16.6C			25	19-31	35	27-48C			13.1	
42 Fetus at term <sup>2</sup>	42	37-47C	27	23-30C	15.0	14.3-16.2C			22	20-28	36	31-47C			12.8	
43 Fetus at term <sup>2</sup>	43	38-48C			13.9	13.4-14.3C			22.5	22-23	28	26-30C			12.8	
44 Fetus at term <sup>2</sup>	44	39-49C			14.1	13.7-14.5C										
45 Fetus at term <sup>2</sup>	45	40-50C			14.1	13.7-14.5C										
46 Newborn	37.5	32-43C	23.4	18-26C					21.7	20-23C	30.9	27-37C	43.11	31-50.11		
47 Newborn	47	42-52C							20.6	16-25C	33.10	27-34C	41.12	24-56.12		
48 Adult female	29	24-33C	18		13.7	12.3-15.3C			21		31		20			13

Corpuscle Volume (μ<sup>3</sup>)

Stage of Development	Man		Cat		Cow		Goat		Pig		Rabbit		Rat		Sheep	
	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)	Value (10)	Range (10)
49 Fetus at term <sup>2</sup>	181	134-235C			83	81-90C			244	204-361					151	119-184C
50 Fetus at term <sup>2</sup>	50	45-55C			80	74-86C			173	131-278						
51 Fetus at term <sup>2</sup>	131	113-150C			77	74-80C			100	80-149					73	
52 Fetus at term <sup>2</sup>	129	116-146C			74	70-78C			94	84-114	120	113-133C				
53 Fetus at term <sup>2</sup>	123	118-136C	134	94-103C	71	69-73C			101	85-112	122	108-136C				
54 Fetus at term <sup>2</sup>	123	118-136C	134	94-103C	71	69-73C			101	85-112	122	108-136C				
55 Fetus at term <sup>2</sup>	123	118-136C	134	94-103C	71	69-73C			101	85-112	122	108-136C				
56 Fetus at term <sup>2</sup>	123	118-136C	134	94-103C	71	69-73C			101	85-112	122	108-136C				
57 Fetus at term <sup>2</sup>	123	118-136C	134	94-103C	71	69-73C			101	85-112	122	108-136C				
58 Newborn	113	90-124C	68.4	63-78C	53.5	50-57C			18.7	16-21C	84.9	80-100C	126.11	98-172.11		
59 Newborn	113	90-124C	68.4	63-78C	53.5	50-57C			18.7	16-21C	84.9	80-100C	126.11	98-172.11		
60 Adult female	87	74-98C	31		50	47-54C			64.8	55-68C	87.10	85-102C	107.12	77-110.12		

1/1 Values quoted for adult female are not necessarily those of the mother. Additional values for man may be found in the table on Erythrocyte Values in Pregnancy and Postpartum Man. 2/2 Gestation period for man in days is 280; cat, 60; goat, 147; pig, 114; rabbit, 31; rat, 21; sheep 147. 3/3 Age: 1-12 days. 4/4 Age: 3-12 days. 5/5 Probably by caesarian section. 6/6 Age: 1-12 hrs. 7/7 Age: 1-12 hrs. 8/8 Age: 2-18 hrs. 9/9 Age: 2-18 hrs. 10/10 Age: 24-48 hrs. 11/11 Age: 1-7 days. 12/12 Age: 8-14 days.

# 47. ERYTHROCYTE OXYGEN CONSUMPTION<sup>1</sup>

47. ERYTHROCYTES		Blood (B); Cells (C)		Temperature of Measurement (°C)	Method
Animal	$\mu\text{l O}_2/\text{mg dry wt/hr.}$				
	In Serum	In Ringer or Saline			
(A)	(B)	(C)	(D)	(E)	
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26	Man	B <sup>4</sup>	-0.0152	37	Chemical
		B <sup>4</sup>	-0.0172	37	
		C <sup>5</sup>	-0.0422	37	Manometric
		C <sup>6</sup>	-0.062	37	
Alligator, American	C <sup>4</sup>	-0.0673	25	Manometric	
	B <sup>4</sup>	-0.212	38		
	B <sup>4</sup>	-0.35	25	Chemical	
	B <sup>4</sup>	-0.26	37		
Chicken	C <sup>6</sup>	-0.182	20	Manometric	
	C <sup>6</sup>	-0.227	20		
	C <sup>6</sup>	-0.0753	20	Manometric	
	C <sup>6</sup>	-0.112	25		
Fish, puffer	C <sup>6</sup>	-0.0513	37	Manometric	
	C <sup>6</sup>	-0.111	37		
	Fish, sea robin	C <sup>6</sup>	-0.402	39	Chemical
	Fish, toad	B <sup>4</sup>	-0.72	25	
Frog, bull	B <sup>4</sup>	-0.67	38	Manometric	
	B <sup>4</sup>	-0.25	37		
	Goose	B <sup>4</sup>	-0.064	37	Chemical
	B <sup>4</sup>	-0.062	37		
Rabbit	B <sup>4</sup>	-0.0492	25	Manometric	
	B <sup>4</sup>	-0.0242	37		
	B <sup>4</sup>	-0.0813	25	Manometric	
	C <sup>7</sup>	-0.0833	25		
Snake, garter	C <sup>6</sup>	-0.062	25	Manometric	
	C <sup>6</sup>	-0.0673	25		
	C <sup>6</sup>	-0.0813	25		
	C <sup>6</sup>	-0.0753	25		
Snake, water	C <sup>6</sup>	-0.096	25	Manometric	
	C <sup>6</sup>	-0.158	25		
	C <sup>6</sup>	-0.119	25		
	C <sup>6</sup>	-0.119	25		

These signs are a convention to indicate loss of the quantity of oxygen in contact with the red corpuscles. All values calculated using 70% by weight. /2/ Ringer.

buffy coat removed.

/1/ The minus signs are a convention to indicate loss of the quantity of oxygen from the atmosphere in contact with the red corpuscles. All values calculated on the basis of water content of cells being 70% by weight. /2/ Ringer. /3/ Isotonic saline. /4/ Defibrinated. /5/ Washed, buffy coat removed. /6/ Washed cells of whole blood. /7/ Suspension of R B C.

# 48. ERYTHROCYTE CARBOHYDRATE METABOLISM

Animal	Substrate <sup>1</sup>	Rate of Lactic Acid Formation <sup>2</sup>		Rate of Methemoglobin Reduction <sup>3</sup>	Enzymes Probably Involved in Utilization <sup>4</sup>
		Value	Range		
(A)	(B)	(C)	(D)	(E)	(F)
1	-	-2.6	-6.1 to 0.8b		
2	se	25.8	12.1-39.5b	0.06	
3	se	24.5	1.8-47.2b	0.77	
4	se	19.6	4.7-34.5b	0.79	
5	se	-0.5	-4.0 to 3.0b	0.47	Glycolytic enzymes <sup>6</sup>
6	se	4.3	-2.8 to 11.4b	0.15	Unknown
7	se	8.9	1.1-16.7b	1.07	Malic dehydrogenase, oxaloacetic acid decarboxylase
8	se	3.8	-0.1 to 7.1b	1.12	Malic dehydrogenase, oxaloacetic acid decarboxylase
9	se	21.0	13.9-28.1b	1.15	Lactic dehydrogenase
10	se	20.5	11.1-29.5b	0.78	
11	se	22.0	14.2-29.6b	0.86	
12	se	1.2	-0.6 to 3.0b	0.69	
13	se	17.4	4.5-30.3b	0.26	Glycolytic enzymes <sup>6</sup>
14	se	33.8	19.1-48.5b	1.19	Unknown
15	se	3.5	-0.4 to 7.4b	1.20	Malic dehydrogenase, oxaloacetic acid decarboxylase
16	se	48.7	28.3-69.1b	1.27	Malic dehydrogenase, oxaloacetic acid decarboxylase
17	se	59.3	36.4-82.2b	0.46	Lactic dehydrogenase
18	se	56.8	32.7-80.9b	2.92	
19	se	13.5	7.4-19.6b	3.07	Glycolytic enzymes <sup>6</sup>
20	se	38.3	24.2-52.4b	2.80	
21	Rabb	44.5	29.4-59.6b	1.21	Unknown
22	Rabb			2.08	Malic dehydrogenase, oxaloacetic acid decarboxylase
23	Rabb			1.82	Malic dehydrogenase, oxaloacetic acid decarboxylase
24	Rabb			5.92	Lactic dehydrogenase

/1/ RBC is not able to utilize succinate. /2/ CO<sub>2</sub>/hour/4ml of a 50% RBC suspension in Krebs-Ringer bicarbonate buffer under anaerobic conditions. /3/ gm of reduced hb/hour (100ml of a 50% suspension of methemoglobin corpuscles in cat and dog cells but not for the hexoses in rabbit cells and for the metabolic intermediates in cat, dog, and rabbit cells except when expressed as the log of the methemoglobin concentration. /4/ Exclusive of methemoglobin reductase and of the pyridine nucleotide coenzymes which are probably utilized in methemoglobin reduction. /5/ Free or endogenous control. /6/ And possibly zwischentferment.

# 49. BLOOD LEUKOCYTE VALUES, BIRTH TO MATURITY: MAN<sup>1</sup>

Age	Leukocytes Total <sup>2</sup>		Neutrophils Total		Neutrophils Band Forms <sup>3</sup>		Neutrophils Segmented Forms		Eosinophils		Basophils		Lymphocytes		Monocytes	
	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>
	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)
Thousands/mm <sup>3</sup> —% of total leukocytes <sup>4</sup>																
1 At Birth	18.1 100	9.0-30.0	11.0 61	6.0-28.0	1.63 9.1		9.4 52	0.40 2.2	0.02-0.85	0.10 0.6	0.10 0.6	0-0.64	5.5 31	2.0-11.0	1.05 5.8	0.40-3.17
2 End 12 hours <sup>7</sup>	22.8 100	13.0-36.0	15.5 68	6.0-28.0	2.33 10.2		13.2 58	0.45 2.0	0.02-0.03	0.10 0.4	0.10 0.4	0-0.50	5.5 24	2.0-11.0	1.20 5.3	0.40-3.67
3 End 24 hours <sup>7</sup>	18.9 100	9.4-34.0 <sup>6</sup>	11.5 61	5.0-21.0	1.75 9.2		9.8 52	0.45 2.4	0.03-1.00	0.10 0.5	0.10 0.5	0-0.30	5.8 31	2.0-11.5	1.10 5.8	0.20-3.1 <sup>7</sup>
4 End 1 week <sup>7</sup>	12.2 100	5.0-21.0	5.5 45	1.5-10.0	0.83 6.8		4.7 39	0.30 4.1	0.07-1.10	0.05 0.4	0.05 0.4	0-0.25	5.0 41	2.0-17.0	1.10 9.1	0.30-2.7 <sup>7</sup>
5 End 2 weeks <sup>7</sup>	11.4 100	5.0-20.0	4.5 40	1.0-9.5	0.63 5.5		3.9 34	0.35 3.1	0.07-1.00	0.05 0.4	0.05 0.4	0-0.23	5.5 48	2.0-17.0	1.00 8.8	0.20-2.4 <sup>7</sup>
6 End 4 weeks <sup>7</sup>	10.8 100	5.0-19.5	3.8 35	1.0-9.0	0.49 4.5		3.5 30	0.30 2.8	0.07-0.30	0.03 0.5	0.03 0.5	0-0.20	6.0 58	2.5-18.5	0.70 6.5	0.15-2.0 <sup>7</sup>
7 End 2 months <sup>7</sup>	11.0 100	5.5-18.0	3.8 34	1.0-9.0	0.49 4.4		3.3 30	0.30 2.7	0.07-0.85	0.05 0.5	0.05 0.5	0-0.20	6.3 57	3.0-16.0	0.85 5.8	0.13-1.3 <sup>7</sup>
8 End 4 months <sup>7</sup>	11.5 100	6.0-17.5	3.8 33	1.0-9.0	0.45 3.9		3.3 29	0.30 2.6	0.07-0.80	0.05 0.4	0.05 0.4	0-0.20	6.8 59	3.5-14.5	0.80 5.2	0.10-1.5 <sup>7</sup>
9 End 6 months <sup>7</sup>	11.9 100	6.0-17.5	3.8 32	1.0-8.5	0.45 3.6		3.3 28	0.30 2.5	0.07-0.75	0.05 0.4	0.05 0.4	0-0.20	7.3 61	4.0-13.5	0.58 4.8	0.10-1.3 <sup>7</sup>
10 End 8 months <sup>7</sup>	12.2 100	6.0-17.5	3.7 30	1.0-8.5	0.41 3.3		3.3 27	0.30 2.5	0.07-0.70	0.05 0.4	0.05 0.4	0-0.20	7.6 82	4.5-12.5	0.58 4.7	0.06-1.2 <sup>7</sup>
11 End 10 months <sup>7</sup>	12.0 100	6.0-17.5	3.6 30	1.0-8.5	0.40 3.2		3.2 27	0.30 2.5	0.06-0.70	0.05 0.4	0.05 0.4	0-0.20	7.5 83	4.5-11.5	0.55 4.6	0.03-1.2 <sup>7</sup>
12 End 12 months <sup>7</sup>	11.6 100	6.0-17.5	3.5 31	1.5-8.5	0.35 3.1		3.2 28	0.30 2.5	0.05-0.70	0.05 0.4	0.05 0.4	0-0.20	7.0 61	4.0-10.5	0.55 4.8	0.05-1.1 <sup>7</sup>

1. The data are from smoothed curves plotted from averages of means from the literature. Apparent discrepancies in addition are due to rounding off after calculations were made. 2. Values in Bar are sums of values in D, J, N, P. 3. Includes a small percentage of myelocytes during first several days after birth. 4. The upper of the two numbers for each age represents thousands per mm<sup>3</sup>, the lower, the percentage of cells of the given type relative to the total leukocyte count. 5. Ranges are from smoothed curves plotted from averages of ranges from the literature. Range values are estimates of the 95% range, and may be regarded as conforming to type "d", i.e., common experience. 6. The range of counts and the range of percentages for any given cell type and age are not necessarily related. 7. Approximately 3 nucleated red cells per 100 leukocytes have been found at birth. 8. An extreme range of 3,600-45,000 has been observed.

# 49. BLOOD LEUKOCYTE VALUES, BIRTH TO MATURITY: MAN (Concluded)

Age		Leukocytes Total <sup>2</sup>		Neutrophils Total		Neutrophils Band Forms <sup>3</sup>		Neutrophils Segmented Forms <sup>3</sup>		Eosinophils		Basophils		Lymphocytes		Monocytes	
		Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>	Value	Range <sup>5</sup>
(A)		(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)
23	End 2 years	10.6	6.0-17.0	3.5	1.5-8.5	0.32		3.2		0.28	0.04-0.65	0.05	0-0.20	6.3	3.0-9.5	0.53	0.05-1.07
26		100		33		3.0		30		2.6		0.5		59		5.0	
27	End 4 years	9.1	5.5-15.5	3.8	1.5-8.5	0.27	0-1.0	3.5	1.5-7.5	0.25	0.02-0.65	0.05	0-0.20	4.5	2.0-8.0	0.45	0-0.8
28		100		42		3.0	0-1.0	39	16-60	2.8		0.6		50		5.0	
29	End 6 years	8.5	5.0-14.5	4.3	1.5-8.0	0.25	0-1.0	4.0	1.5-7.0	0.23	0-0.65	0.05	0-0.20	3.5	1.5-7.0	0.40	0-0.8
30		100		51		3.0	0-1.0	48	16-60	2.7		0.6		42		4.7	
31	End 8 years	8.3	4.5-13.5	4.4	1.5-8.0	0.25	0-1.0	4.1	1.5-7.0	0.20	0-0.60	0.05	0-0.20	3.3	1.5-6.8	0.35	0-0.8
32		100		53		3.0	0-1.0	50	18-60	2.4		0.6		39		4.2	
33	End 10 years	8.1	4.5-13.5	4.4	1.8-8.0	0.24	0-1.0	4.2	1.8-7.0	0.20	0-0.60	0.04	0-0.20	3.1	1.5-6.5	0.35	0-0.8
34		100		54		3.0	0-1.0	51	16-60	2.4		0.5		38		4.3	
35	End 12 years	8.0	4.5-13.5	4.4	1.8-8.0	0.24	0-1.0	4.2	1.8-7.0	0.20	0-0.55	0.04	0-0.20	3.0	1.2-6.0	0.35	0-0.8
36		100		55		3.0	0-1.0	52	16-60	2.5		0.5		38		4.4	
37	End 14 years	7.9	4.5-13.0	4.4	1.8-8.0	0.24	0-1.0	4.2	1.8-7.0	0.20	0-0.50	0.04	0-0.20	2.9	1.2-5.8	0.38	0-0.8
38		100		56		3.0	0-1.0	53	18-60	2.5		0.5		37		4.7	
39	End 16 years	7.8	4.5-13.0	4.4	1.8-8.0	0.23		4.2		0.20	0-0.50	0.04	0-0.20	2.8	1.2-5.2	0.40	0-0.8
40		100		57		3.0		54	25-70	2.6		0.5		35		5.1	
41	End 18 years	7.7	4.5-12.5	4.4	1.8-7.7	0.23		4.2		0.20	0-0.45	0.04	0-0.20	2.7	1.0-5.0	0.40	0-0.8
42		100		57		3.0		54	25-70	2.6		0.5		35		5.2	
43	End 20 years	7.5	4.5-11.5	4.4	1.8-7.7	0.23	0-0.7	4.2	1.8-7.0	0.20	0-0.45	0.04	0-0.20	2.5	1.0-4.8	0.38	0-0.8
44		100		59		3.0	0-6	56	37-75	2.7		0.5		33		5.0	
45	End 21 years	7.4	4.5-11.0	4.4	1.8-7.7	0.22	0-0.7	4.2	1.8-7.0	0.20	0-0.45	0.04	0-0.20	2.5	1.0-4.8	0.30	0-0.8
46		100		59		3.0	0-5	56	37-75	2.7		0.5		34		4.0	

# 50. BLOOD LEUKOCYTE VALUES AT ALTITUDE: MAN

Cell Type	At Sea Level <sup>1</sup>			At Altitude of 3,730 Meters <sup>2</sup>			At Altitude of 4,500 Meters <sup>3</sup>		
	Value		Range	Value		Range	Value		Range
	(B)	(C)		(D)	(E)		(F)	(G)	
(A)									
1 Leukocytes, total, (per mm <sup>3</sup> b1)	6,800	3,480-14,840		6,500	3,400-9,600		6,900	4,700-10,900	
2 Neutrophils, Segmented, %	55.1	38-70		52.8	32-79		49.9	25-71	
3 Neutrophils, Band, %	2.6	0-6		5.0	0-10		3.0	0-5	
4 Eosinophils, %	4.2	0-15		2.5	0-3		2.2	0-2	
5 Basophils, %	0.6	0-2		0.1	11-54		0.2	21-62	
6 Lymphocytes, %	29.8	18-49		34.8	0-9		39.4	1-13	
7 Monocytes, %	7.2	2-14.5		4.9			5.5		

/1/ Lima, Peru. /2/ Oroya, Peru. /3/ Morococha, Peru.

# 51. BLOOD LEUKOCYTE VALUES IN PREGNANCY AND POSTPARTUM: MAN

Cell Type	2nd-3rd month		4th-5th month		6th-7th month		8th-9th month		5th-9th week postpartum	
	Value		Value		Value		Value		Value	
	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)
(A)										
1 Leukocytes, total	10,100	10,600	10,600	10,600	10,400	10,400	10,400	10,400	8,200	8,200
2 Leukocytes, range	5,000-15,000	5,500-15,500	5,500-15,500	5,500-15,500	5,500-15,500	5,500-15,500	5,500-15,500	5,500-15,500	4,500-11,900	4,500-11,900
3 Neutrophils, %	62.5	67.0	67.0	69.5	69.0	69.0	69.0	69.0	51.5	51.5
4 Eosinophils, %	2.0	2.0	2.0	1.5	1.5	1.5	1.5	1.5	0.3	0.3
5 Basophils, %	0.2	0.2	0.2	0.2	0.2	0.2	0.2	0.2	41.0	41.0
6 Lymphocytes, %	31.0	27.0	27.0	25.0	25.0	25.0	25.0	25.0	4.0	4.0
7 Monocytes, %	4.5	4.0	4.0	4.0	4.0	4.0	4.0	4.0		

/1/ Venous blood. For values in non-pregnant woman, see Table 49.

# 52. BLOOD LEUKOCYTE VALUES<sup>1</sup>; CERTAIN LABORATORY ANIMALS

Cell Types	Count per mm <sup>3</sup>			% of Total			Count per mm <sup>3</sup>			% of Total		
	Value (B)	Range (C)	Cat	Value (D)	Range (E)	Rat	Value (F)	Range (G)	Value (H)	Range (I)		
(A)	Cat						Mouse					
1 Total Leukocytes	16,000	9,000-24,000 <sup>c</sup>	(100%)	59.5%	44-82% <sup>c</sup>	14,000	5,000-25,000 <sup>c</sup>	(100%)	22%	9-34% <sup>c</sup>		
2 Neutrophils	9,500	5,500-16,500 <sup>c</sup>	59.5%	5.4	2-11 <sup>c</sup>	3,100	1,100-6,000 <sup>c</sup>	22%	2.2	0-6 <sup>c</sup>		
3 Eosinophils	850	200-2,500 <sup>c</sup>	5.4	0.1	0-0.5 <sup>c</sup>	300	0-700 <sup>c</sup>	2.2	0.5	0-1.5 <sup>c</sup>		
4 Basophils	20	0-100 <sup>c</sup>	0.1	31	15-44 <sup>c</sup>	100	0-200 <sup>c</sup>	0.5	73	65-84 <sup>c</sup>		
5 Lymphocytes	5,000	2,000-9,000 <sup>c</sup>	31	4	0.5-7 <sup>c</sup>	10,200	7,000-16,000 <sup>c</sup>	73	2.3	0-5 <sup>c</sup>		
6 Monocytes	650	50-1,400 <sup>c</sup>	4			300	0-650 <sup>c</sup>	2.3				
	Dog						Guinea Pig					
7 Total Leukocytes	12,000	8,000-18,000 <sup>c</sup>	(100%)	68%	62-80% <sup>c</sup>	8,000	4,000-12,000 <sup>c</sup>	(100%)	25.5%	12-44% <sup>c</sup>		
8 Neutrophils	8,200	6,000-12,500 <sup>c</sup>	68%	5.1	2-14 <sup>c</sup>	2,000	700-4,000 <sup>c</sup>	25.5%	2	0-5 <sup>c</sup>		
9 Eosinophils	600	200-2,000 <sup>c</sup>	5.1	0.7	0-2 <sup>c</sup>	150	0-500 <sup>c</sup>	2	0.5	0-1 <sup>c</sup>		
10 Basophils	85	0-300 <sup>c</sup>	0.7	21	10-28 <sup>c</sup>	50	0-100 <sup>c</sup>	0.5	68	54-85 <sup>c</sup>		
11 Lymphocytes	2,500	900-4,500 <sup>c</sup>	21	5.2	3-9 <sup>c</sup>	5,500	3,000-8,500 <sup>c</sup>	68	4	0-15 <sup>c</sup>		
12 Monocytes	650	300-1,500 <sup>c</sup>	5.2			300	0-1,300 <sup>c</sup>	4				
	Rabbit						Guinea Pig					
13 Total Leukocytes	9,000	6,000-13,000 <sup>c</sup>	(100%)	46%	36-52% <sup>c</sup>	10,000	7,000-19,000 <sup>c</sup>	(100%)	42%	22-50% <sup>c</sup>		
14 Neutrophils <sup>2</sup>	4,100	2,500-6,000 <sup>c</sup>	46%	2	0.5-3.5 <sup>c</sup>	4,200	2,000-7,000 <sup>c</sup>	42%	4	2-12 <sup>c</sup>		
15 Eosinophils	180	0-400 <sup>c</sup>	2	5	2-7 <sup>c</sup>	400	200-1,300 <sup>c</sup>	4	0.7	0-2 <sup>c</sup>		
16 Basophils	450	150-750 <sup>c</sup>	5	39	30-52 <sup>c</sup>	70	0-300 <sup>c</sup>	0.7	49	37-64 <sup>c</sup>		
17 Lymphocytes	3,500	2,000-5,600 <sup>c</sup>	39	8	4-12 <sup>c</sup>	4,900	3,000-9,000 <sup>c</sup>	49	4.3	3-13 <sup>c</sup>		
18 Monocytes	725	300-1,300 <sup>c</sup>	8			430	250-2,000 <sup>c</sup>	4.3				

<sup>1</sup>/ Values and ranges in this table are approximate averages of highly variable unweighted means and ranges from the literature. <sup>2</sup>/ In rabbits and guinea pigs, cells classed as "neutrophils" include "pseudo-eosinophils," "amphophils," or "heterophils."



## 50. BLOOD LEUKOCYTE VALUES AT ALTITUDE: MAN

Cell Type	At Sea Level <sup>1</sup>		At Altitude of 3,730 Meters <sup>2</sup>		At Altitude of 4,500 Meters <sup>3</sup>	
	(C)		(E)		(G)	
	Value	Range	Value	Range	Value	Range
(A)	(B)	(C)	(D)	(E)	(F)	(G)
1 Leukocytes, total, (per mm <sup>3</sup> b1)	6,800	3,480-14,840	6,500	3,400-9,600	6,900	4,700-10,900
2 Leukocytes, Segmented, %	55.1	38-70	52.8	32-79	49.9	25-71
3 Neutrophils, Band, %	2.6	0-6	5.0	0-10	3.0	0-5
4 Eosinophils, %	4.2	0-15	2.5	0-3	2.2	0-2
5 Basophils, %	0.6	0-2	0.1	11-54	0.2	21-62
6 Lymphocytes, %	29.8	18-49	34.8	0-9	39.4	1-13
7 Monocytes, %	7.2	2-14.5	4.9		5.5	

/1/ Lima, Peru /2/ Oroya, Peru. /3/ Morococha, Peru.

## 51. BLOOD LEUKOCYTE VALUES IN PREGNANCY AND POSTPARTUM: MAN

Cell Type	2nd-3rd month		4th-5th month		6th-7th month		8th-9th month		5th-9th week postpartum	
	(B)		(C)		(D)		(E)		(F)	
	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)
1 Leukocytes, total	10,100	5,000-15,000	10,600	5,500-15,500	10,600	5,500-15,500	10,400	5,500-15,500	8,200	4,500-11,900
2 Leukocytes, range	62.5		67.0		69.5		69.0		51.5	
3 Neutrophils, %	2.0		2.0		1.5		1.5		3.0	
4 Eosinophils, %	0.2		0.2		0.2		0.2		0.3	
5 Basophils, %	31.0		27.0		25.0		25.5		41.0	
6 Lymphocytes, %	4.5		4.0		4.0		4.0		4.0	
7 Monocytes, %										

/1/ Venous blood. For values in non-pregnant woman, see Table 49.

# 52. BLOOD LEUKOCYTE VALUES<sup>1</sup>: CERTAIN LABORATORY ANIMALS

Cell Types	Count per mm <sup>3</sup>			% of Total			Count per mm <sup>3</sup>			% of Total		
	Value (B)	Range (C)	Value (D)	Range (E)	Value (F)	Range (G)	Value (H)	Range (I)				
(A)	Cat						Rat					
Total Leukocytes	16,000	9,000-24,000 <sup>c</sup>	(100%)	44-82% <sup>c</sup>	14,000	5,000-25,000 <sup>c</sup>	(100%)	9-34% <sup>c</sup>				
1 Neutrophils	9,500	5,500-16,500 <sup>c</sup>	59.5%	2-11 <sup>c</sup>	3,100	1,100-6,000 <sup>c</sup>	22%	0-6 <sup>c</sup>				
2 Eosinophils	850	200-2,500 <sup>c</sup>	5.4	0-0.5 <sup>c</sup>	300	0-700 <sup>c</sup>	2.2	0-1.5 <sup>c</sup>				
3 Basophils	20	0-100 <sup>c</sup>	0.1	15-44 <sup>c</sup>	100	0-200 <sup>c</sup>	0.5	65-84 <sup>c</sup>				
4 Lymphocytes	5,000	2,000-9,000 <sup>c</sup>	31	0.5-7 <sup>c</sup>	10,200	7,000-16,000 <sup>c</sup>	73	0-5 <sup>c</sup>				
5 Monocytes	650	50-1,400 <sup>c</sup>	4		300	0-650 <sup>c</sup>	2.3					
	Dog						Mouse					
Total Leukocytes	12,000	8,000-18,000 <sup>c</sup>	(100%)	62-80% <sup>c</sup>	8,000	4,000-12,000 <sup>c</sup>	(100%)	12-44% <sup>c</sup>				
1 Neutrophils	8,200	6,000-12,500 <sup>c</sup>	68%	2-14 <sup>c</sup>	2,000	700-4,000 <sup>c</sup>	25.5%	0-5 <sup>c</sup>				
2 Eosinophils	600	200-2,000 <sup>c</sup>	5.1	0-2 <sup>c</sup>	150	0-500 <sup>c</sup>	2	0-1 <sup>c</sup>				
3 Basophils	85	0-300 <sup>c</sup>	0.7	10-28 <sup>c</sup>	50	0-100 <sup>c</sup>	0.5	54-85 <sup>c</sup>				
4 Lymphocytes	2,500	900-4,500 <sup>c</sup>	21	3-9 <sup>c</sup>	5,500	3,000-8,500 <sup>c</sup>	68	0-15 <sup>c</sup>				
5 Monocytes	650	300-1,500 <sup>c</sup>	5.2		300	0-1,300 <sup>c</sup>	4					
	Rabbit						Guinea Pig					
Total Leukocytes	9,000	6,000-13,000 <sup>c</sup>	(100%)	36-52% <sup>c</sup>	10,000	7,000-19,000 <sup>c</sup>	(100%)	22-50% <sup>c</sup>				
1 Neutrophils <sup>2</sup>	4,100	2,500-6,000 <sup>c</sup>	46%	0.5-3.5 <sup>c</sup>	4,200	2,000-7,000 <sup>c</sup>	42%	2-12 <sup>c</sup>				
2 Eosinophils	180	0-400 <sup>c</sup>	2	2-7 <sup>c</sup>	400	200-1,300 <sup>c</sup>	4	0-2 <sup>c</sup>				
3 Basophils	450	150-750 <sup>c</sup>	5	30-52 <sup>c</sup>	70	0-300 <sup>c</sup>	0.7	37-64 <sup>c</sup>				
4 Lymphocytes	3,500	2,000-5,600 <sup>c</sup>	39	4-12 <sup>c</sup>	4,900	3,000-9,000 <sup>c</sup>	49	3-13 <sup>c</sup>				
5 Monocytes	725	300-1,300 <sup>c</sup>	8		430	250-2,000 <sup>c</sup>	4.3					

/1/ Values and ranges in this table are approximate averages of highly variable unweighted means and ranges from the literature. /2/ In rabbits and guinea pigs, cells classed as "neutrophils" include "pseudo-eosinophils," "amphophils," or "heterophils."

53. BLOOD LEUKOCYTE RANGES: FARM ANIMALS

53. BLOOD LEUKOCYTE RANGES: FARM ANIMALS							
Animal	Leukocytes Thousands/mm <sup>3</sup> blood	Number/mm <sup>3</sup> blood					Monocytes
		Neutrophils	Eosinophils	Basophils	Lymphocytes	(G)	
(A)							
1 Cattle	5-12 <sup>d</sup>	1,200-4,800 <sup>d</sup>	180-1,800 <sup>d</sup>	0-100 <sup>d</sup>	2,700-6,900 <sup>d</sup>	150-1,800 <sup>d</sup>	
2 Goat	5-14 <sup>d</sup>	2,100-3,350 <sup>d</sup>	0-1,100 <sup>d</sup>	0-600 <sup>d</sup>	2,100-11,250 <sup>d</sup>	50-600 <sup>d</sup>	
3 Horse	5-11 <sup>d</sup>	3,000-6,900 <sup>d</sup>	50-600 <sup>d</sup>	0-100 <sup>d</sup>	1,200-4,800 <sup>d</sup>	100-1,450 <sup>d</sup>	
4 Sheep	4-10 <sup>d</sup>	1,000-4,500 <sup>d</sup>	50-700 <sup>d</sup>	0-200 <sup>d</sup>	2,500-7,000 <sup>d</sup>	50-800 <sup>d</sup>	
5 Swine	7-20 <sup>d</sup>	2,400-10,000 <sup>d</sup>	50-2,000 <sup>d</sup>	0-800 <sup>d</sup>	3,200-12,000 <sup>d</sup>	50-2,000 <sup>d</sup>	
6 Chicken	16-40 <sup>d</sup>	4,000-16,000 <sup>d</sup>	400-4,000 <sup>d</sup>	200-1,600 <sup>d</sup>	8,000-24,000 <sup>d</sup>	1,000-6,000 <sup>d</sup>	

# 54. MORPHOLOGY OF BLOOD LEUKOCYTES AND PLATELETS: MAN<sup>1</sup>

54. MORPHOLOGY OF BLOOD

NUCLEUS

CYTOPLASM

CYTOPLASM				NUCLEUS						
Cell Type	Size $\mu$	Granules	Relative Amount	Color	Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Parachromatin Pattern and Amount
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)
1 Band cell, Neutrophilic <sup>2</sup> (Regenerative)	10-15	Full complement of specific granules, pink to rose-violet			Rod-like, or deeply indented no segmentation				Thickened chromatin strands form coarse chromatin masses with a few distinct interstices	Small amount; pale blue or pink, seen as distinct clumps between chromatin masses.
2 Granulocytes, segmented <sup>2</sup>		Full complement of specific granules <sup>3</sup>			Polymorpho-nuclear, elongated, constricted					
3 (a) Neutrophilic	9-15	Specific granules are small, fine pink to rose-violet and irregularly distributed throughout cytoplasm	Abundant	Colorless or slightly acidophilic	(a) 2 or more lobes, normal maximum = 5.	Central or eccentric.	Thick, heavy	None visible	(a)(b) Thickened chromatin strands form coarse chromatin masses with a few distinct interstices.	(a)(b) Small amount; small irregular, but distinct clumps between chromatin masses.
4 (b) Eosinophilic		Specific granules are red and throughout cytoplasm			(b) 2 to 4 lobes					
5 (c) Basophilic		Specific granules are metachromatic, water soluble, vary in size, and may overlie nucleus; are irregularly distributed.			(c) Slightly indented or irregular; less lobulation than (a)(b)				(c) Chromatin pattern less distinct than (a)(b); amudged.	(c) Much less distinct than in (a)(b)

ETS: MAN (Concluded)

55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS				
Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions		Biological Properties and Functions (E)
		Nucleus and Cytoplasm (C)	Nucleus (D)	
			Chemical Contents	
1	Physical Properties: Cell Specific gravity=1.066. Migrate towards anode. Cathaphoretic velocity slower than lymphocytes.	Acid phosphatase; Alkaline phosphatase; Amylase. Catalase; Cytochrome oxidase. Fibrinolysin. Histamine. Lipase. Nucleotidase. Phosphorylase. Proteases. Adenosinase. Cathelapsin. Lysosyme. Nuclease. Peptidase. Trypsinase (trypsin). Trypanolase. Verdoperoxidase.	Sulfur <sup>11</sup> .	Possess ameboid and wormlike locomotion (28-34, per minute). Show diapedesis and adhere to endothelium under stimulation. May show marked cytoplasmic budding. Exhibit chemotaxis positive and negative for about 1 mm distance. Retain integrity best in isotonic, slightly hypertonic, or slightly hypotonic solution. Fragility inversely proportional to ascorbic acid content. Phagocytic for bacteria, quite common for blood, tissue cells, debris, antigen-antibody compounds, and foreign bodies. Phagocytic ability proportional to ascorbic acid content. Phagocytosis enhanced by opsonins and bacteriotropins. Phagocytosis enhanced by coagulase. Migrate into areas of inflammation, especially acute. Elaborate soluble, antigenically active material from particulate antigens. Associated with complement formation.
5	Polarized, but may reverse direction. Are sensitive to irradiation; particularly in hemopoietic tissues. Constriction rings due to external factors <sup>1</sup> .			
10	Physical Properties: Granules			
15	Slightly refractile. Little or no electrophoretic movement during active flow during movement of cell.			
20	Chemical Properties			
25	Are very sensitive to moderate pH changes from normality. Are very sensitive to benzene and related compounds; sensitivity may be property of regenerative not of adult forms. Adult forms more resistant to IHN, than regenerative forms. Respond to leukotoxine. Exhibit chemotaxis <sup>1</sup> . Exhibit oxidase effusion. High glycolytic activity. Reduce potassium tellurite.	Sudan black B positive. Acridine orange positive.	Feulgen positive.	Granules: Acid fast. Nile blue sulphate positive. Sudan III positive. Stain red-lilac with Wright's stain. Stain violet with Ehrlich's triacid stain. Sudan black B positive. Acid hematein positive. Neutral red positive. Non-specific tanophil granules.
30				Produce agglutinins <sup>1</sup> . Agglutinated by coagulase. Are source for oxidase effusion. Are source for verdoperoxidase. High oxygen consumption. High glycolytic activity. Digest fibrin. Post mitotic. Extravascular maturation. Life span in blood 0.5 to 5 days. Disintegrate to "pus corpuscles". Disappear quickly in tissue culture explants.
35				
40				

1/1/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. 1/2/Questioned. 1/3/Inhibited by leukocidins; decreased by alcohol. Positive chemotaxis of leukocytes: cellulose, starch, glycogen, glucose, maltose, levulose, fructose, absorbent cotton. No chemotaxis: olive oil, various inorganic salts, citric acid, Na<sub>2</sub> K<sub>2</sub> Mg, and Ca palmate, stearic acid, carbon, quartz, glass, washed MnO<sub>2</sub>, non-absorbent cotton, raw cotton. Negative chemotaxis: aluminum oxide. 1/4/Less than lymphocytes. 1/5/Contain major portion of alkaline phosphatases of white cell suspensions. 1/6/Trace. 1/7/May be a common ester. 1/8/Present in significant amounts only in rabbit neutrophils. 1/9/In cats; in man found in exudates rich in neutrophilic segmented cells. 1/10/Stuccus contains 1.8% sulfur. 1/11/Amount decreases with maturation. 1/12/Extractable with acetone.

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)

Cell Type (A)	Chemical Contents and Stain Reactions				Biological Properties and Functions (F)
	Physical Properties (B)	Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
45	Physical Properties Core of granules offers little resistance to electron.	Amylase. Cytochrome oxidase. Histamine. Lipase. Papainase. Trypsinase. Verdoperoxidase	Chemical Contents Alkaline phosphatase.	Cytoplasm proper: Glycogen (extragranular). Mucopolysaccharide (neutral). Granules: Acid phosphatase. Cytochrome c oxidase. Iron.	Possess amoeboid locomotion, but are usually slower than neutrophils (5-9, per minute). Occasionally travel very rapidly in supravital preparations. Migrate into areas of inflammation. Phagocytic (usually less than neutrophils). Transport anaphylactogenic substance (antigen). Source of blood histamine. Cells are slowly affected by anisotonia ("hardy" cells). Extravascular maturation. Survive 10 days or more in vitro.
50	Chemical Properties Form Charcot-Leyden crystals. Nucleus selectively dissolved with wetting agents			Iron-porphyrin-protein complex. Histamine. Phospholipids. Tyrosine. Verdoperoxidase. Corpora in low molecular weight compounds.	
55	Leukocyte. Eosinophilic (granulocyte, eosinophilic, segmented).		Stain Reactions Feulgen positive.	Granules: Sudan black B: positive for bull. Negative for core. Nile blue sulphate positive. Acid hematein: positive for core. Negative for bull. Neutral red positive. Eosin positive.	
60		Acridine orange positive			
65					
70					
75					

11/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. 12/Questioned. 13/Probably nuclear in origin. Crystals are hexagonal pyramids with bases opposed. 14/Rate depends upon site and surface. 15/Especially indammation due to parasites, repeated foreign protein allergy, epidermal destruction, and tumors; also subacute and chronic inflammations.

55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)					
Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
80			Chemical Contents		
		No acid phosphatase.			
85	Tissue Eosinophil*				
			Chemical Contents		
				Cytoplasm proper: No ribonucleoprotein. No glycogen. Granules: Alkaline phosphatase. No glycogen.	
90			Stain Reactions		
			Feulgen positive		
95				Cytoplasm proper: HIO-Schiff (PAS) positive Sudan III positive Granules: HIO-Schiff (PAS) positive. Sudanophilic Stain green with thionine.	
100			Chemical Contents		
	Physical Properties			Granules: Oxidase. Glycogen. Peroxidase. Lipid Phospholipids.	Possess amoeboid locomotion (slower than neutrophils or eosinophils). Migrate into areas of inflammation. Phagocytic. May produce heparin. Post mitotic. Extravascular maturation.
105	Chemical Properties				
	Granular metachromatic material is soluble in aqueous solutions Positive chemotaxis to egg albumin				
110	Leukocyte, Basophilic (granulocyte, segmented).				
		Sudan III positive Sudan black B positive Acridine orange positive.	Stain Reactions Feulgen positive	Granules: Snow metachromasia. HIO-Schiff (PAS) positive Acid hematein".	

/1/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned. /16/ May be identical with blood eosinophil /17/ Variable. /18/ Selective and rare.



# 53. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)

Cell Type (A)	Physical and Chemical Properties (B)		Chemical Contents and Stain Reactions		Biological Properties and Functions (F)
	Physical Properties	Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
45	Core of granules offers little resistance to electric currents	Amylase, Cytochrome oxidase, Lipase, Papainase Trypsinase, Verobutyrooxidase.	Chemical Contents Alkaline phosphatase.	Cytoplasm proper: Glycogen (extra-granular), Mucopolysaccharide (neutral).	Possess amoeboid locomotion, but are usually slower than neutrophils (5-8 μ per minute). Occasionally travel very rapidly in supravital preparations. Migrate into areas of inflammation. Phagocytic (usually less than neutrophils). Transport anaphylactogenic substances (antigen). Source of blood histamine. Cells are slowly affected by anisotonia ("hardy" cells). Extravascular maturation. Survive 10 days or more in vitro.
50	Chemical Properties Form Charcot-Leyden crystals. Nucleus selectively dissolved with wetting agents			Granules: Acid phosphatase, Cytochrome c oxidase, Iron porphyrin-protein complex. Histamine, Phospholipids. Tyrosine. Verdoperoxidase. Core rich in low molecular weight compounds	
55	Leukocyte, Eosinophilic (granulocyte, eosinophilic, segmented).			Stain Reactions Feulgen positive.	
60		Acridine orange positive		Granules: Sudan III positive. Sudan black B: positive for hull, negative for core. Nile blue sulphate positive. Acid hematein: positive for core, negative for hull. Neutral red positive. Eosin positive.	
65					
70					
75					

1/ Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/ Questioned.  
/3/ Probably nuclear in origin. Crystals are hexagonal pyramids with bases opposed. /4/ Rate depends upon site and surface. /5/ Especially inflammation due to parasites, repeated foreign protein allergy, epidermal destruction, and tumors; also subacute and chronic inflammations.

# 55 PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS\* (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
80			Chemical Contents No acid phosphatase.	Cytoplasmic Contents Cytoplasm proper: No ribonucleoprotein No glycogen Granules: Alkaline phosphatase <sup>1</sup> . No glycogen	
85 Tissue Eosinophil <sup>16</sup>			Stain Reactions Feulgen positive.	Cytoplasm proper: HIO-Schiff (PAS) positive Sudan III positive Granules HIO-Schiff (PAS) positive. Sudanophilic Stain green with thionine	
90					
95					
100	Physical Properties More resistant to irradiation than other granulocytes		Chemical Contents	Granules: Oxidase Glycogen Peroxidase. Lipid Phospholipids	Possess amoeboid locomotion (slower than neutrophils or eosinophils). Migrate into areas of inflammation Phagocytic. May produce heparin. Post mitotic Extravascular maturation.
105 Leukocyte, Basophilic (granulocyte, segmented).	Chemical Properties Granular metachromatic material is soluble in aqueous solutions Positive chemotaxis to egg albumin		Stain Reactions Sudan III positive. Sudan black B positive. Acridine orange positive	Granules: Show metachromasia. HIO-Schiff (PAS) positive Acid hematein <sup>17</sup> .	
110					

<sup>1</sup>/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. <sup>2</sup>/Questioned <sup>16</sup>/ May be identical with blood eosinophil <sup>17</sup>/ Variable <sup>18</sup>/ Selective and rare

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions		Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)
115	Granules more soluble in water than blood basophil granules. Resistant to ribonuclease digestion. Exhibit chemotaxis, especially to egg albumin.		Chemical Contents No acid or alkaline phosphatase.	Cytoplasm proper: Heparin. Cytoplasm and granules: Heparin. No lipase. No veridoperoxidase. No free iron. No glycogen. No nucleic acid. Granules: Acid mucopolysaccharides. Acid phosphatase. Alkaline phosphatase. Cytochrome oxidase. Heparin. Monosulfuric acid mucopolysaccharides. Granules chemically not related to mucin or pigment cell granules. Golgi area: Acid phosphatase.
120				
125				
130	Tissue Mast Cell			
135				
140				
145				
150				

/1/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned. /19/Less soluble in rat and mouse than in man. /20/Changes to di-, tri-, and tetra- in plasma. /21/Metachromasia resistant to hyaluronidase digestion. /22/May be by granular solution and vacuolization.

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS: (Continued)

Cell Type	Chemical Contents and Stain Reactions				Biological Properties and Functions (F)
	Physical and Chemical Properties (B)	Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
(A)	Physical Properties	Chemical Contents			
155	Cataphoretic velocity slightly higher than that of neutrophils and thrombocytes. Very sensitive to irradiation and HN <sub>2</sub> , particularly in hemopoietic tissues <sup>1</sup> .	Adenosinase. Acid phosphatase <sup>2</sup> . Alkaline phosphatase. Amylase <sup>3</sup> . Cathepsin. No cytochrome oxidase (neg in blood). Esterase (low). Gamma globulin <sup>4</sup> . Lipase <sup>5</sup> .	Chromatin complex. Nucleohistone. Desoxyribonucleic acid.	Cytoplasm proper: Ribonucleoprotein (basophilic). Acid hematein negative. Verticillidase (neg. in blood; may be pos. in field of inflammation). Granules: Glycogen <sup>6</sup> . Mitochondria. Nucleoproteins. Phospholipids <sup>7</sup> . Cytochromes (Golgi zone). Acid phosphatase.	Possess polarized ameboid rotary locomotion (15 to 30% per minute). Phagocytic for bacteria, yeast, blood, and tissue cells <sup>8</sup> . Phagocytic into areas of inflammation. Migrate into G I tract. May show marked cytoplasmic budding <sup>9</sup> . Cytoplasmic budding may be concurrent with rise in antibody titre <sup>10</sup> . Cells are relatively resistant in hypotonic solutions. Oxygen consumption less than that of granulocytes. Synthesize protein (globulin) <sup>11</sup> . Participate in antibody formation <sup>12</sup> . Trephocytic. Transform into monocytes <sup>13</sup> , into plasma cells.
165	Are less sensitive than granulocytes to benzene and related compounds. Synthesize protein (globulin) <sup>14</sup> . Aerobic glycolysis: slight or none. Glycolytic activity half or less than that of granulocytes. Do not exhibit chemotaxis <sup>15</sup> .	Lysozyme. Nucleinase. Purine-nucleosidase.			Transform into histiocytes (macrophages) in inflammation and into macrophages, epithelioid cells, giant cells in tissue culture <sup>16</sup> . Form granulocytes <sup>17</sup> . Chromosome size varies with tissue and with age. Develop nuclear vacuoles during degeneration. Collagen deposition may be associated with lymphocyte disintegration. Sensitive to or killed by adrenal cortical extract, HN <sub>2</sub> , or x-rays in vitro. Survive best at 7-17° C. in vitro. Intermitotic. Life span in blood possibly less than 24 hours; in tissues, days or weeks.
175			Stain Reactions		
180			Lack stainable lipoidal substances in blood. Stain with acridine orange.	Cytoplasm proper: Siderophilic. Granules: Iodophilic. Azurophilic granules. Give negative HIO-Schiff (PAS) reaction. Cytochromes. Occasional rosette of neutral red vacuoles near cytocentrum.	
185					
190					

1/1 Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. 2/2 Questioned. 3/3 More sensitive in node than in the rest of the node. 4/4 Evidence of protein synthesis for "export" disputed. 5/5 Contain major portion of acid phosphatase in leukocyte suspensions. 6/6 Small amounts: 1/100 that of neutrophils. 7/7 Highly soluble in alcohol. 28/28 May be first step in disintegration. 29/29 May be due to premeation and segregation into vacuoles instead of phagocytosis. 30/30 Directly or by supplying essential basic building blocks and by stimulating phosphorylation.

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS: (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
195 Leukocytoid Lymphocyte (of infectious mononucleosis)		No peroxidase. No glycogen <sup>1</sup> .			Possess depolarized locomotion <sup>1</sup> . Phagocytic for bacteria <sup>1</sup> .
			Stain Reactions Feulgen positive.	Cytoplasm proper: Methylene blue positive.  Cytocentrum: Rosette of neutral red vacuoles about cytocentrum.	
200					
203	Exhibit chemotaxis for about 25 $\mu$ distance. Often oxidase positive.	No alkaline phosphatase <sup>1</sup> . Carbohydrase. Lipase. Nucleinase. Proteolytic enzymes. Verdoperoxidase.		Chemical Contents Cytoplasm: Glycogen. Granules: Lipids. Phospholipids.	Locomotion is depolarized Phagocytic for bacteria, cellular debris, pigment, and parasites. Phagocytic for vital dyes <sup>1</sup> . Exhibit pinocytosis. May show cytoplasmic budding <sup>1</sup> . Migrate into areas of inflammation. Participate in antibody formation <sup>1</sup> . Transform into macrophages, epithelioid cells, giant cells, and possibly into fibroblasts. Extravascular maturation.
210					
215	Monocyte (blood mononuclears).	Acridine orange positive.	Feulgen positive.	Cytoplasm: Droplets stain with Sudan III <sup>1</sup> . Acid hematein positive. Droplets stain with Nile blue sulfate Cytocentrum: Rosette of neutral red vacuoles <sup>1</sup> . Granules: Sudan III positive. Sudan black B positive.	
220					
223					
230					

<sup>1</sup>/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned. /29/May be due to permeation and segregation into vacuoles instead of phagocytosis. /31/Not specific. /32/May be first step in disintegration. /33/Possibly by elaborating soluble antigenically active material for particulate antigens.

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS: (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
		Chemical Contents			
235	Do not exhibit chemotaxis. Are less sensitive to lower pH than other leukocytes.	Carbohydrase Lipase. Nuclease Proteinases No verticillinase.			on is depolarized. ic for bacteria, cells, tissue, parasites as well as foreign ic for lipids and then called distocytes" or "foam cells" al dyes Inocytosis as large scavenger cells of nation. into, or are activated in, of inflammation. s than other leukocytes", e in antibody formation", d with the intermediate me- n of blood and bile pigments, erol and iron. n into epithelioid cells, for- dy type, or Langerhans type ells, and possibly into fibro- ed animals.
240					
245	Etiocyte (Macro- phage).		Stain Reactions Feulgen positive.	Neutral red vacuoles are dispersed in cytoplasm or form rosette about cyto- centrum". Acid hematein positive.	
250					
255					
260	Megakary- ocyte	Acid phosphatase. Alkaline phosphatase (low)		Chemical Contents Cytoplasm: Ribonucleoprotein. Granular glycogen. Granules: Oxidase	May be phagocytic. Possess slight motility. Form platelets by pseudopodial for- mation and disintegration of cyto- plasm.
265			Stain Reactions	Cytoplasm: Exhibits metachro- masia. Acid hematein posi- tive. Granules: Iodophilic. Mitochondria: Sudan black B positive	
270					

/1/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned.  
/31/Not specific. /33/Possibly by elaborating soluble antigenically active material for particulate antigens.

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions (CDE)	Biological Properties and Functions (F)
	Physical Properties Are negatively charged. Cathaphoretic velocity same as that of neutrophils. Chromomeres traversed by ultra-violet rays. Chemical Properties Chromomere dissolves in acetic acid. Positive Bial's test in acid hydrolysates.	Chemical Contents Acid phosphomonoesterase; Ascorbic acid; Catalase; Dopamine; Histamine; Oxidase; Peptolytic enzymes; No veridoperoxidase; No phosphatase; Granular glycoprotein; Lipids; Phospholipids; Cephalin, Free cholesterol, Cholesterol esters, Neutral fats, Ribonucleic Acid; Thromboplastin; Platelet accelerator factor; Vasoconstrictor substance; Hypotensive principle.  Stain Reactions Granules: Iodophilic Acid hematein positive. Sudan black B positive. Feulgen negative. Neutral red positive bodies.	Release thromboplastin in clotting. Release accessory (accelerator) clotting factors. Interact with plasma antithrombotic factor to form thromboplastin. Possess agglutinative function, are sticky and adhesive, and lend these properties to the clot. Form capillary thrombi in test for bleeding time <sup>a</sup> when more than 30,000-60,000 per mm <sup>3</sup> . Adhere to microorganisms, "platelet loading." Associated with coarsening and shortening of fibrin strands in synecrisis. Liberate a vasoconstrictor substance. Liberate a hypotensive principle which is stronger than vasoconstrictor substance. Support capillary walls in tourniquet tests when more than 30,000-60,000 per mm <sup>3</sup> , and endothelial cells are normal. In shed blood, transform into dendritic, then into expanded forms. In shed blood, contains fibrils which in turn are composed of spherules (50-200 millimicrons) disseminated into surrounding medium. Consists of bits of megakaryocyte cytoplasm. Oxygen consumption of platelets in man is 8.4 mm <sup>3</sup> per mg per hr. Life span in blood is 3-5 days.
275			
280			
285			
290			
295			
300			

<sup>1/1</sup>Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. <sup>2/2</sup>Questioned. <sup>3/3</sup>In rabbits. <sup>4/4</sup>In horses. <sup>5/5</sup>15% of dried weight consists of lipids. <sup>6/6</sup>37/68% of phospholipids is cephalin. <sup>7/7</sup>Critical level in dispute. <sup>8/8</sup>129/

# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS (Continued)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
303	Relatively resistant to ir- radiation HN, and to toxic stimuli of all va- rieties. Tendency to mat and to form colonies in vitro		Stain Reactions	Cytoplasm proper: May have azure gran- ules in smears.  Granules: Hemosiderin gran- ules. Argrophilic. Sudan black B.	Possess pseudopodial migration. Form stromal support for hema- topoietic tissues. Line sinuses of bone marrow, lymph nodes, and spleen. Line sinuses of liver, adrenal cortex, and possibly anterior lobe of pitui- tary gland. Associated with intermediate metabo- lism of blood and bile pigments, cholesterol and iron. Stem cells for myeloblasts, mono- cytes, megakaryoblasts, lympho- blasts, promonoblasts, promegalo- blasts, and plasmablasts. Form osteoblasts and possibly osteo- clasts. May form cartilage cells (chondrocy- tes). Form osteocytes. Transform into adipose cells Form argyrophil (reticular) fibers Possess all functions ascribed to his- tiocyte.
310	Reticulum Cell (Mesenchymal cell).				
315					
320					
325					
330	Relatively resistant to ir- radiation and HN.	Alkaline phos- phatase. No peroxidase.	Chemical Contents	(Cytoplasm proper: Acid phosphatase. Ribonucleoprotein. Mucoprotein. Russell bodies; Peroxidase. Mucoprotein. Ribonucleic acid	Are not phagocytic. May show marked peripheral cyto- plasmic dissolution. Participate in antibody formation after reinjection of antigen. Participate in antibody formation after simple injection of antigens. Secrete Erythrocytes and mucoglo- bulins. May transform into plasma mast cells.
335					
340			Stain Reactions	Neutral red vacuoles may be arranged as rosettes about cyto- centrum Stains red with Unna's stain. Pyronin of methy- green-pyronin positive. Acridine red of mala- chite-green- acridine-red positive Russell bodies stain pink with H & E.	
345					
350					

/1/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned.  
/25/May be first step in disintegration. /40/No longer serve as stem cells except pathologically. /41/Usually.

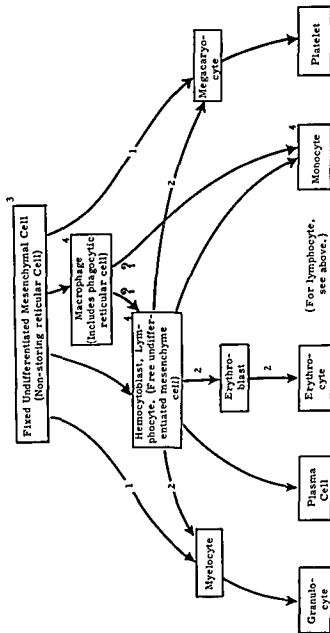


# 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS<sup>1</sup>(Concluded)

Cell Type (A)	Physical and Chemical Properties (B)	Chemical Contents and Stain Reactions			Biological Properties and Functions (F)
		Nucleus and Cytoplasm (C)	Nucleus (D)	Cytoplasm (E)	
333		Vitamin A. Free alpha amino acid groups.		Cytoplasm proper: Fine, granular, glycogen.  Hof: Phospholipids, Alkaline phosphatase negative.	Exhibit frequent cytoplasmic bud- ding. May form Russell bodies. Cells often associated with abnor- mally increased plasma globulins. Cells often associated with abnormal plasma protein formation (Hence- Jones protein). Produce less globulin than mature plasma cells.
380	Myeloma Cell <sup>o</sup>				
383					
370					

<sup>1</sup>/Statement of a positive finding in this table does not necessarily imply a negative finding in the same respect elsewhere in the table. /2/Questioned.  
/28/May be first step in disintegration. /42/Usually considered to be a malignant form of plasmacytic series.

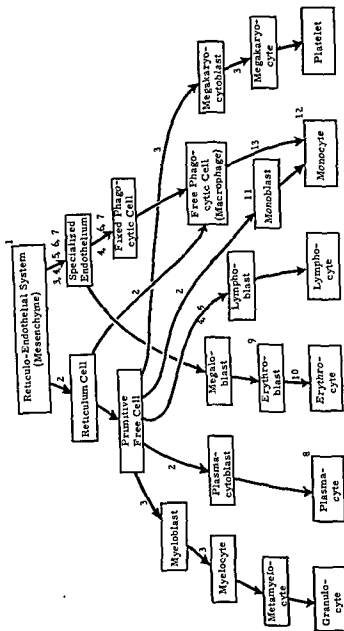
56. GENEALOGY OF THE FORMED ELEMENTS OF BLOOD  
Monophyletic Theory, William Bloom, (1952)



/1/ In ectopic myelopoiesis. /2/ Normally in bone marrow, in lymphatic and other tissues in ectopic myelopoiesis. /3/ Also gives rise directly to the fibroblast, fat cell, chondrocyte, osteoblast, and in the embryo, to the tissue mast cell and the endothelial cell. The fibroblast, chondrocyte, and osteoblast each may give rise to the osteocyte which in turn may give rise to the osteoclast, and thus to the reticular cell. Whether fibroblasts can become non-storing reticular cells in mammals is not clear. /4/ Gives rise in tissue culture and inflammation to the macrophage, and thus to the fibroblast.



58. GENEALOGY OF THE FORMED ELEMENTS OF BLOOD  
Polyphyletic (Dualist) Theory, Charles A. Doan, (1952)



/1/ Also gives rise to the lipoblast which gives rise to the fat cell, both the former and the latter cells are found in special fat organs, diffuse connective tissue and bone marrow /2/ In diffuse connective tissue and organ parenchyma. /3/ In bone marrow. /4/ In lymph nodes. /5/ In spleen. /6/ In Kupffer cells in liver. /7/ In endothelium (or sinusoids) of locations cited. /8/ Rare in blood, more common in connective tissue. /9/ Both early and late forms. /10/ Genealogical sequence from erythroblast is: normoblast, reticulocyte and erythrocyte. /11/ May give rise to myeloblast in tissue culture. /12/ May give rise to fibroblast like cell in tissue culture and may revert to monoblast on demand. Monocyte also may give rise to epithelioid cell in fatty degeneration (of tuberculosis caseation) and to the Langhans giant cell, and foreign body giant cell. /13/ On demand. There is evidence that macrophage may also give rise to fat cell or the reverse.

## ORGANS

1. FOR CELLS OF BLOOD AND BLOOD-FORMING ORGANS	Specific Cell Type	Assay
		Assay

59. NOMENCLATURE RECOMMENDED* FOR			
General Cell Type	Specific Cell Type		General Cell Type
	Term to be used	Terms to be avoided	
(A)	(B)	(C)	(D)
Erythrocytic Series	Rubriblast	Erythroblast, megakaryoblast, promyeloblast, promegakaryoblast, normoblast, hemocytoblast, lymphoblast, erythroblast	Myeloblast
		Erythroblast, megakaryoblast, promyeloblast, normoblast, macroblast, prokaryocyte	Progranulocyte
		Normoblast, pronormoblast, megakaryoblast, promegakaryocyte, polychromatophilic normoblast, karyocyte	Metamyelocyte
Erythrocytic Series	Rubricyte	Normoblast, erythroblast, metakaryocyte	Band Cell
		Metarubricyte	Segmented
		Reticulocyte <sup>2</sup>	Plasmablast
Erythrocytic Series	Lymphoblast	Red blood cell, erythroplastic, normocyte, erythrocyte	Propiasecyte
		Megakaryoblast, hemocytoblast, lymphoblast, stem cell, lymphocyte	Plasmacyte
		Large lymphocyte, pathologic lymphocyte, atypical lymphocyte, immature monocyte, immature lymphocyte	Thrombocyte
Lymphocytic Series	Pro-lymphocyte	Small, medium, or large lymphocyte, normal lymphocyte, small, medium, or large mononuclear	Megakaryoblast
		Megakaryoblast, hemocytoblast, lymphocyte, stem cell, immature monocyte	Promegakaryocyte
		Promonocyte, hemoblastoblast, immature monocyte, Ferrata cell	Megakaryocyte
Lymphocytic Series	Lymphocyte	Large mononuclear, transitional, plasmalocyte, endometrial leukocyte, histiocyte, resting wandering cell	Disintegrated cell
		Large mononuclear, transitional, plasmalocyte, endometrial leukocyte, histiocyte, resting wandering cell	Not identifiable
		Large mononuclear, transitional, plasmalocyte, endometrial leukocyte, histiocyte, resting wandering cell	Not identifiable

\*The Nomenclature of Cells and Diseases of the Blood and Blood-forming Organs. This Committee is sponsored by the American Medical Association. 1/21 It is recommended that the reticulocyte stage

1. By the Committee as reported by the American Society of the erythrocyte stage.

# 60. BONE MARROW (STERNAL) DIFFERENTIAL CELL COUNT: MAN

Author	Isaacs (1946)	Diggs (1949)	Whitby & Britton (1950)	Leitner (1945)	Lucia (1947)	Herman (1949)	Wintrobe (1947)	Vaughan (1947)	Osmond (1944)
Amount aspirated (ml)	0.1-0.2	0.2-0.5	0.2-0.5	0.2-0.5	0.5	1.5	1.0-2.0	3.0	0.5-1.0
Number of cases	10	10	10	10	10	10	10	10	10
Cell count in per cent (%)	Range (10)	Range (10)	Range (10)	Range (10)	Range (10)	Range (10)	Range (10)	Range (10)	Range (10)
	0-5-4	0-2	0-4	0-2	0-6-3	0-4-5	0-4	0-6	0-6
	1-5	1-7	4-15	3-2-8	8-9	10-5-10	1-8	0-12	0-12
	12-20	2-5-13	6-15	24-2-11	12-0-11	10-5-10	7-32	6-0-15	4-8-6
	6-10	7-20-11	7-19	1-2	1-0-0-3-3	1-16	0-3-5	3-0-17	1-5-6
	0-3-2	0-3	0-2-5	2-2-1	4-0-21	0-7-6	0-3	0-4-19	0-10
	1-8	1-5-20	0-5-21	0-0-2	10-2	6	0-3	1-4-21	0-3-6
	5-20	2-8	0-1	12-6-22	10-2	2-18	13-32	8-2-16	0-12-6
	13-32-27	5-12-28	10-25	1-4	0-8	0-5-2	0-0-5	6-2-26	3-10-6
	20-40	20-40	0-2-5	0-8	1-3	0-2-3	23-9	12-34	17-33-6
	7-26	9-30	10-40-27-31	28-4	10-0-31	0-34	18-23	6-34	12-0-34
	0-5-40	0-30	0-4-21	1-8	0-3-11	17	0-5-4	0-6-5	2-0-4
	0-1	0-1	0-1-31	0-0-2	0-2-3	2	0-0-7	0-1-5	0-0-5
	3-20	5-15	5-20	7-6	10-3	0-2-2	3-6	7-35	3-2-16
	0-5-5	0-1	0-5	1-4	0-9	0-2-2	2-4	0-2-6	0-4-6
	0-2	0-1-35	0-1	0-8	0-0-3-3	0-4	0-1-5	0-1-5	0-1-5
	3-5	3-5	3-5	3-5	3-5	3-5	3-5	3-5	3-5
Cells	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000	60,000-100,000

1/1 Smoothed weighted mean. 2/2 Per 100 leukocytes. 3/3 Megakaryoblasts. 4/4 Promonoblasts. 5/5 In per cent of red series. 6/6 Karyoblasts. 7/7 Early erythroblasts. 8/8 Macrophages. 9/9 Erythroblasts. 10/10 Basophilic normoblasts. 11/11 Normoblasts. 12/12 Prokaryocytes. 13/13 Late erythroblasts. 14/14 Polychromatic normoblasts. 15/15 Karyocytes. 16/16 Orthochromatic normoblasts. 17/17 Metakaryocytes. 18/18 Myeloblasts & leukoblasts. 19/19 Granuloblasts. 20/20 Promyelocytes. 21/21 Promyelocytes. 22/22 Neutrophilic band neutrophils & neutrophilic myelocytes. 23/23 Includes early neutrophilic myelocytes. 24/24 May include eosinophilic metamyelocytes. 25/25 Young forms. 26/26 Metagranulocytes. 27/27 Eosinophilic, eosinophilic, & basophilic. 28/28 Includes eosinophilic band neutrophils, metamyelocytes, band cells, and seg cells. 29/29 Juvenile neutrophils. 30/30 Juvenile neutrophils, eosinophilic, & basophilic. 31/31 Polymorphonuclear neutrophils, eosinophilic, & basophilic. 32/32 May include eosinophilic band neutrophils. 33/33 Filament cells, neutrophilic, eosinophilic, & basophilic. 34/34 Endothelial cells. 35/35 Eosinophilic cells. 36/36 Eosinophilic cells. 37/37 Eosinophilic cells. 38/38 Eosinophilic cells. 39/39 Eosinophilic cells. 40/40 Eosinophilic cells.

61. BONE MARROW (STERNAL)<sup>1</sup> DIFFERENTIAL CELL COUNT.  
SUMMARY: MAN<sup>2</sup>

Cell Types			Value <sup>3,5</sup> %	Range <sup>4,5</sup> %
			(D)	(E)
(A)	(B)	(C)		
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17	Red Series 19.1%	Proerythroblasts Early Normoblasts	0.5 2.4	0.2 - 4.0 1.5 - 5.8
	Early 2.9%	Intermed. Normoblasts	11.7	5.0 - 26.4
	Late 16.2%	Late Normoblasts	4.5	1.6 - 21.5
	Granulo- cytic 57.4%	Myeloblasts	1.2	0.3 - 3.1
		Progranulocytes	3.0	0.5 - 4.5
		Myelocytes	8.7	0.9 - 20.3
		Metamyelocytes	11.0	5.6 - 22
		Band Cells	17.9	6.1 - 36
	Others 12.6%	Segmented Cells	15.6	8.7 - 27
		Lymphocytes	9.8	2.7 - 24
		Monocytes	1.4	0.7 - 2.8
		Megakaryocytes	0.2	0.03- 0.4
		Plasmacytes	0.6	0.1 - 1.5
		Reticulum Cells	0.6	0.03- 1.6
	Not Identifiable 10.9%	Unclassified Cells	1.7	0.02- 3.3
		Disintegrated Cells	9.2	1.1 - 20.8

/1/ Sternal aspirations of 0.1-10.0 ml of bone marrow. /2/ Men and women, 750 adults. /3/ Grand mean of 21 authors' means.  
/4/ Range of authors' means. /5/ These values are averages of results from different laboratories using different techniques.  
Values from any one laboratory may be expected to show a consistent departure from the averages.

# 62. BONE MARROW (STERNAL) DIFFERENTIAL CELL COUNT IN PREGNANCY: MAN

Cell Types	First Trimester				Second Trimester				Third Trimester			
	Cells per mm <sup>3</sup>		% of Total		Cells per mm <sup>3</sup>		% of Total		Cells per mm <sup>3</sup>		% of Total	
	Value	Range	(A)	(B)	Value	Range	(C)	(D)	Value	Range	(E)	(F)
1 Proerythroblasts ("Rubriblasts")	35	0-116	0.2	0.0-0.4	73	0-203	0.2	0.0-1.0	61	0-210	0.3	0.0-0.8
2 Early Normoblasts ("Proerythrocytes")	749	86-1,314	2.3	0.6-3.2	797	251-1,875	2.0	1.0-4.0	1,050	51-3,937	5.1	3.9-11.2
3 Intermediate Normoblasts ("Metarubricytes")	1,139	259-2,428	3.2	1.6-5.0	1,803	314-7,000	4.0	2.0-6.4	1,865	51-4,616	9.1	6.1-14.1
4 Late Normoblasts	944	28-2,700	2.2	0.2-6.0	835	250-2,000	2.2	0.4-7.0	836	119-2,719	4.1	3.1-11.1
5 Myeloblasts	53	0-118	0.1	0-0.5	173	0-1,250	0.3	0-1.0	96	0-435	0.5	0.0-1.5
6 Progranulocytes	252	38-675	0.7	0.0-1.5	445	0-3,508	0.7	0-2.8	362	51-1,050	1.9	1.0-4.0
7 Myelocytes, Eosinophilic <sup>1/2</sup>	213	28-494	0.7	0.2-2.6	351	0-1,408	0.4	0-1.6	167	0-700	0.9	0.0-2.4
8 Myelocytes, Neutrophilic <sup>1/2</sup>	2,162	172-4,316	6.0	1.2-10.6	3,231	659-13,125	7.0	4.0-11.2	2,138	391-7,337	11.1	8.1-18.1
9 Metamyelocytes, Neutrophilic	2,165	258-5,913	7.7	1.8-9.0	2,823	251-10,250	5.4	1.6-8.6	1,879	340-4,900	9.6	6.6-14.6
10 Band Neutrophils	11,659	2,059-24,135	30.2	9.0-38.0	9,444	1,416-24,135	19.3	8.6-24.6	11,613	4,240-29,135	60.3	40.3-70.3
11 Segmented Eosinophils <sup>3</sup>	371	48-1,140	1.3	0.2-6.0	239	18-799	0.8	0.1-1.8	228	31-740	1.2	0.0-3.2
12 Segmented Basophils	78	0-197	0.3	0-1.0	47	0-128	0.2	0-0.6	49	0-191	0.3	0.0-0.8
13 Lymphocytes	3,975	2,894-6,200	13.5	4.5-24.8	6,467	1,684-10,000	12.9	5.6-23.0	3,719	1,118-14,000	19.5	13.5-25.5
14 Monocytes	333	1-965	1.1	0.1-3.0	453	0-1,645	1.0	0-4.0	194	0-732	1.0	0.0-2.4
15 Plasma Cells	4	0-8	0.0	0-0.0	13	0-11	0.0	0-0.0	4	0-13	0.0	0-0.0
16 Total Increased Cells	24,580	14,400-68,700	13.5	9.0-18.0	41,510	15,700-125,000	12.4	6.0-23.5	33,930	16,700-70,000	18.1	13.1-23.1
Fourth, Second, Third Trimesters												
Cell Types	Fourth Trimester (Continued)		Second, Third Trimesters		Non-Pregnant Females		Cells per mm <sup>3</sup>		% of Total		Cells per mm <sup>3</sup>	
	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range
	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)	(R)
1 Proerythroblasts ("Rubriblasts")	0.2	0.0-0.5	64	0-210	0.2	0.0-1.0	28	0-122	0.1	0.0-0.4	0.3	0.0-0.8
2 Early Normoblasts ("Proerythrocytes")	2.8	0.3-6.2	886	51-3,937	2.4	0.3-6.2	616	62-1,396	2.5	0.6-4.0	1.0	0.1-2.2
3 Intermediate Normoblasts ("Metarubricytes")	5.3	0.3-13.6	1,462	51-8,626	4.3	0.3-13.6	1,199	136-2,924	5.0	1.3-8.0	0.5	0.0-1.6
4 Late Normoblasts	2.4	0.6-6.5	863	28-2,730	2.3	0.2-7.0	455	61-1,089	1.9	0.4-3.2	0.3	0.0-0.8
5 Myeloblasts	0.2	0.0-1.0	115	0-1,250	0.2	0.0-1.0	89	0-279	0.3	0.0-0.8	0.3	0.0-0.8
6 Progranulocytes	0.8	0-2.3	393	0-2,500	0.9	0-2.3	231	11-736	1.0	0.1-2.2	0.5	0.0-1.6
7 Myelocytes, Eosinophilic <sup>1/2</sup>	0.4	0.0-1.0	209	0-1,408	0.5	0.2-2.6	147	0-593	0.5	0.0-1.6	0.3	0.0-0.8
8 Myelocytes, Neutrophilic <sup>1/2</sup>	5.7	2.3-12.5	2,562	172-13,125	6.5	1.2-15.2	1,568	168-4,160	6.0	1.4-11.0	1.0	0.1-2.2
9 Metamyelocytes, Neutrophilic	5.3	2.0-9.0	2,511	251-10,250	5.3	1.6-9.0	1,537	188-4,324	5.6	1.4-9.4	1.0	0.1-2.2
10 Band Neutrophils	34.9	21-62.5	10,415	2,189-46,250	33.3	15.2-48.0	6,048	1,091-14,490	23.7	12-43.4	3.6	1.0-6.0
11 Segmented Eosinophils <sup>3</sup>	0.7	0-1.6	576	59-1,140	0.9	0-3.0	345	1,132-7,094	16.6	10-43.1	1.0	0.1-2.2
12 Segmented Basophils	0.2	0.0-0.6	36	0-197	0.2	0-1.0	53	60-142	2.3	0-6.6	0.3	0.0-0.8
13 Lymphocytes	12.0	5.5-23.3	4,645	1,318-10,000	12.7	4.5-28.0	3,428	1,721-6,520	16.8	9.2-30.8	1.4	0.0-3.5
14 Monocytes	0.7	0.0-2.4	246	1,037-11,809	0.8	0-3.0	378	1,081-6,536	16.2	10.5-24.0	0.3	0.0-0.8
15 Plasma Cells	13.2	5.5-23.6	4,599	14,000-125,000	12.9	5.5-23.6	3,500	1,500-46,000	16.2	10.5-24.0	0.3	0.0-0.8
16 Total Increased Cells	38,740	14,400-68,700	13.5	9.0-18.0	41,510	15,700-125,000	12.4	6.0-23.5	33,930	16,700-70,000	18.1	13.1-23.1

1/1 40 pregnant females and 28 non-pregnant females, 10 ml marrow aspirated. Progranulocytes, plasmacytes, basophilic myelocytes, early myelocytes (eosinophilic and basophilic), not isolated but present in numbers less than 0.1%. 2/2 Includes eosinophilic metamyelocytes. 3/3 Includes band eosinophils.



# 63. BONE MARROW (RIB) DIFFERENTIAL CELL COUNT: DOG

Author	Stasney & Higgins (1937)			Van Loon (1943)			Mulligan (1941)			Rakers & Coulter (1948)			Mulligan (1945)		
	Adult			Adult			Adult			19-24 mos.			21		
	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value
Age															
Number of cases	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value	Range	Value
Cell count in per cent (%)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)
(A)															
1 Myeloblasts	2.41	0-5.1 <sup>b</sup>	0.6	0.2-1 <sup>c</sup>	0.6	0.2-4.6 <sup>c</sup>	2.7	2.2-11 <sup>c</sup>	4.7	5.6-20 <sup>c</sup>	5.1	17-63 <sup>c</sup>	0.2-3.7 <sup>c</sup>	0.8 <sup>c</sup>	0-2.2 <sup>c</sup>
2 Early Myelocytes	2.82	0-5.8 <sup>b</sup>	6.0	2.7-10 <sup>c</sup>	4.7	5.6-20 <sup>c</sup>	42.4	17-63 <sup>c</sup>	20.6	15-28 <sup>c</sup>	9.7	7-12 <sup>c</sup>	9.7	7-12 <sup>c</sup>	15-28 <sup>c</sup>
3 Myelocytes, all	8.9	2.8-15 <sup>b</sup>	3.4	1.1-4.6 <sup>c</sup>	10.5	17-54 <sup>c</sup>	5.1	17-63 <sup>c</sup>	20.6	15-28 <sup>c</sup>	9.7	7-12 <sup>c</sup>	9.7	7-12 <sup>c</sup>	15-28 <sup>c</sup>
4 Neutrophils	1.2	0-2.4 <sup>b</sup>	3.4	1.1-4.6 <sup>c</sup>	10.5	17-54 <sup>c</sup>	5.1	17-63 <sup>c</sup>	20.6	15-28 <sup>c</sup>	9.7	7-12 <sup>c</sup>	9.7	7-12 <sup>c</sup>	15-28 <sup>c</sup>
5 Eosinophils	15.3	7.2-23 <sup>b</sup>	11.7	6.8-17 <sup>c</sup>	31.0	17-54 <sup>c</sup>	5.1	17-63 <sup>c</sup>	20.6	15-28 <sup>c</sup>	9.7	7-12 <sup>c</sup>	9.7	7-12 <sup>c</sup>	15-28 <sup>c</sup>
6 Metamyelocytes			30.1	17-44 <sup>c</sup>	3.9	0.2-8.5 <sup>c</sup>	4.73	0.2-19 <sup>c</sup>	2.43	0.8-6.6 <sup>c</sup>	3.4	0.8-6.6 <sup>c</sup>	3.4	0.8-6.6 <sup>c</sup>	0.8-6.6 <sup>c</sup>
7 Band Cells	5.1	0-12.5 <sup>b</sup>	2.0	0.4-3.8 <sup>c</sup>	3.73	1-6.8 <sup>c</sup>	0.2	0-1.3 <sup>c</sup>	3.3	1-6.6 <sup>c</sup>	3.3	1-6.6 <sup>c</sup>	3.3	1-6.6 <sup>c</sup>	1-6.6 <sup>c</sup>
8 Segmented Cells	2.8	0-6.8 <sup>b</sup>	0.9	0.2-2.7 <sup>c</sup>	1.9	0-6.6 <sup>c</sup>	0.7	0-1.1 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	0.4-3.2 <sup>c</sup>
9 Neutrophils	0.1	0-0.3 <sup>b</sup>	0.2	0-0.3 <sup>c</sup>											
10 Eosinophils	1.2	0-2.3 <sup>b</sup>	0.5	0-1.4 <sup>c</sup>	0.54	0-4.3 <sup>c</sup>	0.7	0-1.1 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	0.4-3.2 <sup>c</sup>
11 Basophiles			0.64	0.2-2.7 <sup>c</sup>	0.54	0-4.3 <sup>c</sup>	0.7	0-1.1 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	0.4-3.2 <sup>c</sup>
12 Lymphocytes	0.1	0-0.5 <sup>b</sup>	0.5	0-1.4 <sup>c</sup>	0.54	0-4.3 <sup>c</sup>	0.7	0-1.1 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	1.3 <sup>a</sup>	0.4-3.2 <sup>c</sup>	0.4-3.2 <sup>c</sup>
13 Monocytes			7.86	6.4-10 <sup>c</sup>	1.56	0.4-3.8 <sup>c</sup>	28.27	8-54 <sup>c</sup>	45.189	33-57 <sup>c</sup>	45.189	33-57 <sup>c</sup>	45.189	33-57 <sup>c</sup>	33-57 <sup>c</sup>
14 Megakaryocytes			16.48	11-26 <sup>c</sup>	38.189	19-64 <sup>c</sup>									
15 Proerythroblasts			17.49	9-26 <sup>c</sup>											
16 Early Normoblasts	59	40-76 <sup>b</sup>													
17 ("Prorubricytes")															
18 ("Rubricytes")															
19 Late Normoblasts															
20 ("Metarubricytes")															
21 Plasmacytes	1.	0-2.1 <sup>b</sup>													
22 Reticulum Cells	0.211	0-0.7 <sup>b</sup>													
23 Unclassified Cells															
24 Total Nucleated Cells (millions/mm <sup>3</sup> )															

1/ Includes "Leukoblasts." 2/ Promyelocytes. 3/ Eosinophilic cells, all forms. 4/ Pronormoblasts. 5/ Megakaryoblasts. 6/ Basophilic normoblasts. 7/ Erythroblasts. 8/ Polychromatic normoblasts. 9/ Orthochromatic normoblasts. 10/ Normoblasts. 11/ Includes "Heterophils." 12/ Mean of the means. 13/ Range of the means.

64. BONE MARROW (RIB)<sup>1</sup> DIFFERENTIAL CELL COUNT, SUMMARY: DOG<sup>2</sup>

Cell Types				Value <sup>4,6</sup> %	Range <sup>5,6</sup> %
(A)	(B)	(C)		(D)	(E)
1 2 3 4	Red Series 43.6%	Early 5.4% Late 38.2%	Proerythroblasts Early Normoblasts Intermed. Normoblasts Late Normoblasts	0.5 4.9 22.3 15.9	0.3 - 0.6 1.5 - 7.8 11.0 - 26. 4.6 - 17.4
5 6 7 8 9 10 11	White Series and Others 54.9%	Granulo- cytic 52.2%	Myeloblasts Early Myelocytes, Neutrophil. Myelocytes, Neutrophilic Metamyelocytes, Neutrophilic Band Cells, Neutrophilic Segmented Cells, Neutrophil. <sup>3</sup> Eosinophilic Cells, all types	1.2 1.4 4.8 7.4 24.6 9.6 3.1	0.6 - 2.4 0.7 - 2.8 2.7 - 8.9 3.4 - 15.3 11.7 - 42 3.9 - 30 2.0 - 4.7
12 13 14 15 16		Others 2.7%	Lymphocytes Monocytes Megakaryocytes Plasmacytes Reticulum Cells	0.9 0.2 0.4 0.3 0.9	0.7 - 1.9 0.0 - 0.2 0.1 - 0.6 0.0 - 0.4 0.0 - 1.0
17 18	Not Identifiable 1.5%		Unclassified Cells	1.5	0.0 - 3.0

/1/ Rib extrusions. /2/ 187 adult dogs. /3/ Includes basophilic cells (0.05-0.2 range of means). /4/ Grand mean of 4 authors' means.

/5/ Range of 4 authors' means. /6/ These values are averages of results from different laboratories using different techniques. Values from any one laboratory may be expected to show a consistent departure from the averages.

# 65. MORPHOLOGY OF FORMED ELEMENTS OF BONE MARROW: MAN<sup>1</sup>

## CYTOPLASM

Cell Type	Size $\mu$	Granules	Relative Amount	Color
(A)	(B)	(C)	(D)	(E)
1 Reticulum cell (Reticulo-endothelial cell, Stem cell of modern pathologists, Mesenchymal cell, Hemohistioblast)	15-25	May or may not contain fine, angular, purple, azurophil granules	Usually moderate; more than myeloblast.*	Pale to dark blue; may be mottled with abundant, colorless, or slightly acidophilic hyaloplasm.
2 Myeloblast* (Hemocytoblast, Stem cell of classic hematologists)	10-18	May or may not contain fine, angular, purple, azurophil granules	Usually a narrow band	Pale to dark blue.
3 Progranulocyte* (Leukoblast, "Myeloblast" of Italian authors)	14-21 mean = 17	May or may not contain fine, angular, purple, azurophil granules; are less commonly round and larger; may be numerous; may overlie nucleus.	Usually moderate; more than myeloblast.*	Varying shades of blue but usually pale blue or mottled.
(Early) Myelocyte* (Promyelocyte)	12-18	May or may not contain azurophil granules. At this stage the first few specific granules are seen.*	Usually moderate to abundant.	Varying shades of blue, usually pale; paler areas in region of cytocentrum, darker at periphery.
(a) Neutrophilic		(a) Pink specific granules first appear about cytocentrum; may show diffuse pink masses of neutrophilic substances.		
(b) Eosinophilic		(b) Round, red, large specific granules; a few early ones are blue to purple.		
(c) Basophilic		(c) Few very large, metachromatic, specific granules; vary in size; water soluble.		
(Late) Myelocyte (a) Neutrophilic	10-16	(a) Specific granules are pink and more numerous than in early myelocyte	Moderate to abundant.*	(a) Usually clear or colorless
(b) Eosinophilic		(b) Specific granules are red and more numerous than in early myelocyte		(b) May be pale blue.
(c) Basophilic		(c) Specific granules are metachromatic and more numerous than in early myelocyte		(c) May be pale blue.

/1/ Table applies only to elements as observed in ordinary air-dried smears after Romanowsky stains. For characteristics of the corresponding elements as observed in tissue sections and moist smears, refer to Chapter 5, Maximow, A., and Bloom, W., "A Textbook of Histology," W. B. Saunders, Philadelphia, 1948. For characteristics of the corresponding elements as observed with the supravital technique, refer to Tompkins, E. H., and Cunningham, R. S., in Sections IX and X in Vol. I of "Handbook of Hematology," Hal Downey, Ed.

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

## NUCLEUS

Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Parachromatin Pattern and Amount
(F)	(G)	(H)	(I)	(J)	(K)
Round, oval, reniform, or rarely indented	Central or eccentric	Moderately thin	0-6; usually 3-6; round irregular	Irregular network of light, red-violet	Small amount; pink or pale blue; fills irregular chinks in chromatin interstices; abundant and distinct from chromatin
			ance.	unequal in size.	
Round or oval	Usually central	Very thin and indistinct	0-5, pale blue, round or oval, occasionally irregular, chromatin may delimit nucleoli	Fine network of chromatin strands with regular, equal, small interstices; chromatin strands are thin, or the pattern may appear as one of fine regular stippling, no clumping; stains a light lavender.	Small amount; fine pink or pale blue parachromatin in small interstices of chromatin; distinct from chromatin.
Round or oval	Central or eccentric.	Thin	clumped about periphery	clumped clumping of chromatin, especially about nucleoli	
Round or oval	Central or eccentric	Thin	Occasionally present, usually not visible; when present, pale blue and round or oval	Network of chromatin filaments, slightly coarsened with less regular interstices	Small amount; pale blue or pink; distinct from chromatin
Round or oval	Central or eccentric	Distinct, thick	None visible	Thickened chromatin strands form a coarse meshwork, staining a deep purple	Small amount; pale blue or pink; distinct from chromatin

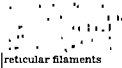
tor, Paul B. Hoeber, New York, 1938 /2/ Nomenclature recommended by Committee for Clarification of the Nomenclature of Cells and Diseases of the Blood and Blood-forming Organs (Blood 4:89, 1949) /3/ Vacuoles are common in cytoplasm, may have clear ectoplasm /4/ Abundant hyaloplasm /5/ All characterized by specific granules and basophilic spongioplasm /6/ All characterized by increase in colorless hyaloplasm and in specific granules

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

CYTOPLASM				
Cell Type	Size μ	Granules	Relative Amount	Color
(A)	(B)	(C)	(D)	(E)
6 Metamyelocyte* (Juvenile) (a) Neutrophilic	10-16	(a) Specific granules are pink to rose-violet and more numerous than in late myelocyte.	Abundant	Colorless or slightly acidophilic.
(b) Eosinophilic		(b) Specific granules are red and more numerous than in late myelocyte.		
(c) Basophilic		(c) Specific granules are metachromatic and more numerous than in late myelocyte.		
7 Granulocyte, neutrophilic, band (Regenerative).	(See Table on Morphology of Blood Leukocytes and Platelets: Man)			
8 Granulocytes, segmented: Neutrophilic Eosinophilic Basophilic.				
9 Lymphoblast*	10-18	Azurophil granules are usually absent	Usually forms narrow band; may have large cell body†	Varying shades of pale to dark blue.
10 Prolymphocyte* (Immature lymphocyte).	10-18	Azurophil granules may or may not be present; usually absent.	Usually forms very narrow band, may have large cell body	Varying shades of pale to dark blue.
11 Lymphocyte Small Medium	(See Table on Morphology of Blood Leukocytes and Platelets: Man)			
12 Leukocytoid Lymphocyte (Pathological lymphocyte, e.g., as in infectious mononucleosis)	10-16	May or may not contain round, purple azurophil granules. Granules, if present, may be small or large, sparse or numerous	May have increased azurophilic granules.	

1/ Table applies only to elements as observed in ordinary air-dried smears after Romanowsky stains  
 2/ Nomenclature recommended by Committee for Clarification of the Nomenclature of Cells and Diseases of

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

NUCLEUS					
Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Pattern and Amount Parachromatin
(F)	(G)	(H)	(I)	(J)	(K)
Reniform, moderately indented	Central or eccentric	Thick, heavy	None visible	Thickened chromatin strands form a coarse meshwork, staining a deep purple	Small amount; pale blue or pink, distinct from chromatin in the interstices of the latter
Round or oval	Usually central	Thin	0-5, usually 1-2; pale blue, small, round, chromatin does not mass about nucleoli	Fine network of chromatin strands with regular, small interstices. Chromatin strands are thin, or  reticular filaments	Small amount; fine pink or pale blue, in small interstices of chromatin; dis-
Round, oval, or reniform, slightly indented	Central or eccentric	Thin to thick	Occasional blue, round, unmasked nucleolus	In some areas, fine to coarse network of chromatin strands, such areas blend imperceptibly with areas with indistinct, coarse, clumped chromatin masses	Small amount, pale blue, in some areas still distinct; in other areas blends with chromatin.
Round, oval, deeply indented, reniform, or irregular	Usually eccentric	Thick		Pattern may be lymphocyte-like (11J) if accompanied by extensive cytoplasmic changes, or pattern may be plasmacytic,	Varying amounts of

the Blood and Blood-forming Organs (Blood 4, 89, 1949) /7/ Cytoplasm is homogeneous /8/ Downey recognized three cell types on the basis of these characteristics

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

## CYTOPLASM

Cell Type		Size $\mu$	Granules	Relative Amount	Color
(A)		(B)	(C)	(D)	(E)
13	Monoblast <sup>2,3</sup>	14-18	May or may not contain fine azurophilic granules.	Moderate or abundant.	Slightly basophilic or gray.
14	Monocyte <sup>4</sup> (Transitional or Blood mononuclear)	(See Table on Morphology of Blood Leukocytes and Platelets: Man)			
15	Megakaryoblast <sup>5</sup>	circa 15-50	Usually absent. A few coarse granules may be present.	As cell matures to promegakaryocyte, amount varies from narrow band to abundant.	Varying shades of blue. Usually zoned in lighter and darker areas. <sup>6</sup>
16	Promegakaryocyte <sup>4,11</sup>	circa 18-50	Azurophilic granules begin to make their appearance near cytocentrum and spread through cell body.	Usually abundant.	Basophilic, but becomes paler blue as maturation approaches.
17	Megakaryocyte <sup>4</sup>	circa 30-70	Azurophilic granules	Large amounts. Pseudopodial processes frequent.	Pale blue. May begin
18	Thrombocyte (Platelet)	(See Table on Morphology of Blood Leukocytes and Platelets: Man)			
19	Pronormoblast <sup>12</sup> or Proerythroblast (Megakaryoblast of Supravitalists, Rubriblast) <sup>4</sup>	10-15	Usually absent.	Usually narrow band; ordinarily not quite as much as myeloblast, but both vary; almost always less than promegakaryoblast (24D).	Basophilic; stains deeply blue and is usually more homogeneous and condensed than that of myeloblast or promegakaryoblast; may have clear perinuclear zone.
				Because of shrinking of	Basophilic; stains

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

NUCLEUS					
Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Parachromatin Pattern and Amount
(F)	(G)	(H)	(I)	(J)	(K)
Round, oval, may be indented	Central or eccentric	Thin	0-10, pale blue, well circumscribed.	more darkly than myeloblast pattern	work.
Irregular or polylobular "Rarely multinucleated"	Usually eccentric	Thin	Usually less numerous than megakaryoblast, (15f) but may still be very numerous	Stains more heavily than megakaryoblast; a regular distinct network remains but chromatin threads are coarser than in megakaryoblast and may show areas of clumping.	Small amount of pink between chromatin strands where they are not clumped
Markedly multilobular and complicated in form, less frequently round, oval, or multinuclear.	Central or eccentric	Thick, heavy	None visible	visible	chromatin may be seen
Round or oval	Central or eccentric	Very thin	Present but faint; appear to be dissolved out; give blue tint.	Similar to that of myeloblast (2J) but chromatin strands forming network are slightly thicker and there is some clumping, thus appearing more heavily stained, interstices are distinct but slightly irregular.	Small amount, less than myeloblast and much less than promegaloblast, stains pink and is distinct from chromatin
Round and smaller than pronormoblast	Central or eccentric	Thin	May still be present though usually none visible	Network now complete	Small amount; ar-

usually identified by "the company it keeps" /10/ Light area of hyaloplasm around nucleus /11/ Large cells of this group are called Intermediate Megakaryocytes /12/ Nucleus larger than megakaryocyte. /13/ There is a gradual decrease in nuclear size due to condensation as all the following cells in this series differentiate



# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

CYTOPLASM				
Cell Type	Size $\mu$	Granules	Relative Amount	Color
(A)	(B)	(C)	(D)	(E)
21 Polychromatophilic Normoblast <sup>14</sup> or Polychromatophilic Erythroblast (Late Rubricyte) <sup>2</sup>	circa 8-12	Absent	Because of shrinking	Hemoglobin first appearing in center
22 Orthochromatic or Acidophilic Normoblast. (Metarubricyte) <sup>2</sup>	circa 7-10	Absent	Because of condensed nucleus, cytoplasmic to nuclear ratio is increased but is less than that of orthochromatic megaloblast.	Red, homogenous, and smooth because of hemoglobin predominance.
23 Erythrocyte <sup>2,14</sup>	7.2-7.8 mean $\approx 7.5$	Absent	Non-nucleated corpuscle formed by loss of nucleus from the orthochromatic normoblast.	Orange-red hemoglobin; center is slightly paler than periphery.
24 Promegaloblast <sup>14</sup> (Rubriblast, Pernicious Anemia type)	13-25	Usually absent		
25 Basophilic Megaloblast (Prorubricyte, P.A. type and Rubricyte, P.A. type) <sup>2</sup>	13-20	Absent		
26 Polychromatophilic Megaloblast (Rubricyte, late, P.A. type) <sup>2</sup>	Usually 10-18 (May be much larger).	Absent		Varying amounts of eosinophilic (red)
27 Orthochromatic Megaloblast <sup>14</sup> (Late Rubricyte, P.A. type and Metarubricyte, P.A. type) <sup>2</sup>	Usually 8-15 (May be much larger)	Absent		

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

## NUCLEUS


Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Parachromatin Pattern and Amount
(F)	(G)	(H)	(I)	(J)	(K)
Round but smaller than basophilic normoblast	Central or eccentric	Thick	None visible.	Chromatin strands so coarse that in many places they are condensed into masses; they may be arranged as wheel-spokes ("rad-kern")	Small amount; arranged as pink, irregular chinks in interstices of chromatin
Round but smaller than polychromatophilic normoblast.	Central or eccentric	Thick, coarse, heavy	None visible	Nucleus is condensed, no longer is any pattern visible	May either be absent or one or two pink chinks may still remain

Round or oval	Central or eccentric.	Very thin, indistinct		Delicate structure, al-	More abundant than in myeloblast and much more than in pronormoblast. Appears as pink, comma-shaped areas distinct from chromatin.
Round and about same size as promegaloblast	Central or eccentric	Thin	Nucleoli often absent but they are more often present than in corresponding basophilic normoblast.		
Round	May be central but usually eccentric	Thin to thick.	May still be present, usually none visible.	Pattern remains the same as in the basophilic megaloblast; i.e., there is a slightly coarsened network with prominent interstices. Reticular structure still evident.	Remains abundant, pink; fills interstices in chromatin network from which it is distinct.
Round	Usually eccentric,	Thick	May still be present; usually	Pattern shows all gradations from a slightly coarsened network with numerous distinct interstices to	Parachromatin is pink; varies from abundant, when chromatin interstices are large to

tingu-  
decre-  
as the,

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Continued)

## CYTOPLASM

Cell Type		Size μ	Granules	Relative Amount	Color
(A)		(B)	(C)	(D)	(E)
28	Megalocyte <sup>10</sup>	Greater than 7.8	Absent	Non-nucleated corpuscle formed from the cytoplasm of the orthochromatic megaloblast, often pear-shaped.	Orange-red hemoglobin; homogeneous or slightly dark red through thicker center.
29	Plasmablast <sup>10,11,12</sup> (Reticular plasma cell).	25-40	Usually absent. (Azurophilic granules are rarely described in abnormal forms).	Large amounts.	Varying shades of blue, usually light blue or moderately deep; basophilic spongioplasm is mottled with bars of hyaloplasm (mitochondria); canaliculated appearance; Hof is indistinct; may show peripheral cytoplasmic dissolution.
30	Proplasmacyte <sup>10</sup>	circa 15-30	Usually absent.	Abundant, but less than plasmablast.	With shrinkage of cytoplasm, basophilia become more prominent. An eccentric, clear, perinuclear area, or Hof, develops; may show peripheral cytoplasmic dissolution.
31	Plasmacyte <sup>10</sup> (Marschalko plasma cell).	8-25	Usually absent.		Deep blue cytoplasm with large pale stains.
32	Tissue Mast Cell <sup>10</sup>	8-25	Entire cytoplasm is stuffed or packed with metachromatic granules <sup>10</sup> which are spherical and uniform in size. May be so abundant as to mask nucleus; granules more water soluble than those of blood basophils.	Abundant; cytoplasmic to nuclear ratio favors the cytoplasm.	Colorless

# 65. BONE MARROW MORPHOLOGY<sup>1</sup> (Concluded)

NUCLEUS					
Shape	Position	Nuclear Membrane	Nucleoli	Chromatin Pattern	Parachromatin Pattern and Amount
(F)	(G)	(H)	(I)	(J)	(K)
Round or oval	Eccentric.	Thin	0 to 6, blue.	Irregular network of fine chromatin strands with irregular interstices. Resembles pattern of reticulum cell nucleus (1J) but has thicker strands and may show some clumping of chromatin.	Pink or yellow; distinct from chromatin strands; fills interstices of chromatin.
Round or oval and smaller than that of plasmablast.	Eccentric	Thin to thick.	1 or 2 nucleoli may still be present	Contraction of nucleus leads to further coarsening and thickening of reticular network; irregular interstices are still distinct.	Small amount; distinct in interstices of chromatin although interstices are irregular.
Round or oval; may be binucleate or multinucleate. Usually smaller than nucleus of proplasmacyte (30F).	Eccentric.	Thick	None visible.	Chromatin further condensed; tends to pyknosis. Coarse chromatin masses arranged often in smears or imprints.	Small amount; dis-
Small, round, or oval; may be binucleate.	Eccentric.	Varies in thickness.	None visible	Coarse chromatin masses arranged irregularly in clumps, or almost homogeneously	Small amount.

pute, the lymphocyte at times is considered to function as the plasmacyte precursor /20/ Cell outline may be spherical, ellipsoidal, or irregular. /21/ Metachromatic material is believed by some to be in the cytoplasm between the granules (Jaques).

# 66. HISTOCHEMICAL PROPERTIES OF BLOOD AND BONE MARROW CELLS: MAN

Cytoplasm proper (C); Specific Granules (G); Juxtanuclear bodies (J); Mitochondria (M); Nucleoli (N)							
Cell Type (A)	Lipid <sup>1</sup> (B)	Phospho- lipid <sup>2</sup> (C)	Acid Phosphatase <sup>3</sup> (D)	Alkaline Phosphatase <sup>3</sup> (E)	Ribonucleo- protein <sup>4</sup> (F)	Carbohydrate <sup>5</sup> (G)	Peroxidase <sup>6</sup> (H)
1 Nuclei (all types)	Neg	Neg	Pos	Pos <sup>7</sup>	Pos N	Neg	---
2 Cytoplasm of:							
3 Myeloblasts	Pos M	Pos M	Neg	---	Pos C <sup>8</sup>	Neg	Neg
4 Progranulocytes	Pos GM	Pos G <sup>7</sup> M	Pos G	Neg <sup>9</sup>	Pos C <sup>8</sup>	Neg <sup>7,10</sup>	Pos G
5 Myelocytes	Pos GM	Pos G <sup>7</sup> M	Pos GC	Neg <sup>9</sup>	Neg	Pos <sup>7,10</sup>	Pos
6 Metamyelocytes	Pos G	Pos G <sup>7</sup>	Pos G	Pos C <sup>8,10</sup>	Neg	Pos <sup>7,10</sup>	Pos
7 Segmented neutrophils	Pos G	Pos G <sup>7</sup>	Pos G	Pos C <sup>8,10</sup>	Neg	Pos <sup>7,10</sup>	Pos
8 Segmented eosinophils	Pos G	Pos G	Pos G	Neg	Neg	Pos G <sup>10</sup>	Pos
9 Tissue eosinophils	Pos G	Pos G <sup>7</sup>	---	---	---	Pos <sup>7</sup>	Neg
10 Lymphoblasts	Pos G	Pos GM	Neg	Neg	Pos C <sup>8</sup>	Neg	Pos
11 Lymphocytes	Pos M	Pos M	---	---	Pos C <sup>8</sup>	---	Pos
12 Monocytes	Neg	Neg	Pos JG	Neg <sup>12</sup>	Neg C <sup>8</sup>	---	Neg
13 Megakaryocytes	Pos GM	Pos G	---	---	Pos C <sup>8</sup>	Neg <sup>13</sup>	Neg
14 Thrombocytes	Pos GM	Pos G <sup>7</sup> M	Pos <sup>8</sup>	Pos C <sup>8</sup>	Pos C <sup>8,9</sup>	Pos C	Pos <sup>11</sup>
15 Proerythroblasts ("Rubriblasts")	Neg	Pos C <sup>8</sup> G	---	---	Pos C <sup>8,9</sup>	Pos <sup>10</sup>	---
16 Early normoblasts ("Prorubricytes")	Pos M	Pos M	Pos J	Neg	Pos C <sup>8</sup>	Neg	Neg
17 Intermediate normoblasts ("Rubricytes")	Pos M	Pos M	Pos J	Neg	Pos C <sup>8</sup>	Neg	Neg
18 Late normoblasts ("Metarubricytes")	Neg	Neg	---	Neg	Pos C <sup>8</sup>	Neg	Neg
19 Erythrocytes	Neg	Neg <sup>8</sup>	Neg <sup>14</sup>	Neg	Neg	Neg	Neg
20 Plasmacytes	---	---	Pos JG	Pos C <sup>8</sup>	Neg	Neg	Pos
21 Reticulum Cells	Pos G	Pos G <sup>6</sup>	Pos G	---	Neg	Pos <sup>7</sup>	Neg
22 Mast Cells	Pos C <sup>16</sup>	Pos G	Pos G	Pos G	Neg	Pos G <sup>15</sup>	Pos

/1/ Stained with Sudan black B unless specified otherwise. /2/ Detected by acid-hematein method; negatively evaluated by pyridine extraction method. /3/ Detected by modified Gomori method using glycerophosphate substrate. /4/ Detected by Feulgen reaction; negatively evaluated by digestion with ribonuclease. /5/ Detected by either Periodic Acid-Schiff, or by Gomori silver-methenamine methods. /6/ Detected by benzidine-peroxide. /7/ Controversial. /8/ Diffuse staining. /9/ May be positive (Rabinovitch and Andreucci). /10/ As glycogen. /11/ Trace. /12/ May be positive. /13/ Positive according to Stowell, Wachstein. /14/ Positive when substrate is A.T.P. /15/ Probably is phagocytosed material (Lillie). /16/ Detected by staining with Sudan IV. /17/ Granules positive in many cells, negative in others.

67. BLOOD WATER AND SOLIDS						
Blood (B), RBC (C), Plasma (P), Serum (S)						
Animal		Water g/100 ml <sup>1</sup>		Solids g/100 ml		
		Value	Range	Value	Range	
(A)		(B)	(C)	(D)	(E)	
1	Man	B	83	81-86	23	20-25
2		C	72 <sup>2</sup>	70-75 <sup>2</sup>	37	34-39
3		P	94 <sup>2</sup>	93-95 <sup>2</sup>	8.6	7.9-9.1
4		S	93 <sup>3</sup>	93-94 <sup>c</sup>		
5	Cat	B	84 <sup>4</sup>		21 <sup>4</sup>	
6		C	62 <sup>3</sup>		38 <sup>3</sup>	
7		S	93 <sup>3</sup>		7 <sup>3</sup>	
8	Chicken	B	(87) <sup>4,5</sup>	(86-89) <sup>4,5</sup>		
9		C	72 <sup>4</sup>	72-74 <sup>4</sup>		
10		P	94 <sup>4</sup>	92-95 <sup>4</sup>		
11	Cow	B	85 <sup>4</sup>		20 <sup>4</sup>	
12		C	64 <sup>4</sup>		44 <sup>4</sup>	
13		S	91 <sup>3</sup>		9 <sup>3</sup>	
14	Dog	B	84	83-85	22	20-24
15		C	72	64-80 <sup>b</sup>		
16		P	93	91-95		
17		S	92 <sup>3</sup>		7.6 <sup>3</sup>	
18	Goat	B	84 <sup>4</sup>		20.5 <sup>4</sup>	
19		C	61 <sup>3</sup>		39 <sup>3</sup>	
20		S	91 <sup>3</sup>		9 <sup>3</sup>	
21	Horse	B	81 <sup>4</sup>		24 <sup>4</sup>	
22		C	61 <sup>3</sup>		39 <sup>3</sup>	
23		S	91 <sup>3</sup>		9 <sup>3</sup>	
24	Pig	B	83 <sup>4</sup>		22 <sup>4</sup>	
25		C	63 <sup>3</sup>		37 <sup>3</sup>	
26		S	92 <sup>3</sup>		8 <sup>3</sup>	
27	Rabbit	B	86 <sup>4</sup>		19 <sup>4</sup>	
28		C	70 <sup>4</sup>		40 <sup>4</sup>	
29		S	93 <sup>3</sup>	92-94 <sup>3</sup>	6.7 <sup>3</sup>	6.0-7.8 <sup>3</sup>
30	Rat	B	86 <sup>4</sup>	84-87 <sup>4</sup>	20 <sup>4</sup>	19-22 <sup>4</sup>
31		P	95 <sup>4</sup>		7.8 <sup>4</sup>	
32	Sheep	B	87 <sup>4</sup>		19 <sup>4</sup>	
33		C	67 <sup>4</sup>		42 <sup>4</sup>	
34		S	92 <sup>3</sup>		8.3 <sup>3</sup>	

/1/ Determined by oven-drying methods unless specified otherwise. /2/ 75 (73-76) g/100ml RBC, and 95 (95-96) g/100ml plasma by distillation method. /3/ As grams per 100 grams. /4/ Source data presented in terms of g/100 g; calculated to grams per 100ml by application of values for specific gravity of blood, plasma, or cells. Solids calculated as (100-water content)% by weight, then calculated to g/100ml as above. /5/ Parentheses enclose values calculated on basis of standard hematocrit values and contents in cells and plasma.

# 68. BLOOD CARBOHYDRATES AND RELATED SUBSTANCES

Blood (B), R B C (C), Plasma (P), Serum (S), Leukocytes (L), Celomic fluid (F)

Constituent	Animal	mg/100ml		Constituent	Animal	mg/100ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
1	Man, fasted	B 90	80-200 <sup>b</sup>		Opossum	C 81.3 <sup>8</sup>	73-89 <sup>b</sup> , 3.8
2	Man, fasted	B 86	76-96 <sup>b</sup>		Opossum	P 123.3 <sup>8</sup>	107-139 <sup>b</sup> , 3.8
3	Man	C 74.3 <sup>4</sup>	46-102.3 <sup>4</sup>		Ox	C 15.3 <sup>4</sup>	
4	Man	S 97.3 <sup>4</sup>	61-150 <sup>b</sup> , 3.4		Ox	S 85.3 <sup>4</sup>	
5	Man, newborn	B 34	20-30 <sup>b</sup> , 5.6		Pig	C 0.3 <sup>4</sup>	
6	Man, fetus	B 35	36-116 <sup>b</sup> , 7		Pig	S 90.3 <sup>4</sup>	
7	Calf	C 48.3 <sup>4</sup>			Rabbit	B 85.3 <sup>4</sup>	67-107 <sup>b</sup> , 11
8	Calf	S 118.3 <sup>4</sup>			Rabbit	C 41.3 <sup>4</sup>	
9	Camel	B 48			Rabbit	S 145.3 <sup>4</sup>	
10	Camel	S 108			Rat, Wistar	H 66.5 <sup>4</sup> , 10	56-76 <sup>b</sup> , 5.10
11	Cat, fasted	B 74.3 <sup>8</sup>	64-84 <sup>b</sup> , 3.8		Rat, Yale	H 85.3 <sup>4</sup> , 10	78-93 <sup>b</sup> , 5.10
12	Cat	C 76.3 <sup>4</sup>			Sheep	H 35.5 <sup>4</sup> , 10	18-57 <sup>b</sup> , 5.10
13	Cat	S 297.3 <sup>7</sup>			Sheep	C 10.3 <sup>4</sup>	
14	Cattle	B 46.9	36-57 <sup>9</sup>	Glucose	Sheep	S 80.3 <sup>4</sup>	
15	Dog	B 60.9	44-75 <sup>9</sup>		Sheep, fetus	H 20.2	
16	Dog, fasted	B 77.5 <sup>6</sup>	67-87 <sup>b</sup> , 5.6		Chicken	H 170.5 <sup>10</sup>	126-204 <sup>b</sup> , 5.10
17	Dog	C 41.3 <sup>4</sup>			Duck	H 114.5 <sup>11</sup>	90-138 <sup>b</sup> , 5.11
18	Dog	S 132.3 <sup>4</sup>			Owl	H 152.5 <sup>11</sup>	200-350 <sup>b</sup> , 5.10
19	Fox	B 104	70-134		Pigeon, fasted	H 99.5 <sup>13</sup>	117-189 <sup>b</sup> , 5.11
20	Guinea pig, fasted	B 95.5 <sup>10</sup>	60-125 <sup>b</sup> , 5.10		Alligator, fasted	H 25-111 <sup>b</sup>	21-205 <sup>b</sup> , 13
21	Guinea pig	B 96.5 <sup>10</sup>	82-107 <sup>b</sup> , 5.10		Philodryas	B 63	46-88 <sup>b</sup>
22	Guinea pig	C 53.3 <sup>4</sup>			Pit viper, male	B 67	22-98 <sup>b</sup>
23	Guinea pig	S 155.3 <sup>4</sup>			Pit viper, female	B 60	36-49 <sup>b</sup>
24	Horse	B 73 <sup>9</sup>	54-95 <sup>9</sup>		Frog	B 42	
25	Mink	B 122	72-172 <sup>b</sup>		Carp	B 98-256	
26	Monkey	C 119.3 <sup>4</sup>			fresh water	B 44-141	
27	Monkey	S 148.3 <sup>4</sup>			Eel	B 0.2	
28	Mouse, fasted	B 109.3 <sup>8</sup>	75-143 <sup>b</sup> , 3.8		Gastrophilus		
29	Mouse	B 174.3 <sup>8</sup>	82-266 <sup>b</sup> , 3.8		intestinalis, larva		

1/1 Arterial blood. 2/2 Venous blood. 3/3 Tungstic acid filtrate. 4/4 Fermentation. 5/5 Zinc hydroxide filtrate. 6/6 Ferrihydride-iodometric titration. 7/7 Fermentable sugar in cord blood. 8/8 Color formation with ferric ferrocyanide. 9/9 "True blood sugar" substance". 10/10 Copper-iodometric titration. 11/11 Color formation with copper arsenomolybdate. 12/12 "Non-fructose, fermentable reducing substance". 13/13 Copper molybdate color formation.

# 68. BLOOD CARBOHYDRATES AND RELATED SUBSTANCES (Continued)

		Blood (B), R B C (C), Plasma (P), Serum (S), Leukocytes (L), Cerebrospinal fluid (F)					
Constituent	Animal	mg/100ml		Constituent	Animal	mg/100ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
57 Arencola	P	12		Glycogen 10,19	Man	0	0.5-11.2
58 Dasybranchus	F	8	2.3-8.7		Horse	3.3	
59 Spingulus	F		7.5-26		Dog	3.1	
60 Carinus	P	0			Man	67	61-78
61 Carinus, fasted	P				Man, aged	81	70-89
62 Cancer	P	22	1-4	Glucosamine 20	Man, 3-8 yr	63	52-69
63 Crayfish	P				Man, fetus	48	42-55
64 Bombyx	P	16			Rabbit	71	51-91
65 Hydrophilus	P		6-31		Man	102.2	73-131.2
66 Mollusc	P		2.1-8.7		Man	111	93-126
67 Man, fetus	S	7.5-15		Polysaccharide 21	Man, aged	129	104-138
68 Sheep, fetus	B	70	0-1		Man, 3-8 yr	105	94-118
69 Gastrophilus	B		12-128.6		Man, fetus	80	62-103
70 Intestinalis, larva	B	171.5			Rat	167	2.6-4.8
71 pupa, 10-11th day	B	22.5		Pentose 23,24	Man	3.7	2.6-4.8
72 Blowfly	B	5.17		total	Rat, fasted	5.4	4.4-6.4
73 Grasshopper	B	0.17			Rabbit, fasted	5.3	2.9-7.7
74 Waxmoth, larva	B	0.17		Pentose,	Man	2.1	
75 Man	B		0-trace	phosphorylated	Rabbit	2.9	1.5-4.2
76 Woman, pregnant	P		0-trace		Man	6.7	4.1-9.3
77 Lactose 3,13,18	P				Man	0.6	
	P				Man	0.8	
	P				Man	0.9	
	P				Rat	0.4	0.5-1.3
	P				Rabbit	1.4	0.0-3.0
	P				Rabbit	0.7	0.0-1.9
78 Man	B	5.5	0-2				
79 Man	L	2.5	1.2-16.2				
80 Man	P	0	1.0-1.8				

14/ "Fermentable reducing sugar." 15/ Color development with diphenylamine. 16/ Cadmium hydroxide filtrate. 17/ Color development with resorcinol. 18/ Destruction of non-lactose sugar by B. proteus. 19/ Cells hemolyzed in water, digested with KOH; precipitation with alcohol. hydrolysis. 20/ Color development with acetylacetone on hydrolysate of alcohol precipitate. 21/ Absorption at 500 mμ by product of tryptophan reaction with acidified alcohol precipitate. 22/ Carbazole reaction with alcohol-precipitated, acid-treated serum. 23/ Trichloroacetic acid filtrate. 24/ Quantitative orcinol reaction. 25/ 2.6N saline extract of acetone precipitate. 26/ As glucuronic acid, color development with naphtho-resorcinol, with preliminary hydrolysis but without deproteinization, unless specified otherwise. 27/ Tungstate-sulfuric acid filtrate.



## 68. BLOOD CARBOHYDRATES AND RELATED SUBSTANCES (Continued)

# 68. BLOOD CARBOHYDRATES AND RELATED SUBSTANCES (Concluded)

Blood (B), R B C (C), Plasma (P), Serum (S), Leukocytes (L), Cerebro fluid (F)									
Constituent	Animal	mg/100ml		Constituent	Animal	mg/100ml			
		Value (C)	Range (D)			Value (C)	Range (D)		
(A)	(B)			(A)	(B)				
165	Rat	P	146	189	Man	B	1.3		
166	Sheep	C	<3.5	190	Man, infant	B	0.8		
167	Birds	C	0	191	Dog	B	1.0		
168	Reptiles	C	0	192	Rabbit	B	1.0		
169	Fish	C	0	193	Rat	B	1.1		
170	Mammals	C	0	194	Pigeon	B	0.9		
171	Chicken	C	251	195	Alpha-ketoglutaric acid	B	0.2-1		
172	Duck	C	264	196	Man	P	0.7-1		
173	Pigeon	C	285	197	Man, adult	S	2.4-5		
174	Turtle	C	95	198	Man, child	S	2.8-5		
175	Man	B	24-39	199	Man, newborn	S	3-6		
176	Man	C	12-39	200	Man	B	1.9-4		
177	Man	P	36-39	201	Man	P	2.3-4		
178	Man	B	19-40	202	Malic acid	P	0.1-0.9		
179	Dog	B	19-39	203	Fumaric acid	B	<0.3-48		
180	Dog	C	22.5-39	204	Succinic acid	P	0.5		
181	Rat	B	13-39	205	Ascorbic acid <sup>49</sup>	P	0.5		
182	Sheep	B	21-39						
183	Carp	B	16-39						
184	Man	B	0.3-1						
185	Man	P	1.2-4						
186	Guinea pig	P	2.3						
187	Rabbit	P	1.9-4						
188	Rat	P	1.9-4						

/37/ Color formation with naphtho-resorcinol after precipitation by lead acetate. /38/ Precipitation as calcium salt, determination of phosphorus. /39/ Tungstic acid filtrate, oxidation by Mn+++ to acetaldehyde and titration of bound bisulfite with iodine. /40/ Conversion to acetaldehyde by concentrated sulfuric acid, color formation with para-hydroxy diphenyl; cutaneous blood from finger tip. /41/ Separation of dinitrophenylhydrazones by chromatography prior to color formation with sodium carbonate. /42/ Color formation with sodium carbonate. /43/ Formation and extraction of dinitro-phenylhydrazones; color formation with sodium carbonate. /44/ Oxidation to succinic acid, manometric measurement with succinic dehydrogenase. /45/ Formation of penta-bromo-acetone, color formation with sodium sulfide. /46/ Measurement of bromine in penta-bromo-acetone by reaction of bromate with iodine. /47/ Formation of fluorescent compound with orcinol. /48/ Chromatographic separation in column of silica gel, titration with alkali. /49/ See table on Blood Vitamin Content.

# 69. BLOOD LIPIDS

Blood (B), RBC (C), Plasma (P), Serum (S)

Constituent	Animal	Lipids mg/100 ml		Constituent	Animal	Lipids mg/100 ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
1	Lipids, total <sup>1,2</sup>	Man	B (559) <sup>3</sup>	32	Phospholipid <sup>2</sup>	Rabbit	B (145) <sup>3</sup>
2		Man	C 596	33		Rabbit	C 240
3		Man	P 530	34		Rabbit	P 78
4		Cat	P 376	35		Rat	P 83
5		Chicken	P 520	36		Sheep	P 90
6		Cow	P 348	37	Lecithin	Man	B (115) <sup>3</sup>
7		Dog	P 580	38		Man	C (70) <sup>6</sup>
8		Goat	P 300	39		Man	P 117 <sup>5</sup>
9		Guinea pig	P 169	40		Man	S 107
10		Rabbit	P 243	41		Cow	S 54
11	Neutral fat <sup>2</sup>	Rat	P 230	42		Dog	P 370
12		Man	B 134	43		Dog	S 288
13		Man	C 93	44		Pig	S 72
14		Man	P 142	45		Rabbit	C 85
15		Cat	P 108	46	Cephalin	Man	B 65
16		Chicken	P 225	47		Man	C (210) <sup>6</sup>
17		Cow	P 105	48		Man	P 7 <sup>5</sup>
18		Guinea pig	P 73	49		Cow	S 3
19		Rabbit	B (78) <sup>3</sup>	50		Dog	P 22
20		Rabbit	C 41	51		Pig	S 3
21		Rabbit	P 105	52		Rabbit	B (60) <sup>3</sup>
22		Rat	P 85	53		Rabbit	C 107
23	Phospholipid <sup>2</sup>	Man	B (247) <sup>3</sup>	54		Rabbit	P 27
24		Man	C 350 <sup>4</sup>	55	Sphingomyelin	Man	B 186
25		Man	P 165	56		Man	C (70) <sup>6</sup>
26		Cat	P 132	57		Man	P 41 <sup>5</sup>
27		Chicken	P 155	58		Cow	S 22
28		Cow	P 84	59		Dog	S 55
29		Cow	S 80	60		Pig	S 21
30		Guinea pig	P 51	61		Rabbit	B (42) <sup>3</sup>
31		Pig	P 96	62		Rabbit	C 47
				63	Cholesterol <sup>2</sup>	Rabbit	P 38
				64		Man	B
				65		Man	C 173
				66		Man	P 152

(Continued on the next page)

/1/ Total lipids = Cholesteryl esters + Free Cholesterol + Phospholipid + Neutral Fat. /2/ Determined by oxidimetric methods on oxalated blood of fasted animals, unless specified otherwise. /3/ Parentheses ( ) enclose values calculated on basis of hematocrit and content of constituent in blood, cells or plasma. /4/ Phospholipid P (determined by method of Blomgren and al.). /5/ Phospholipid P (determined by method of Blomgren and al.). /6/ Values calculated on basis of following probable approximate partition of total phospholipid in human red blood cells: cephalin, 60%, lecithin, 20%, sphingomyelin 20%.

# 69. BLOOD LIPIDS (Concluded)

# IDS (Concluded)

Constituent		Animal	Blood (B), RBC (C), Plasma (P), Serum (S)				Constituent	Animal	Lipids mg/100 ml														
(A)	(B)		Lipids mg/100 ml		Value	Range			(C)	Value	Range	(D)											
			(C)	(D)																			
67	Cholesterol <sup>2</sup>	Chicken	P	100		52-148b	98	Cholesterol, esterified	Chicken	P	66	26-106b											
68		Cat	P	93		43-143b	99		Cow	P	73	25-121b											
69		Dog	P	110		8-212b	100		Guinea pig	P	21	35-55											
70		Goat	P	173		138-214d	101		Horse	S	30	12-30b											
71		Guinea pig	P	32		46-95d	102		Rabbit	B	30	(0-28) <sup>3</sup>											
72		Horse	S	77		21-43b	103		Rabbit	C	0												
73		Monkey	S	118			104		Rabbit	P	22												
74		Mouse, male	B	1808			105		Rat	P	31	0-47b											
75		Mouse, female	B	148 <sup>8</sup>		149-244 <sup>8</sup>	106		Man	B	0	11-51b											
76		Rabbit	B	(82) <sup>3</sup>		132-166 <sup>8</sup>	107		Man	C	0	79-194											
77	Cholesterol, free	Rabbit	C	133		(54-110) <sup>3</sup>	108	Cholesteryl esters <sup>7</sup>	Man	C	0												
78		Rabbit	P	45		115-151b	109		Man	P	179	127-232											
79		Rat	P	52		10-80b	110		Cat	P	106	25-188											
80		Man	B	140		82-113	111		Chicken	P	112	44-179											
81		Man	P	46		119-161b	112		Cow	P	123	42-204											
82		Cat	P	30		30-62b	113		Dog	P	66	59-93											
83		Chicken	P	34		9-51b	114		Guinea pig	P	15	9-22											
84		Cow	P	37		15-53	115		Horse	S	51												
85		Dog	P	134		0-85b	116		Rabbit	C	0												
86		Guinea pig	P	11		103-159d	117		Rabbit	P	37												
87	Cholesterol esterified	Horse	S	47		7-15b	118	Fatty acids, 2 total	Rat	P	52	0-79											
88		Rabbit	B	(68) <sup>3</sup>		(48-90) <sup>3</sup>	119		Man	P	316	19-86											
89		Rabbit	C	133		115-151b	120		Cat	P	228	149-483b											
90		Rabbit	P	22		0-47b	121		Chicken	P	361	56-400b											
91		Rat	P	21		5-37b	122		Cow	P	202	206-516b											
92		Man	B	0		47-115	123		Guinea pig	P	116	26-378b											
93		Man	C	0			124		Rat	P	169	92-140b											
94		Man	P	106			125		Rabbit	P	153	40-298b											
95		Cat	P	73		75-137b	126					108-198b											
96						25-121b	127																
97							128																

/7/ Cholesteryl esters = 1.69 x esterified cholesterol  
 modification

/8/ Determined by Sperry-Schoenheimer method, or  
 from neutral fats = 0.95 x neutral fats  
 from phospholipids = 0.65 x Phospholipids  
 from cholesteryl esters = 0.43 x cholesteryl esters

/7/ Cholesteryl esters = 1.69 x esterified cholesterol modification

from neutral fats = 0.95 x neutral fats  
from phospholipids = 0.65 x Phospholipids  
from cholesteryl esters = 0.43 x cholesteryl esters

/8/ Determined by Sperry-Schoenheimer method, or

70. BLOOD PROTEINS: MAN

Constituent	Blood		Erythrocytes		Plasma <sup>1</sup>	
	Value	Range	Value	Range	Value	Range
	g/100ml (B)	g/100ml (C)	g/100ml (D)	g/100ml (E)	g/100ml (F)	g/100ml (G)
1 Protein, total	20.5		36.8		7.41	
2 Hemoglobin	14.9 <sup>2</sup>		33.5 <sup>3</sup>	30-40 <sup>b</sup>	trace	0.001-0.005
3 Corpuscle stroma	1.5 <sup>3</sup>		3.3			
4 Plasma protein	4.1 <sup>3</sup>		—	—	7.41	
5 Fibrinogen	0.16 <sup>3</sup>		—	—	0.28	0.23-0.36 <sup>d</sup>
6 Serum protein	3.9 <sup>3</sup>		—	—	7.13	6.4-7.9 <sup>b</sup>

/1/ Salt fractionation methods. /2/ Average of male, value = 15.8 (range = 14-18<sup>b</sup>) and female, value = 13.9 (range = 11.5-16<sup>b</sup>). /3/ Calculated on the basis of hematocrit value of 44.5% (see Table 37).

# 71. PLASMA PROTEINS: MAN

Constituent	Salt Fractionation Method <sup>1</sup>			Salt Fractionation Method <sup>2</sup>			Electrophoretic Method			Electrophoretic Meth. (Corrected) <sup>3</sup>		
	Value		Range	Value		Range	Value		Range	Value		Range
	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	
(A)	100	7.48		100	7.41		100	7.30		100	6.38	
1 Plasma protein, total												
2 Fibrinogen	3.7	0.28 <sup>4</sup>	0.23-0.36 <sup>4</sup>	3.8	0.28 <sup>4</sup>	0.23-0.36 <sup>4</sup>	4.0	0.29		4.7	0.31	
3 Serum protein, total	96.3	7.2	6.5-7.9	96.2	7.13	6.4-7.9 <sup>b</sup>	96.0	7.01	6.3-7.7 <sup>b</sup>	95.3	6.27	
4 Albumin	59.5	5.2	4.7-5.7	61.8	4.58	4.1-5.1 <sup>b</sup>	60.5	4.42	4.0-4.8 <sup>b</sup>	50.1	3.30	
5 Globulin, total	26.8	2.0	1.3-2.5	34.4	2.55	3.8-3.3 <sup>b</sup>	35.5	2.59		45.2	2.97	
6 Alpha <sub>1</sub> -globulin							9.0	0.29	0.2-0.4 <sup>b</sup>			
7 Alpha <sub>2</sub> -globulin							7.7	0.56	0.4-0.7 <sup>b</sup>	14.0		
8 Beta-globulin							11.5	0.94	0.8-1.1 <sup>b</sup>	19.3	1.27	
9 Gamma-globulin							12.3	0.90	0.6-1.2 <sup>b</sup>	11.9	0.78	
10 Euglobulin	2.7	0.2	-0.4	21.4	1.59	0.9-2.2 <sup>b</sup>						
11 Pseudoglobulin, total	24.1	1.8		13.0	0.96	0.6-1.35 <sup>b</sup>						
12 Pseudoglobulin I	17.4	1.3	0.8-1.9									
13 Pseudoglobulin II	6.7	0.5	0.2-0.8									
14 Albumin/globulin ratio		2.6			1.8				1.71	1.3-2.2 <sup>b</sup>		1.1

/1/ Method of Howe, P.E., 1921. /2/ Micro-Major method, 1947. /3/ Corrected for nitrogen factor, refractive increment differences, ionic strength, total protein concentration effects. Red Cross pooled plasma used, corrected for dilution with citrate. /4/ Value not from Milne-Major or Howe. See bibliography.

# 72. PLASMA PROTEINS<sup>1</sup>: ANIMALS OTHER THAN MAN

Animal	Plasma Protein Total		Fibrinogen		Serum Protein Total		Albumin		Globulin	
	g/100ml		g/100ml		g/100ml		g/100ml		g/100ml	
	Value (H)	Range (C)	Value (D)	Range (E)	Value (F)	Range (G)	Value (H)	Range (I)	Value (J)	Range (K)
1 Cat										
2 Chicken	8.32	7.4-10.2			7.58	6.1-9.0 <sup>b</sup>	4.01	3.5-4.6 <sup>b</sup>	3.57	2.1-5.0 <sup>b</sup>
3 Cow	6.72	6.1-7.6	0.72		3.60	2.6-4.6	1.82		1.78	
4 Dog	7.27		0.52	0.42-0.64	7.602		3.632,3		3.972,3	
5 Goat	4.70		0.60		6.202	4.9-7.9	3.572,4	3.1-4.0	2.632,4	2.0-3.3
6 Guinea pig	6.84		0.33		6.672		3.962,5		2.712,5	
7 Horse	5.82		0.34	0.29-0.43	4.37		3.03		1.34	
8 Monkey			0.32	0.25-0.40	6.502		3.252,6		3.252,6	
9 Mouse					5.50	5.84-6.20 <sup>b</sup>	4.50		1.52	
10 Rabbit	6.29		0.25	0.16-0.34 <sup>b</sup>	6.02		4.202,7		1.502,7	
11 Rat	5.74		0.36		5.702	5.5-6.5 <sup>b</sup>	3.86	3.4-4.3 <sup>b</sup>	2.18	1.8-2.5 <sup>b</sup>
12 Sheep					6.04		3.07		2.31	
13 Swine					6.30		2.03		3.27	

<sup>1/1</sup> The data represent chemical methods (salting-out, colorimetric, etc.); in most instances 6.25 used as Nitrogen factor in calculating protein. "Fraction V," precipitated with the albumin in the Howe salting-out method, should be subtracted from the albumin and added to the globulin to permit comparison with results of later methods. <sup>2/2</sup> Total serum protein partitioned into albumin and globulin by means of average of measured A/G ratios. <sup>3/3</sup> A/G = 0.92. <sup>4/4</sup> A/G = 1.46. <sup>5/5</sup> A/G = 1.0. <sup>6/6</sup> A/G = 2.80.

### 73. PLASMA PROTEINS, CERTAIN PROPERTIES AND REACTIONS: MAN

Classification	Purified Protein Component	Electrophoretic Fraction	Interacts with	General Properties
(A)	(B)	(C)	(D)	(E)
1 Blood coagulation agents	Antihemophilic globulin			Clots hemophilic blood
2	Fibrinogen		Thrombin	Forms fibrin clot
3	Plasmin		Proteins	Dissolves plasma clot
4	Plasminogen		Streptokinase	Enzyme precursor
5	Prothrombin		Thromboplastin	Catalyzes formation of fibrin clot
6	Thrombin		Fibrinogen	Antibodies for pathogenic organisms
7 Serological agents	Antibody gamma-	Gamma-globulin	Antigens	Contain typhoid "O" agglutinins
8	Diphtheria antitoxin	Gamma-globulin	Antigens	Anti-A, anti-B, & anti-RH agglutinins
9	Antibody euglobulin	Beta- & gamma-globulin	Antigens	
10	Isoagglutinins	Beta- & gamma-globulin	Antigens of incompatible red blood cells	
11	Complement components C'1, C'2	Alpha- & beta-globulin	Antigen-antibody complex	
12 Enzymes <sup>1</sup>	Choline esterase		Choline esters	
13	Peptidase		1-Leucylglycylglycine	
14	Phosphatase alkaline		Phosphoric acid monoesters	
15	Plasmin (see 3A-E)			
16	Thrombin (see 6A-E)			
17	Iodoglobulin	Alpha-globulin		
18	Thyrotrophic hormone	Alpha-globulin		Thyroid-active protein
19	Albumin	Albumin	Fatty acids, bile salts, dyes, & drugs	Influences thyroid activity
20	Mercaptalbumin	Albumin	Mercury	Osmotic regulation of blood volume
21	Alpha <sub>1</sub> -lipoprotein	Alpha <sub>1</sub> -globulin	Steroids	Have sulfhydryl groups
22	Alpha <sub>1</sub> -bilirubin globulin	Alpha <sub>1</sub> -globulin	Bilirubin	35% lipid
23	Alpha <sub>2</sub> -glycoproteins	Alpha <sub>2</sub> -globulin	Carbohydrates & barium	
24	Alpha <sub>2</sub> -muproteins	Alpha <sub>2</sub> -globulin	Carbohydrates & barium	
25	Beta <sub>1</sub> -lipoproteins	Beta <sub>1</sub> -globulin	Estriol, carotenoids, & other steroids	75% lipid, carrier for vitamins & hormones
26	Beta <sub>1</sub> -metal-combining protein <sup>2</sup>		Iron & Copper	Binds 2 Fe <sup>+++</sup> or Cu <sup>++</sup> per molecule <sup>3</sup>
27	Beta <sub>1</sub> -lipid-poor euglobulins			
28	Ceruloplasmin		Copper	

<sup>1</sup>/ Other enzymes include acid phosphatase, amylase, tributyrin esterase, beta-glucuronidase  
<sup>2</sup>/ Blends iron much more strongly than copper

<sup>3</sup>/ = Siderophilin, trans-



74. PLASMA PROTEINS, PHYSICAL PROPERTIES: MAN

Purified Component	Concentrated in Fraction	Estimated Amount g/100 g Plasma Protein	Approx. Isoelectric Point pH	Molecular Weight M	Sedimentation Constant S <sub>20,w</sub>	Specific Volume V	Intrinsic Viscosity $\eta_{sp}/c$ , 10 <sup>-2</sup>	Frictional Ratio $f/f_0$	Approx. Dimensions Angstrom Units	
									Length	Diameter
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)
1 Antihemophilic globulin	I	0.15	<5.3							
2 Cold-insoluble globulin <sup>1</sup>	I-1	4	<5.3	400,000	9		25	1.98	700	38
3 Fibrinogen	I-2		6.3	156,000	7	0.739	6	1.38	235	44
4 Gamma-globulins	II	11 { 9	7.3	300,000	10	0.739				
5 Diphtheria antitoxin	II	0.001								
6 Beta <sub>1</sub> -lipoprotein	III-0	5	5.4	1,300,000	7	0.950	4.1		185	185
7 Beta <sub>1</sub> -lipid-poor euglobulins	III-0	3 { 2	5.5		7					
8 Cereuloplasmin	III-0	3 { 1	5.5		20					
9 Isoagglutinins	III-1		4.4							
10 Antibody euglobulins	III-1		6.3							
11 Prothrombin	III-2	0.03								
12 Complement components C'1, C'2	III-2&IV	0.1								
13 Plasminogen	III-3	0.4								
14 Plasmin	III-3									
15 Beta <sub>2</sub> -globulins	III		6.3							
16 Alpha <sub>1</sub> -lipoprotein	IV-1	3	5.2	200,000	7	0.841	6.6	1.38	300	50
17 Thyrotrophic hormone	IV-4	3			5					
18 Alpha <sub>2</sub> -glycoprotein	IV-6	1.2	4.9		9					
19 Alpha <sub>2</sub> -mucoprotein	IV-6	0.5	4.9		9					
20 Cholinesterase	IV-6	0.005	4.5		9					
21 Peptidase	IV									
22 Phosphatase, alkaline	IV									
23 Iodoglobulin	IV-6									
24 Beta <sub>1</sub> -metal combining protein	IV-7	3	5.8	90,000	5	0.725	5.5	1.37	190	37
25 Albumin, serum	V	52	4.9	69,000	4.6	0.733	4.2	1.28	150	38
26 Mercaptalbumin	V	34								
27 Alpha <sub>1</sub> -bilirubin globulin	V-1	0.05	4.7							
28 Alpha <sub>2</sub> -protein	VI-1	0.1								
29 Beta <sub>1</sub> -protein	VI-1	0.05								
30 Alpha <sub>1</sub> -small acid protein <sup>2</sup>	VI-2	0.5	3.0		2.9					
31					5					
32					3.5					

/1/ Non-clottable protein, insoluble at low temperature. /2/ Acid glycoprotein.

# 75. BLOOD FREE AMINO ACIDS<sup>1</sup>

Constituent	Animal	100 ml Blood mg		100 ml RBC mg		100 ml Plasma <sup>2</sup> mg	
		Value	Range	Value	Range	Value	Range
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
Alanine	Man	4.0	2.8-5.2 <sup>b</sup>	4.0	2.5-5.6 <sup>b</sup>	4.0 <sup>3</sup>	2.6-5.3 <sup>b</sup>
	Mouse					5.9	5.3-6.6
	Rat	12.3	9.0-15.6 <sup>b</sup>				
Arginine	Man	1.0	0.6-1.7 <sup>b</sup>	0.3	0.1-0.6 <sup>b</sup>	2.3	1.1-3.5 <sup>b</sup>
	Dog	3.7	1.7-5.2	4.2		3.3 <sup>3</sup>	1.8-4.8 <sup>b,4</sup>
	Mouse					0.97	0.9-1.0
	Rat					3.9	2.7-4.9 <sup>b,5</sup>
Aspartic acid	Man						0.9-1.2
Citrulline <sup>6</sup>	Man						0.3-1.0
	Dog						0.8-1.5
Cysteine	Horse	0.3		0.5		0.2	
	Man	0.9	0.6-1.2 <sup>b</sup>	0.4	0.3-0.5 <sup>b</sup>	1.4	0.8-2.0 <sup>b</sup>
	Dog					0.9	0.5-1.5
	Horse					1.6	
	Pig						0.6-0.9
	Rabbit						0.9-1.1
	Rat						0.7-0.9
Glutamic acid	Man					0.8 <sup>7,8</sup>	0.6-1.7 <sup>7</sup>
	Cat					2.1	
	Dog					<0.6	
	Mouse					3.3	2.9-3.6
	Man		1.8-2.5	2.4	1.6-3.1 <sup>b</sup>	1.8	1.3-2.3
	Guinea pig					2.5	
	Mouse						
	Rabbit					1.9	1.7-2.3
	Rat					4.0	
						3.1	1.5-4.6 <sup>b,5</sup>
Histidine	Man	1.3	0.9-1.7	1.1	0.8-1.6	1.4	1.1-1.8 <sup>b</sup>
	Dog	1.3	1.0-2.0			1.2	0.9-1.6 <sup>b</sup>
	Mouse					1.6	1.4-1.7
	Rat					1.2	0.9-1.5 <sup>b,5</sup>
Isoleucine	Man	1.3	0.9-1.5	0.9	0.5-1.4	1.6	1.0-2.2 <sup>b</sup>
	Dog	1.8	1.2-2.2 <sup>b</sup>			1.3	0.5-2.1 <sup>b</sup>
	Mouse					1.5	1.2-2.0 <sup>b</sup>
	Rat					1.4	0.7-2.5 <sup>b,5</sup>
Leucine	Man	1.7	1.4-2.0 <sup>b</sup>	1.5	1.0-1.8 <sup>b</sup>	1.9	1.3-2.5 <sup>b</sup>
	Dog	2.5	1.2-3.6			2.1	1.6-2.8
	Mouse					2.4	2.2-2.8
	Rat					2.1	1.1-3.1 <sup>b,5</sup>
Lysine	Man	2.2	1.3-3.0 <sup>b</sup>	1.4	0.9-1.8 <sup>b</sup>	3.0	2.1-3.8 <sup>b</sup>
	Dog	2.5	1.6-3.6			2.4	1.3-3.6 <sup>b</sup>
	Goat					1.8	1.3-2.4
	Mouse					6.4	5.7-7.0
	Rat					7.2	4.0-10.4 <sup>b,5</sup>
Methionine	Man	0.5	0.4-0.6 <sup>b</sup>	0.5	0.3-0.8 <sup>b</sup>	0.5	0.3-0.6 <sup>b</sup>
	Dog	1.2	0.8-1.7				0.2-1.9
	Mouse					1.9	1.7-2.2
	Rat					1.0	0.6-1.3 <sup>b,5</sup>

Note See footnotes on next page

(Continued on the next page)

# 75. BLOOD FREE AMINO ACIDS<sup>1</sup> (Concluded)

Constituent	Animal	100 ml Blood mg		100 ml R B C mg		100 ml Plasma <sup>2</sup> mg	
		Value	Range	Value	Range	Value	Range
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
48 Phenylalanine	Man	1.0	0.8-1.2 <sup>b</sup>	1.0	0.7-1.3 <sup>b</sup>	1.4	0.5-2.2 <sup>b</sup>
49	Dog	1.5	0.9-2.5			1.2	
50	Mouse					2.4	2.0-3.2
51	Rat					1.1	0.7-1.5 <sup>b,5</sup>
52 Proline	Man						2.4-2.7
53	Mouse					1.8	1.6-2.1
54	Rat					3.6	3.3-3.9 <sup>b,5</sup>
55 Threonine	Man	1.6	1.3-2.0 <sup>b</sup>	1.6	1.3-2.1 <sup>b</sup>	2.0	1.1-2.6 <sup>b</sup>
56	Dog	2.4	1.2-3.3			2.6	1.2-4.0 <sup>b</sup>
57	Mouse					3.5	3.0-3.9
58	Rat					4.2	2.3-6.2 <sup>b,5</sup>
59 Tryptophan	Man	0.7	0.5-1.0 <sup>b</sup>	0.24	0.2-0.4 <sup>b</sup>	1.1	0.7-1.5 <sup>b</sup>
60	Cow					1.1	0.8-1.2
61	Dog	1.2	0.6-2.4			1.2	0.8-1.5
62	Horse					1.2	
63	Mouse					2.1	1.5-2.6
64	Pig					1.1	1.0-1.2
65	Rat	1.6	1.5-2.0			1.5	0.8-2.1 <sup>b,5</sup>
66	Sheep					1.1	0.8-1.5 <sup>b</sup>
67 Tyrosine	Man	1.1	0.8-1.4 <sup>b</sup>	1.1	0.7-1.5 <sup>b</sup>	1.5	0.8-2.2 <sup>b</sup>
68	Dog	1.2	0.7-2.0			1.1	0.6-1.5 <sup>b</sup>
69	Goat					1.2	1.0-1.5
70	Mouse					2.5	2.4-2.7
71	Rat					1.5	0.8-2.2 <sup>b,5</sup>
72 Valine	Man	2.4	2.0-2.9 <sup>b</sup>	2.0	1.6-2.5 <sup>b</sup>	2.8	2.2-3.5 <sup>b</sup>
73	Dog	2.6	1.5-4.1			2.2	1.2-3.3 <sup>b</sup>
74	Mouse					4.3	3.8-5.0
75	Rat					2.6	1.5-3.6 <sup>b,5</sup>

for glutamine. <sup>1</sup> - method, corrected

## 76. BLOOD NON-PROTEIN NITROGEN COMPOUNDS

Blood (B), RBC (C), Plasma (P), Serum (S).

Constituent	Animal			Non-Protein N Compounds per 100 ml					
	(A)	(B)	(C)	(D)	Non-Protein N Compounds per 100 ml				
					Value mg	Range mg			
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)		
1 Amino Acids <sup>1</sup>	Man	B	50	38-53	19 Nucleotides	Man	B	41	31-52
2 Ammonia	Man	B	0.18	0.12-0.24	20 Polypeptides	Man	B	<7	
3 Bilirubin	Man	B	0.18	0.1-0.25		Man	B	32	24-40 <sup>b</sup>
4	Man	B	3.9	2.9-4.9 <sup>b</sup>		Man	C	30	25-39 <sup>b</sup>
5 Creatine	Man	C	8.1	6.0-10.2 <sup>b</sup>		Man	P	34	28-40 <sup>b</sup>
6	Man	P	0.23	0.0-0.8 <sup>b</sup>	Urea	Man ♂	B	33	26-46
7	Chicken	B	2.1			Man ♂	P	27	19-36
8	Man	B	1.5	1.0-2.0		Man ♀	B	24	11-29
9 Creatinine	Man	C	1.8	1.7-1.9 <sup>b</sup>		Rat	P	34	
10	Man	P	0.91	0.87-0.95 <sup>b</sup>		Chicken	B	5.7	
11 Ergothioneine <sup>2</sup>						Man	B	3.2	2.2-4.2 <sup>b</sup>
12	Man	P		5.0-12.0		Man	C	1.9	0.8-3.0 <sup>b</sup>
13 Glutamine	Dog	C		3.8-7.2		Man	P	3.8	2.0-5.6 <sup>b</sup>
14	Dog	P		7 -13	Uric Acid	Man ♂	S	4.8	
15	Horse	C	7.2			Man ♀	S	4.0	
16 Glutathione <sup>2</sup>						Dog	P	0.33	
17 Lecithin <sup>3</sup>						Rat	B	1.5	0.5-3.4
18 Methyl guanidine	Man	B	0.25	0.2-0.3	Vitamins <sup>4</sup>	Chicken	B	4.5	

/1/ For details see table on Blood Free Amino Acid Content. /2/ See table on Blood Sulfur Compounds. /3/ See table on Blood Lipid Content. /4/ See table on Blood Vitamin Content.

# 77. NITROGEN CONTENT OF BLOOD NITROGEN COMPOUNDS

Compound		g Nitrogen in 1 g Compound	Compound		g Nitrogen in 1 g Compound
(A)		(B)	(A)		(B)
1	Alanine	0.1572	28	Leucine	0.1068
2	Amino-butanoic acid	0.1358	29	Lysine	0.1916
3	Ammonia	0.8275	30	Methionine	0.0939
4	Arginine	0.3216	31	Nicotinamide	0.2294
5	Asparagine	0.2120	32	Nicotinic acid	0.1138
6	Aspartic acid	0.1052	33	Norleucine	0.1068
7	Bilirubin	0.0978	34	Norvaline	0.1196
8	Biliverdin	0.0927	35	Ornithine	0.2120
9	Biotin	0.1147	36	Pantothenic acid	0.0639
10	Choline	0.1156	37	Para-aminobenzoic acid	0.1022
11	Citrulline	0.2499	38	Phenylalanine	0.0848
12	Creatine	0.3205	39	Proline	0.1217
13	Creatinine	0.3715	40	Proteins	0.1600
14	Cysteine	0.1156	41	Pyridoxin	0.0828
15	Cystine	0.1166	42	Pteroylglutamic acid	0.2278
16	Epinephrin	0.7646	43	Riboflavin	0.1489
17	Ergothionine	0.1849	44	Serine	0.1333
18	Glucosamine	0.0782	45	Thiamin	0.1661
19	Glutamic acid	0.0952	46	Threonine	0.1176
20	Glutamine	0.1917	47	Thyroxin	0.0180
21	Glutathione	0.1367	48	Tryptophan	0.1372
22	Glycine	0.1866	49	Tyrosine	0.1725
23	Histamine	0.3781	50	Urea	0.4665
24	Histidine	0.2708	51	Uric acid	0.3333
25	Hydroxyproline	0.1068	52	Valine	0.1196
26	Indican	0.0401			
27	Isoleucine	0.1068			

# 78. BLOOD PHOSPHORUS, INORGANIC

Animal		Blood (B); R B C (C); Plasma (P); Serum (S)			Phosphorus mg/100 ml		
		Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml		
		Value (B)	Range (C)		Value (B)	Range (C)	
1	Man	B 2.92	2.1-3.82	21	Rat		
2		C 2.42	0.91-3.32	22			
3		P 3.22	2.4-4.42	23			
4	Camel	B 5.63		24	Sheep	B 5.63	3.3-5.83
5	Cat	B 5.64	4.6-6.94	25	Canary	P, S 7.7	6.0-9.03
6	Dog	B 3.1	2.1-4.0	26	Chicken	B 6.4	5.7-7.14
7		P, S 4.5	3.2-6.05	27	Duck	B 3.5	
8	Elephant	B 3.43		28	Goose	B 5.4	
9	Goat	B 5.74	4.0-6.14	29	Pigeon	B 5.2	
10	Guinea Pig	B 4.4	2.6-5.74	30	Turkey	B 6.4	
11	Horse	B 2.34	2.0-2.54	31	Alligator	B 5.6	
12	Kangaroo	B 4.3		32	Snake	B 2.9	
13	Monkey	B 3.64		33	Snake, moccasin	B 7.0	
14	Mouse	B 7.43		34	Turtle, snapping	B 3.3	
15	Opossum	B 3.3		35	Frog, bull	B 5.1	
16	Ox	B 5.84	4.9-7.24	36	Frog, green	B 8.1	
17	Pig	B 8.04	7.2-8.54	37	Necturus	B 5.3	
18	Rabbit	B 3.34	2.7-3.84	38	Bass, black	B 17.1	
19		P, S 5.06	3.4-6.96	39	Catfish	B 7.8	
20	Raccoon	B 4.33		40	Eel, congo	B 5.63	

/1/ Sample range "c" except as noted. /2/ Oxalated blood; determined by Fiske-Subbarow method. /3/ Heparinized blood; determined by Fiske-Subbarow method. /4/ Oxalated blood; determined by Kuttner and Lichtenstein method. /5/ Determined by Gomori method. /6/ Oxalated blood;

# 79. BLOOD PHOSPHORUS, ORGANIC ACID-SOLUBLE<sup>1</sup>

Blood (B); R B C (C)

Animal		Phosphorus mg/100 ml		Animal		Phosphorus mg/100 ml	
		Value	Range			Value	Range
(A)		(B)	(C)	(A)		(B)	(C)
1	Man	B 23.1 <sup>2</sup>	18.6-29 <sup>2</sup>	31	Raccoon	B (22.2) <sup>4</sup>	
2		C 50 <sup>2</sup>	39 -59 <sup>2</sup>	32		C 62 <sup>4</sup>	
3	Camel	B 15.3 <sup>3</sup>		33	Rat	B (23.0) <sup>3</sup>	
4		C 63 <sup>3</sup>		34		C 50 <sup>3</sup>	
5	Cat	B (8.3) <sup>3</sup>	(7.7-9.1) <sup>3</sup>	35	Sheep	B (4.5) <sup>3</sup>	
6		C 20.7 <sup>3</sup>	19.2-22.7 <sup>3</sup>	36		C 14.0 <sup>3</sup>	
7	Dog	B (26) <sup>3</sup>		37	Canary	B (61) <sup>4</sup>	
8		C 57 <sup>3</sup>		38		C 131 <sup>4</sup>	
9	Elephant	B (15.4) <sup>4</sup>		39	Chicken	B (30) <sup>4</sup>	
10		C 48 <sup>4</sup>		40		C 91 <sup>4</sup>	
11	Goat	B 3.5 <sup>3</sup>		41	Duck	B 38 <sup>3</sup>	
12		C 12.1 <sup>3</sup>		42		C 111 <sup>3</sup>	
13	Guinea pig	B (27) <sup>3</sup>		43	Goose	B (44) <sup>3</sup>	
14		C 65 <sup>3</sup>		44		C 101 <sup>3</sup>	
15	Horse	B (16.3) <sup>3</sup>		45	Pigeon	B (52) <sup>3</sup>	
16		C 47 <sup>3</sup>		46		C 120 <sup>3</sup>	
17	Kangaroo	B (27) <sup>4</sup>		47	Turkey	B (45) <sup>3</sup>	
18		C 53 <sup>4</sup>		48		C 94 <sup>3</sup>	
19	Monkey	B (21.5) <sup>4</sup>		49	Alligator	B (2.4) <sup>4</sup>	
20		C 51 <sup>4</sup>		50		C 13.9 <sup>4</sup>	
21	Mouse	B (34) <sup>4</sup>		51	Snake, moccasin	B (33) <sup>4</sup>	
22		C 85 <sup>4</sup>		52		C 165 <sup>4</sup>	
23	Opossum	B (21.8) <sup>4</sup>		53	Turtle, snapping	B (14.1) <sup>4</sup>	
24		C 51 <sup>4</sup>		54		C 59 <sup>4</sup>	
25	Ox	B (3.6) <sup>3</sup>		55	Frog, bull	B (13.2) <sup>4</sup>	
26		C 9.1 <sup>3</sup>		56		C 44 <sup>4</sup>	
27	Pig	B (40) <sup>3</sup>		57	Necturus	B (18.2) <sup>4</sup>	
28		C 97 <sup>3</sup>		58		C 97 <sup>4</sup>	
29	Rabbit	B 37 <sup>3</sup>		59	Catfish	B (21.4) <sup>4</sup>	
30		C 88 <sup>3</sup>		60		C 57 <sup>4</sup>	

/1/ All determinations on whole blood; cell content calculated on basis of blood content of the constituent and the standard hematocrit values. Calculated values are in parentheses ( ). In each of the following groups of figures the first number is the item number in the table, followed by the standard hematocrit value (underscored) and/or the hematocrit value reported by the investigator: (1), 44.5, (3), 24.3; (5), 40, 28.4; (7), 45.5, 34.1; (9), 32.2; (11), 34.5, 28.8; (13) 42.0, 36.4; (15), 35, 41; (17), 49.9; (19), 42, 43; (21), 41.5, 39.8; (23), 42.9; (25), 40, 38.8; (27), 41.5, 41.6, (29), 41.5, 41.6, (31), 36.1, (33), 46.0, 48.8; (35), 32, 31; (37), 46.5; (39), 32, 33.5, (41), 39.5, 44.5; (43), 44, 41; (45), 42.5, 45.1; (47), 38, 40, (49), 17.1, (51), 20.2, (53), 24.1; (55), 30, 30.8; (57), 18.8, (59), 34.6. /2/ Oxalated blood, determined by Lohmann method. /3/ Defibrinated blood, determined by Fiske-Subbarow method. /4/ Heparinized blood; determined by Fiske-Subbarow method.

# 80. BLOOD ADENOSINE TRIPHOSPHATE PHOSPHORUS<sup>1</sup>

Animal		Phosphorus mg/100 ml		Animal		Phosphorus mg/100 ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(C)	(A)	(B)	(C)	(C)
1 Man	B	8.1 <sup>2</sup>		31 Raccoon	B	1.4 <sup>3</sup>	
2 Camel	C	18.2 <sup>2</sup>	5.1-10.4 <sup>c2</sup>	32 Rat	C	4.0 <sup>3</sup>	
3 Cat	B	3.3 <sup>2</sup>	14.3-24.6 <sup>c2</sup>	33 Sheep	B	(6.6) <sup>2</sup>	
4 Dog	C	13.5 <sup>2</sup>		34 Canary	C	14.4 <sup>2</sup>	
5 Elephant	B	3.8 <sup>4</sup>	9-10 <sup>2</sup>	35 Chicken	B	(2.9) <sup>2</sup>	
6 Guinea pig	C	9.6 <sup>2</sup>		36 Duck	C	24.7 <sup>3</sup>	
7 Goat	B	(4.8) <sup>2</sup>		37 Goose	B	(4.7) <sup>2</sup>	11.1-17.7 <sup>c2</sup>
8 Horse	C	10.7 <sup>2</sup>		38 Pigeon	C	12.0 <sup>2</sup>	
9 Kangaroo	B	4.0 <sup>3</sup>		39 Turkey	B	35 <sup>2</sup>	27-34 <sup>c3</sup>
10 Monkey	C	12.5 <sup>3</sup>		40 Alligator	C	26 <sup>2</sup>	31-40 <sup>c3</sup>
11 Mouse	B	(5.8) <sup>3</sup>		41 Snake, moccasin	B	1.5 <sup>3</sup>	
12 Opossum	C	13.7 <sup>3</sup>		42 Turtle, snapping	C	9.0 <sup>3</sup>	
13 Ox	B	2.5 <sup>2</sup>		43 Frog, bull	B	25 <sup>3</sup>	
14 Pig	C	7.3 <sup>2</sup>		44 Frog, green	C	18.1 <sup>3</sup>	(5.2-6.3) <sup>c3</sup>
15 Rabbit	B	1.6 <sup>2</sup>		45 Necturus	B	5.9 <sup>3</sup>	17.6-20.9 <sup>c3</sup>
16 Kangaroo	C	4.5 <sup>2</sup>			C	21.8	
17 Monkey	B	4.6 <sup>3</sup>			C	12.3 <sup>3</sup>	
18 Mouse	C	9.2 <sup>3</sup>				66 <sup>3</sup>	
19 Opossum	B	(5.0) <sup>3</sup>					
20 Ox	C	12.0 <sup>3</sup>					
21 Pig	B	(5.9) <sup>3</sup>					
22 Rabbit	C	14.1 <sup>3</sup>					
23 Kangaroo	B	(5.2) <sup>3</sup>					
24 Monkey	C	12.1 <sup>3</sup>					
25 Mouse	B	(2.1) <sup>2</sup>					
26 Opossum	C	5.1 <sup>2</sup>					
27 Ox	B	(14.2) <sup>2</sup>					
28 Pig	C	34 <sup>2</sup>					
29 Rabbit	B	(10.0) <sup>2</sup>					
30 Kangaroo	C	24 <sup>2</sup>					

/1/ All determinations on whole blood, cell content calculated on basis of blood analysis and hematocrit values. Calculated values in parentheses ( ). The following figures represent first, the item, next, the hematocrit, standard (underscores) and/or value reported by investigator (not underscored). (1), 44.5, 45.2, (3), 24.3, (5), 40, 28.1, (7), 45.5, 34.2, (9), 32.2, (11), 42, 36.4, (13), 34.5, 27.7; (15), 35, 41, (17), 49.9; (19), 42, 42.6, (21), 41.5, 40, (23), 42.9, (25), 40, 39, (27), 41.5, 37, (29), 41.5, (31), 36.1; (33), 46, 48.8, (35), 32, 31.1; (37), 46.5, (39), 32, 32.2, (41), 34.5, (43), 44, 47.9; (45), 45.1, (47), 38, 39.7, (49), 13.8, (51), 20.2, (53), 24.1, (55), 30, 30.8; (57), 27.0, (59), 18.8. "Hydrolyzable" phosphorus determined by Lohmann method, liberated P determined by Fiske-Subbarow method; these values were multiplied by 3/2 (Kerr & Daoud; Guest & Rapoport) to derive A. T. P. P values. /2/ De-fibrinated blood. /3/ Heparinized blood.



# 81. BLOOD DIPHOSSPHOGLYCERATE PHOSPHORUS<sup>1</sup>

Blood (B), R B C (C)				
Animal	Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml
	Value	Range		
(A)	(B)	(C)	(A)	(C)
1 Man	B 12.42	B 1-16.7 <sup>2</sup> C 2.92	19 Monkey	B (13.0) <sup>3</sup> C 31.3
2 Camel	B 10.13	19-40 <sup>2</sup>	20 Mouse	B (21.5) <sup>3</sup> C 53.3
3 Cat	B 11.61		21 Opossum	B 9.83 C 22.83
4 Dog	B 14.11 <sup>3</sup>	3.5-4.5 <sup>4</sup>	22 Ox	B <0.34 C <0.64
5 Elephant	B 6.03		23 Pig	B (18.3) <sup>4</sup> C 44.4
6 Goat	B 10.73		24 Rabbit	B (18.0) <sup>5</sup> C 45.3
7 Guinea pig	B (13.1) <sup>1</sup>		25 Raccoon	B 9.93 C 27.3
8 Horse	B (11.6) <sup>1</sup>		26 Rat	B (15.6) <sup>5</sup> C 14.3
9 Kangaroo	B 9.33	31-53 <sup>3</sup>	27 Sheep	B <0.34 C <0.84
10	B 18.63		28	

<sup>1/1</sup> All determinations on whole blood, R B C values calculated on basis of whole blood analysis and hematocrit values. Calculated values in parentheses ( ). The following figures represent, first, the item, next, the hematocrit value, standard (underscored), first, the item, next, the hematocrit value reported by the investigator (not underscored). (1) 44.5 (11), 34.8 (9), 24.7 (7), 13.2 (4), 8.5 (3), 32.2 (11), 34.5 (27.7), (13), 32.3 (13), 35.10 (5), 17.1 (4), 42.6 (21), 41.5 (39.8), (23), 42.9, (43), 49.45 (27), 41.5 (42.6), 41.5 (42.6), 37.6 (31), 36.1 (33), 46.0, 40.6, (35), 32.3 (21) (Oxidized blood) Diposphoglycerate P = (Total acid soluble P)-P value after hydrolysis with N HCl at 100° C for 180 minutes. <sup>2/2</sup> Heparitized blood. Rapoport method. <sup>3/3</sup> Defibrinated blood, determined by Rapoport method.

# 82. BLOOD NUCLEOTIDE PHOSPHORUS<sup>1</sup>

Blood (B), R B C (C)				
Animal	Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml
	Value	Range		
(A)	(B)	(C)	(A)	(C)
1 Man	B 2.8	2.2-3.4 <sup>2</sup>	19 Pig	B (4.5) C 10.8
2 Camel	B 1.4	1.1-1.7 <sup>2</sup>	20 Rabbit	B (3.2) C 7.7
3 Cat	B 1.1		21 Rat	B (1.9) C 4.1
4 Dog	B 1.1		22 Sheep	B 1.2 C 3.8
5 Goat	B 1.1		23 Chicken	B (1.8) C 5.5
6 Guinea pig	B 2.0		24 Duck	B 4.8 C 14.0
7 Horse	B (0.5)		25 Goose	B (6.2) C 14.0
8 Monkey	B 1.2		26 Turkey	B (3.7) C 9.9
9 Ox	B 0.6		27 Turtle, sea	B 2.4 C 8.8
10	B 3.2		28	
11	B 5.0		29	
12	B 5.5		30	
13	B 1.5		31	
14	B 1.5		32	
15	B 1.2		33	
16	B 3.7		34	
17	B 0.6		35	
18	B 2.0		36	

<sup>1/1</sup> All determinations on defibrinated whole blood by the Kerr and Bligh method, values for RBC content calculated on the basis of the whole blood content and the hematocrit values. Calculated values are in parentheses ( ). The following figures represent, first, the item, next, the hematocrit, standard, (underscored), and/or the value reported by the investigator (not underscored). (1) 44.5 (11), 34.5 (23), 32.3 (13), 35.10 (5), 17.1 (4), 42.6 (21), 41.5 (39.8), (23), 42.9, (43), 49.45 (27), 41.5 (42.6), 41.5 (42.6), 37.6 (31), 36.1 (33), 46.0, 40.6, (35), 32.3 (21) (Oxidized blood) Diposphoglycerate P = (Total acid soluble P)-P value after hydrolysis with N HCl at 100° C for 180 minutes. <sup>2/2</sup> Heparitized blood. Rapoport method. <sup>3/3</sup> Defibrinated blood, determined by Rapoport method.

### 83. BLOOD LIPID PHOSPHORUS

Blood (B); R B C (C); Plasma (P); Serum (S)

Animal	Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml	
	Value	Range		Value	Range
(A)	(B)	(C)	(A)	(B)	(C)
1 Man	B 11.21		4 Dog	S 14.71	
2					
3					
4					

/1/ Determined by Fiske-Subbarow method after treatment with Bloor's mixture.

/2/ Determined by Bell and Doisy method after Bloor's mixture. /3/ 4.8

(2.8-6.7) by Tisdall method after extraction in alcohol, then chloroform.

### 84. BLOOD HEXOSE PHOSPHATE PHOSPHORUS<sup>1</sup>

Blood (B); R B C (C); Plasma (P)

Animal	Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml	
	Value	Range		Value	Range
(A)	(B)	(C)	(A)	(B)	(C)
1 Man	B 3.2	1.4-5.0	3 Man	P 0.04	0.0-0.22
2	C 7.5	3.5-10.7			

/1/ Hexose phosphate P = (P value after hydrolysis in N HCl for 180 minutes at 100°C) - (P value after hydrolysis in N HCl for 7 minutes at 100°C).

### 85. BLOOD PHYTIC ACID PHOSPHORUS<sup>1</sup>

Blood (B); R B C (C)

Animal	Phosphorus mg/100 ml		Animal	Phosphorus mg/100 ml	
	Value	Range		Value	Range
(A)	(B)	(C)	(A)	(B)	(C)
1 Canary	B 26		8 Goose	C 71	
2	C 56		9 Pigeon	B 33	
3 Chicken	B 19.9		10	C 74	71-86
4	C 65	61-68	11 Turkey	B (31)	
5 Duck	B 31		12	C 74	
6	C 69		13 Turtle, snapping	B 6.1	
7 Goose	B (29)		14	C 25	

/1/ Found only in nucleated R B C. All determinations on heparinized whole blood by the Michel-Durand method as modified by Rapoport and Leva, values for R B C calculated on the basis of blood contents and hematocrit values. Parentheses ( ) enclose values calculated on basis of standard hematocrit values.

## 86. BLOOD SULFUR

Blood (B), R B C (C); Plasma (P), Serum (S)

Constituent	Animal	Sulfur mg/100 ml		Constituent	Animal	Sulfur mg/100 ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
1 Sulfate S, total	Man	P 1.1	0.9-1.3	11 Inorganic S	Rabbit	P 5.0	3.6-6.1
	Rabbit	P 5.4	4.0-6.9	12	Rat	P 1.0	
3 Non-protein S	Man	P 2.8	2.4-3.6	13	Man	C 0.015	
	Rabbit	P 6.9	6.0-8.4	14	Man	P 0.1	0-0.2
5	Man	P 0.9	0.8-1.1	15	Cow	C 0.21	
6 Inorganic S	Cow	C 1.45		16	Dog	C 0.54	
	Dog	C 1.35		17	Goat	C 0.15	
	Dog	P 3.2		18	Rabbit	P 0.4	0.1-1
	Goat	C 1.5		19	Man	P 1.7	1.4-2.6
10	Horse	P 3.4		20	Rabbit	P 1.6	1.0-2.1

/1/ Ethereal S.

## 87. BLOOD SULFUR COMPOUNDS

Blood (B), R B C (C), Plasma (P)

Constituent	Animal	Sulfur Compounds per 100 ml		Constituent	Animal	Sulfur Compounds per 100 ml	
		Value mg	Range mg			Value mg	Range mg
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
1 Sulfate, total	Man	P 3.3	2.7-3.9	23	Man	B (15)	25-41
	Rabbit	P 16.2	12-21	24	Man	C 79	
3	Man	P 2.7	2.4-3.3	25	Man	P 0.0	
4 Sulfate, inorganic	Dog	P 9.6		26	Cow	B 40	
	Horse	P 10.2		27	Cow	C (157)	
	Rabbit	P 14.4	10.8-27	28	Cow	P 0.0	
7	Rat	B 3.0		29	Rat	B 40	30-45
8 Sulfate, conjugated <sup>1</sup>	Man	P 0.3	0-0.6	30	Rat	C (182)	(116-205)
10 Cystine <sup>2</sup>	Rabbit	P 1.2	0.3-3.0	31	Rat	P 0.0	
11	Man	B	1.9-5.5	32	Man	B (4)	
12	Man	C 9.6	3.9-17.7	33	Man	C 8.5	
13 Ergothioneine <sup>3</sup>	Man	P 0.0		34	Man	P 0.0	
	Pig	B (8.4)	(6.1-9.8)	35	Cow	B 6	
	Pig	C 20.7	15.6-24	36	Cow	C (24)	
16	Pig	P 0.0		37	Cow	P 0.0	
17	Man	B (39) <sup>6</sup>		38 Indican	Man	P	0.0-0.6
18	Man	C 87		39 Insulin <sup>4</sup>			
19 Glutathione total	Man	P 0.0		40 Methionine <sup>5</sup>			
	Cow	B 46		41 Thiamine <sup>5</sup>			
	Cow	C (181)		42 Thiocyanate	Man	B 0.77	0.5-1.4
22	Cow	P 0.0					

1/ Or ethereal sulfate 2/ See table on Free Amino Acid Content 3/ Or thioneine, present in R B C only.  
4/ See table on Blood Hormone Content. 5/ See table on Blood Vitamin Content. 6/ Figures in parentheses were calculated on basis of standard hematocrit values, see table on Hematocrit Values.

88. BLOOD VITAMINS			
Constituent	Animal	100 ml Blood	
(A)			

		100 ml Blood		100 ml R B C		100 ml Plasma <sup>1</sup>	
(A)	(B)	Value	Range	Value	Range	Value	Range
		(C)	(D)	(E)	(F)	(G)	(H)
1	Vitamin A as carotenol	Man	13 $\mu$ g	9-17 $\mu$ g	0 <sup>2</sup>	24 $\mu$ g	10-60 $\mu$ g
2		Cow	14 $\mu$ g	6-18 $\mu$ g	0	24 $\mu$ g	10-30 $\mu$ g
3		Dog	1.5 $\mu$ g	0-3 $\mu$ g	0	3 $\mu$ g	0-5 $\mu$ g
4		Horse	8 $\mu$ g	6-10 $\mu$ g	0	12 $\mu$ g	9-16 $\mu$ g
5		Pig	12 $\mu$ g	5-20 $\mu$ g	0	20 $\mu$ g	10-35 $\mu$ g
6		Rabbit	25 $\mu$ g	15-70 $\mu$ g	0	45 $\mu$ g	30-130 $\mu$ g
7		Rat	25 $\mu$ g	4-70 $\mu$ g	0	10 $\mu$ g	6-12 $\mu$ g
8		Sheep	6 $\mu$ g	14-31 $\mu$ g	0	35 $\mu$ g	20-45 $\mu$ g
9		Chicken	31 $\mu$ g	10-68 $\mu$ g	0	45 $\mu$ g	15-100 $\mu$ g
10	Vitamin A as carotene	Man	120 $\mu$ g	20-300 $\mu$ g	0	220 $\mu$ g	40-540 $\mu$ g
11		Cow	40 $\mu$ g	25-950 $\mu$ g	0	70 $\mu$ g	50-2000 $\mu$ g
12		Horse	65 $\mu$ g	13-114 $\mu$ g	0	100 $\mu$ g	20-175 $\mu$ g
13		Pig	0		0	10 $\mu$ g	0-20 $\mu$ g
14		Sheep	0		0	50 $\mu$ g	0.1-2.3 $\mu$ g
15	Ascorbic <sup>3</sup> acid	Chicken	7 $\mu$ g		0	0.7 $\mu$ g	0.2-1.5 $\mu$ g
16		Man	24 $\mu$ g	0-14 $\mu$ g	1.0 mg	0.5 mg	0.2-1.2 $\mu$ g
17		Cow	0.62 mg	0.2-0.7 mg	0.5-2.8 mg	0.5 mg	0.2-2.1 mg
18		Horse	0.5 mg	0.2-1.5 mg		0.5 mg	0.05-1.5 mg
19		Pig	0.5 mg	0.2-1.5 mg		0.4 mg	0.4-0.8 mg
20		Dog	0.4 mg	0.2-1.2 mg		0.5 mg	
21		Rat	0.5 mg	0.2-2.1 mg		0.5 mg	
22		Sheep	0.5 mg	0.1-1.5 mg		0.5 mg	
23		Man	0.5 mg	0.4-0.8 mg			
24		Cow <sup>5</sup>	0.08 $\mu$ g	0.06-0.14 $\mu$ g			
25	Dog	0.05 $\mu$ g	0.04-0.05 $\mu$ g				
26	Goat	0.09 $\mu$ g	0.05-0.11 $\mu$ g				
27	Horse	0.07 $\mu$ g	0.06-0.09 $\mu$ g				
28	Mouse <sup>6</sup>	0.21 $\mu$ g	0.12-0.34 $\mu$ g				
29	Mouse <sup>7</sup>	0.23 $\mu$ g	0.22-0.23 $\mu$ g				
30	Pig	0.12 $\mu$ g	0.09-0.14 $\mu$ g				
31	Rabbit	0.10 $\mu$ g	0.08-0.13 $\mu$ g				
32	Rat,	1.01 $\mu$ g	0.64-1.50 $\mu$ g				
33	cotton	0.36 $\mu$ g	0.33-0.38 $\mu$ g				
34	Rat, white						
35	Sheep	0.08 $\mu$ g	0.05-0.12 $\mu$ g				
36	Chicken	0.07 $\mu$ g	0.06-0.09 $\mu$ g				
37	Turkey	0.56 $\mu$ g	0.38-0.66 $\mu$ g				
38	Turtle	0.53 $\mu$ g	0.45-0.60 $\mu$ g				
39	Biotin <sup>3</sup>	Man	0.66 $\mu$ g	0.59-0.69 $\mu$ g			
40		Mouse	1.23 $\mu$ g	0.8-1.7 $\mu$ g			
41		Rat	1.25 $\mu$ g				
42	Vitamin C, see Ascorbic acid			1.5-3.5 $\mu$ g			
43	Carotene, -ol, see Vitamin A						
44	Choline <sup>3</sup> , total	Man			1.3 $\mu$ g	1.0-1.7 $\mu$ g	
45		Cow					
46		Dog					
47		Horse					
48	Mouse	18.5 mg	11-31 mg				
49	Sheep						
50					16.5 mg	26-35 mg	
51					15 mg	12-15 mg	
52					10 mg	8-15 mg	

1/ Or serum when specified  
chemical methods. 2/ Absence from R B C  
manila as test organism 3/ Oxalated blood, determined by the method of Skeggs et al, using  
albino. 4/ In pasture fed cow, in dry lot fed calf, 5/ Swiss albino 6/ 3/ Determined by the method of Skeggs et al, using

1/ Or serum when specified chemical methods.	2/ Absence from R B C not conclusively established	3/ Determined by Lactobacillus leich-
manii as test organism	4/ Oxalated blood, determined by the method of Skeggs et al, using 0.09 (0.08-0.10) µg.	5/ American
albino.	6/ Swiss albino	
Sheep	18.5 mg	16.5 mg
		15 mg
		10 mg
		26-35 mg
		12-15 mg
		8-15 mg

# 88. BLOOD VITAMINS (Continued)

	(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
47	Choline <sup>1</sup> free	Man	2.5 mg	1.0-4.0 mg		4.4-7.5 mg		0.05-2.5 mg
48		Cow					4 mg	
49		Dog						1.0-1.1 mg
50		Guinea pig						2-12 mg <sup>2</sup>
51		Horse					4 mg	
52		Pig					2.6 mg	
53		Rabbit					0.5 mg	
54	Vitamin D <sup>3</sup> as Calcif- erol (D <sub>2</sub> )	Rat						0.05-0.3 mg
55		Sheep					3 mg	1-4 mg
56		Man					2.8 µg <sup>2</sup>	1.7-4.1 µg <sup>2</sup>
57		Cow			1.4 µg		6.8 µg <sup>2</sup>	
58		Dog					1.4 µg <sup>2</sup>	
59		Pig					3.0 µg <sup>2</sup>	2.6-3.4 µg <sup>2</sup>
60		Rabbit					1.3 µg <sup>2</sup>	1.1-1.8 µg <sup>2</sup>
61	Vitamin E <sup>1</sup> (Tocopherol)	Sheep					1.3 µg <sup>2</sup>	1.1-1.8 µg <sup>2</sup>
62		Chicken					2.5 µg <sup>2</sup>	
63		Man					1.2 mg	0.9-1.9 mg <sup>2</sup>
64		Cow					0.40 mg <sup>2</sup>	0.20-0.50 mg <sup>2</sup>
65		Dog					0.6 mg	
66		Pig					0.1 mg	0.09-0.15 mg
67		Rat						0.05-0.06 mg
68		Sheep					0.02 mg	0.01-0.03 mg
	Folic acid, see Pteroylglutamic acid							
69	Inositol <sup>5</sup>	Man					0.50 mg	0.37-0.76 mg
70	Vitamin K	Mouse	6.6 mg					
71	Nicotinic acid <sup>6</sup>	Man	0.6 mg <sup>7,8</sup>	0.2-0.9 mg <sup>1,9</sup>	1.3 mg <sup>10</sup>		0.075 mg <sup>1</sup>	0.025-0.15 mg <sup>1</sup>
72		Cow	0.3 mg <sup>7</sup>					
73		Dog	0.8 mg	0.5-1.3 mg <sup>11</sup>	1.6 mg			
74		Guinea pig		6.5-8.9 mg <sup>12</sup>				
75		Horse	0.6 mg <sup>7,13</sup>					
76		Ox	0.9 mg <sup>7</sup>					
77		Pig	0.5 mg <sup>7</sup>					
78	Pantothenic acid <sup>6</sup>	Rat	1.4 mg <sup>7</sup>	1.2-1.8 mg				
79		Sheep	1.0 mg <sup>7</sup>	0.4-1.4 mg				
80		Chicken	1.1 mg <sup>7</sup>	1.0-1.2 mg <sup>4</sup>				
81		Man	30 µg	15-45 µg	25 µg	15-30 µg	15 µg	6-35 µg
82		Dog	25 µg	15-35 µg	25 µg	20-30 µg	30 µg	15-40 µg
83		Horse	45 µg	35-55 µg	52 µg	45-60 µg	38 µg	30-45 µg
84		Pig	35 µg	30-40 µg	30 µg	25-35 µg	35 µg	30-40 µg
85		Rabbit	20 µg	15-35 µg				20-30 µg
86		Sheep	35 µg	20-50 µg	30 µg	20-40 µg	25 µg	20-30 µg
87		Chicken	45 µg	40-50 µg				

/1/ Determined by chemical method. /2/ Content in serum. /3/ Determined by animal assay method. /4/ 6.5-10.5 mg coenzyme. /5/ Determined by microbiological method with Neurospora. /6/ Determined by microbiological method. /7/ Total nicotinic acid activity. /8/ 3.0 as coenzyme. /9/ 2.0-3.5 mg as coenzyme. /10/ 0.135 mg free nicotinic acid. 7.0 (6.5-9.0) mg as coenzyme. /11/ 5.1-6.6 mg as coenzyme. /12/ As coenzyme. /13/ 0.27 mg free nicotinic acid.

# 88. BLOOD VITAMINS (Concluded)

	(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
88	Para-amino benzoic Acid <sup>1</sup>	Man	3.4 µg					
89		Mouse	29 µg					
90		Man	3.5 µg	2.3-5.3 µg			1.7 µg	1.5-5.0 µg
91	Pteroylglutamic acid <sup>1</sup> , total <sup>2</sup>	Cattle		2.1-3.0 µg				1.8-2.2 µg
92		Pig		2.3-3.1 µg				1.8-2.1 µg
93		Sheep	4.6 µg	3.5-5.5 µg				
94		Chicken		2.2-4.3 µg				3.0-4.2 µg
95		Man	0.085 µg	0.05-0.13 µg			0.05 µg	
96		Cattle	0.19 µg	0.06-0.45 µg			0.05 µg	
97	Pteroylglutamic acid <sup>1</sup> , free	Horse	0.33 µg	0.22-0.6 µg			0.35 µg	0.25-0.48 µg
98		Pig	0.66 µg	0.45-0.99 µg			0.61 µg	0.48-0.87 µg
99		Chicken	0.87 µg	0.44-1.65 µg			0.31 µg	0.14-0.42 µg
100		Turkey	1.68 µg	1.02-2.76 µg			0.6 µg	0.25-1.08 µg
101	Pyridoxine	Monkey	11 µg	5-20 µg	5 µg	2-21 µg	8 µg	1-18 µg
102		Mouse	42 µg					
103		Sheep	12 µg					
104		Man		15-60 µg <sup>1</sup>	22.4 µg <sup>3</sup>	18-26 µg <sup>3</sup>	3.2 µg <sup>3</sup>	2.6-3.7 µg <sup>3</sup>
105		Dog	97 µg <sup>1</sup>	90-100 µg <sup>1</sup>				
106		Cow, young	45 µg <sup>1</sup>	40-50 µg <sup>1</sup>				
107	Riboflavin	Pig	95 µg <sup>1</sup>					
108		Rat	45 µg <sup>1</sup>	20-65 µg <sup>1,5</sup>		100-130 µg <sup>1,4</sup>		
109		Sheep	27 µg <sup>1</sup>	24-30 µg <sup>1</sup>				
110		Chicken	15 µg <sup>5</sup>	12-18 µg <sup>6</sup>			34 µg <sup>6</sup>	33-35 µg <sup>6</sup>
111		Snake		0-310 µg <sup>1</sup>				
112		Man ♂	8.9 µg <sup>12</sup>	6.1-10.7 µg <sup>1,6</sup>	10 µg <sup>1</sup>	7-14 µg <sup>12</sup>		
113		Man ♀	7.6 µg <sup>12</sup>	6.2-9.0 µg <sup>8,12</sup>	6.5 µg <sup>1</sup>	5-8 µg <sup>12</sup>		
114		Man	8.0 µg <sup>8</sup>	4-11 µg <sup>8</sup>	8.0 µg <sup>8</sup>	7-10 µg <sup>8</sup>	7 µg <sup>8</sup>	1-9 µg <sup>8</sup>
115		Cow	8 µg <sup>8,9</sup>	5-12 µg <sup>8,10</sup>				
116		Dog	7.0 µg	5-9 µg <sup>8</sup>			2.10 µg <sup>8</sup>	0.1-4.0 µg <sup>8</sup>
117		Guinea pig		45-80 µg <sup>12</sup>				
118		Horse		1.1-1.7 µg <sup>12</sup>				
119	Thiamine <sup>1</sup>	Ox		4.5-5.1 µg <sup>12</sup>				
120		Pig	20 µg <sup>8</sup>	17-30 µg <sup>8,11</sup>				
121		Rabbit		3-30 µg <sup>8</sup>				
122		Rat	20.5 µg <sup>12</sup>	10-25 µg <sup>12</sup>				
123		Sheep	5.8 µg <sup>8</sup>	3-15 µg <sup>7,8</sup>			3.8 µg <sup>8</sup>	
124		Chicken	6.0 µg <sup>12</sup>	5.6-11 µg <sup>12</sup>				
125		Pigeon	29 µg <sup>8</sup>	21-40 µg <sup>8</sup>				14-22 µg <sup>8</sup>
126		Frog	9.0 µg <sup>12</sup>					
127		Snake			(80 µg) <sup>13</sup>			
Totopherol, see Vitamin E								

/1/ Determined by microbiological method. /2/ Takadiastase treated samples /3/ Determined by Burch-Beesey-Lowry fluorometric method /4/ As coenzyme. /5/ 50-65 µg coenzyme /6/ Determined by fluorometric method. /7/ 4-4.7 µg thiamin pyrophosphate /8/ Total thiamin /9/ 0.5 µg free thiamin. /10/ 0-0.1 µg free thiamin. /11/ 12-20 µg thiamin pyrophosphate /12/ As thiamin pyrophosphate. /13/ In 1 g R B C.

Constituent	Animal	Value			Range
		(A)	(B)	(C)	
1 Adrenocortico-trophic H 1	Man	B	B	B	
2 Androgenic H 2	Rat	B	B	B	
3 as Testosterone	Man	B	B	B	
4 Chloranolic Condo-trophic H 3	Woman	P	2.8 mg	P	2-3-4 mg
5	Woman, pregnant	P	2.8 mg	P	2-3-4 mg
6	Woman, pregnant	P	14000 lu	P	2-3-4 mg
7 Corticosteroids 14	Woman	P	0.23 mg	P	400-60000 lu
8	Dog, male	P	0.11-0.42 mg	P	400-1000 lu
9	Dog, female	P	0.27 mg	P	0.11-0.42 mg
10	Man	P	0.22 mg	P	0.12-0.38 mg
11	Cat	P	0.31 mg	P	0.12-0.38 mg
12 Epinephrine 8	Rabbit	B	B	B	
13	Woman, pregnant	B	B	B	
14	Cow	B	8.4 µg	B	1-6-3.1 µg
15	Cow	B	0.32 mg	B	1-6-3.1 µg
16	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
17	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
18	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
19	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
20	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
21	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
22	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
23	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
24	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
25	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
26	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
27	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
28	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
29	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
30	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
31	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
32	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
33	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
34	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
35	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
36	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
37	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
38	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
39	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
40	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
41	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
42	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
43	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
44	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
45	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
46	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
47	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
48	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
49	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
50	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
51	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
52	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
53	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
54	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
55	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
56	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
57	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
58	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
59	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
60	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
61	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
62	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
63	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
64	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
65	Cow, pregnant	P	0.32 mg	P	1-6-3.1 µg
66	Cow, pregnant	P	0.32 mg		

to determine the normal acrobatic acid depletion  
trophic growth of 100  $\mu$  pregnant urate deple-  
bit, east, pit except as pregnant urate deple-  
suprarenal in blood, trace cited, expected de-  
15, 6 months, 19/14 By acet demonstrable in deoxy  
menopause. 19/14 By acet demonstrable in deoxy  
6.4-20.0  $\mu$  per 100 ml. 12/27 "Oxytocin" one unit equivalent  
2/25 "vag per 100 ml. 12/27 "Oxytocin" one unit equivalent  
progestins." 15/6, 8. 2/27 "Oxytocin" one unit equivalent  
detected in plasma of Cysto horse, 30-70  $\mu$  per 100 ml. on on  
stasis method of D'Angelo horse, 30-70  $\mu$  per 100 ml. on on  
stasis method of D'Angelo horse, 30-70  $\mu$  per 100 ml. on on  
non-pregnant woman. 14/17 L. T. Samuels reports values

# 90. BLOOD ENZYME ACTIVITY

Variable	Blood (B), RBC (C); Plasma (P), Serum (S)				Variable	Animal	Enzyme Activity per 100 ml		Enzyme Activity per 100 ml
	(A)	Animal	Value (C)	Range (D)			(A)	Value (C)	
1 Adenosine deaminase		Rabbit	4151		20 Carbonic anhydrase (continued)	Cattle			
2 Adenosine polyphosphatase, acid		Man	412	21-61b	21	Calf			
3 Adenosine polyphosphatase, alkaline		Man	302	10-51b	22	Calf			
4 Adrenic acid deaminase, muscle		Rabbit	413		23	Guinea pig		44,000 <sup>7</sup>	89,000-140,000 <sup>7</sup>
5 Aldolase		Man	4904		24	Pig		60,000 <sup>7</sup>	
6 Amylase		Man	90,000 <sup>4</sup>	350-800 <sup>d</sup>	25	Rat		59,000 <sup>7</sup>	
7 Arginase		Rat, albino	6,000 <sup>4</sup>		26	Chicken		144,000 <sup>7</sup>	
8		Man	70,000-110,000 <sup>d</sup>		27	Man		22,000 <sup>7</sup>	
9		Man	4,000-9,000 <sup>d</sup>		28	Cow		699	
10		Man	80-150 <sup>5</sup>		29	Man		3311	420-950
11		Man	2,500-7,700 <sup>5</sup>		30	Man		54911	81,600-469,200
12		Monkey	1,800-7,700 <sup>5</sup>		31	Pig		17611	273-40711
13		Mouse	800-3,200 <sup>b</sup>		32	Cal			437-66011
14		Rat	0		33	Dog			139-21911
15		Rat	0		34	Guinea pig			135-36212
16		Man	0		35	Substrate			92-14012
17		Man	0		36	Cholinesterase (acetylcholine)			112-30312
18		Man	0		37	Ox			125-23012
19		Man	0		38	Pig			248-31012
		Man	0		39	Rabbit		4512	430-68512
		Man	0		40	Rat		4012	
		Man	0		41	Sheep		25313	18-3512
		Man	0		42	Chicken		012	
		Man	0		43				
		Man	0		44				

1/1/  $\mu$ g N liberated/hr. 2/2/  $\mu$ M P from ATP/hr at pH 4.8 for acid adenosine polyphosphatase, and at pH 8.2 for alkaline adenosine polyphosphatase. Corrected for inorganic P and non-enzymatic hydrolysis. 3/3/  $\mu$ g N liberated/hr in presence of 0.05% adenylic acid, starch and fructose diphosphate/hr at 38°C at pH 6.6. No change in pregnancy. 4/4/ mg dextrose or equivalent (by copper reduction) adults. 5/5/ g/100 g dry weight. 6/6/ Units as described by Kochian. 7/7/ Amount of RBC Zn concentration in all conditions in acetylcholine hydrolyzed/min, as measured by Dille et al. 8/8/ mg  $H_2O_2$  split/10 min under conditions specified. 9/9/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 10/10/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 11/11/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 12/12/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 13/13/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 14/14/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 15/15/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 16/16/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 17/17/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 18/18/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine. 19/19/  $\mu$ M acetylcholine hydrolyzed/min from 0.01M acetylcholine.



# 90. BLOOD ENZYME ACTIVITY (Continued)

Blood (B); RBC (C); Plasma (P); Serum (S)

Variable	Animal	Enzyme Activity per 100 ml		Variable	Animal	Enzyme Activity per 100 ml	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
45	Man	C 261	69		Man	C Trace	
46		P 5.8	70			P 110	
47	Cat	C 12.1	71		Cat	C 0	
48		P 16.2	72			P 45	
49	Cow	C 211	73		Cow	C Trace	
50		P Trace	74			P Trace	
51	Dog	C 61	75		Dog	C 0	
52		P 14.8	76			P 144	
53	Guinea pig	C 153	77		Guinea pig	C Trace	
54		P 16.6	78			P 60	
55	Horse	C 81	79		Horse	C Trace	
56		P 8.1	80			P 243	
57	Rabbit	C 34	81		Rabbit	C 0	
58		P 25.2	82			P 19.8	
59	Rat	C +	83		Rat	C 0	
60		P 16.2	84			P 5.8	
61	Sheep	C 76	85		Sheep	C 0	
62		P 13.0	86			P 0	
63	Fowl	C Trace	87		Fowl	C Trace	
64		P 72	88			P 4.5	
65	Labrus (bony fish)	C 0	89		Labrus (bony fish)	C Trace	
66		P 0.3	90			P Trace	
67	Scyllium (elasmobranch)	C 0	91		Scyllium (elasmobranch)	C 0	
68		P 7.6	92			P Trace	

/14/  $\mu$ M acetyl-beta-methylcholine hydrolyzed/min as measured by volume  $\text{CO}_2$  evolved/min from 0.03M acetyl-beta-methylcholine. /15/  $\mu$ M benzoylcholine hydrolyzed/min as measured by volume  $\text{CO}_2$  evolved/min from 0.006M benzoylcholine. /16/ Where figures in literature reported in terms of whole blood, hematocrit values of 45 assumed in present conversion. /17/ RBC cholinesterase activity high in reticulocytes and young cells; high in conditions accompanied by hyperactive hematopoiesis.

# 90. BLOOD ENZYME ACTIVITY (Concluded)

Blood (B), R B C (C), Plasma (P), Serum (S)

Variable	Animal	Enzyme Activity per 100 ml		Variable	Animal	Enzyme Activity per 100 ml	
		Value (C)	Range (D)			Value (C)	Range (D)
93 Dehydrogenase	Man	S	35918	106 Lipase	Man	S	0-15022
94 Glucose-6-phosphate dehydrogenase	Rat	C	+	107 Methemoglobin reductase	Rabbit	C	+
95 Beta-Glucuronidase	Man	S		108 Phenolsulphatase	Man	S	30-1,55023
96	Man	S		109 Phosphatase, acid	Man	S	1.0-4.024
97	Man	S		110	Man, adult	S	10.5-13.025
98 Glyoxalase	Man	C	1,398,000.20	111 Phosphatase, alkaline?	Man, adult	S	1.0-4.026
99	Rat	B	611,703.20	112	Man, child	S	5.0-14.028
100	Rat	C	2,040,000.20	113 Prothrombin	Man	P	50-12529
101 Hexokinase	Rat	B	861,000.10	114 Lysine	Man	P	80-10030
102 Histaminase	Man	S	1621				
103	Man	S	1823				
104 Lactic dehydrogenase	Man	S	10-40				
105	Man	C	0-36				

/18/  $\mu$ MNH<sub>3</sub>/10 min from DL-alanyldihydroalanine at 37°C at pH 8.1. /19/  $\mu$ g phenolphthalein/hr from phenolphthalein-glucuronide at 38°C at pH 4.5. Rise in pregnancy from 300  $\mu$ g at 3 mos. to 1350  $\mu$ g at term. /20/  $\mu$ l CO<sub>2</sub>/20 min from methylglyoxal at pH 7.2 at 26°C in presence of glutathione. /21/  $\mu$ g histamine destroyed in 90 min at 37°C. Rise in pregnancy. /22/ ml N/20 NaOH/24 hr from standard olive oil emulsion. /23/ Amount of enzyme which produces color equivalent to 10  $\mu$ g of p-nitrophenol from p-nitrophenyl sulphate. /24/ mg phenol/hr from disodiumphenylphosphate at 37°C at pH 5.0. /25/ mg phenol/30 min from disodiumphenylphosphate at 37.5°C at pH 9.0. /26/ mg P/hr from beta-glycerophosphate at 37°C at pH 5.0. /27/ Rise in oestrodiol activity and extract at 37°C in 120 seconds at 28°C at pH 7.2 in isotonic saline buffered with imidazole. /30/  $\mu$ g folic acid/90 min from yeast

# 91. BLOOD COENZYMES

Blood (B); RBC (C); Plasma (P)

COENZYMES						
Variable		Blood (B); RBC (C); Plasma (P)				
(A)	Animal	$\mu\text{g}/100 \text{ ml}$		Method		
		Value	Range			
(B)	(C)	(D)	(E)			
1 Coenzyme A (as bound pantothenic acid)	Man	C	210-280 <sup>c</sup>	Acetylation of sulfanilamide		
2	Man <sup>1</sup>	B	7.0	2.8-11.2 <sup>b</sup>	Manometric determination of CO <sub>2</sub> evolved from pyruvate	
3 Ox <sup>1</sup>	B	5.7	2.7-8.7 <sup>b</sup>			
4 Pigeon <sup>1</sup>	B	20.2	0.6-40 <sup>b</sup>			
5	Man ♂	B	8.9	7.0-14.0 <sup>c</sup>	Microbiological assay	
6	Man ♀	C	10.0 <sup>2</sup>	5.0-8.0 <sup>c</sup>		
7	Cattle, young	B	7.6	7.9-10.1 <sup>c</sup>		
8 Guinea pig	C	6.5 <sup>2</sup>	45-80 <sup>c</sup>			
9 Horse	B		1.1-1.7 <sup>c</sup>			
10 Ox	B	20.5	4.5-5.1 <sup>c</sup>			
11 Rat, albino	B		10-25 <sup>c</sup>			
12 Sheep	B		4.0-4.7 <sup>c</sup>			
13 Swine	B		12-20 <sup>c</sup>			
14 Chicken	B		5.6-11.0 <sup>c</sup>			
15 Frog	B	6.0				
16	Man	C	9.0		Modification of alanine test of Warburg & Christian	
17			75			
18		P	10			
19	Rat, albino	B		50-65 <sup>c</sup>	Microbiological assay	
20	Man	B	3600	2600-4600 <sup>b</sup>		
21		C	7700	6100-9300 <sup>b</sup>	Alkaline acetone method	
22		P	70	20-120 <sup>c</sup>		
23						

/1/ All blood cocarboxylase is intracellular. /2/ Sex difference is P values is significant, males have 1.49  $\mu\text{g}/100$  billion RBC. /3/ Ratio of DPN/TPN in RBC entirely in RBC.

/1/ All blood cocarboxylase is intracellular. /2/ Sex difference in RBC cocarboxylase values is significant, males have 1.49  $\mu\text{g}/100$  billion RBC; females have 1.28  $\mu\text{g}/100$  billion RBC. /3/ Ratio of DPN/TPN in RBC is estimated to be about 8/1, DPN is present almost entirely in RBC.

# 92. BLOOD ELECTROLYTES

Blood (B); R B C. (C); Plasma (P); Serum (S); Colonic Fluid (F)

Constituent	Animal	Electrolytes mEq per 1000 ml <sup>1</sup>		Constituent	Animal	Electrolytes mEq per 1000 ml <sup>1</sup>	
		Value (C)	Range (D)			Value (C)	Range (D)
1	Man	B 83	72-91	Potassium	Duck	B	4.8
2	Man	C 18.6, 23	8.7-28.6, 23		Toad	S	3.9
3	Man	S 1382.3	132-146, 2.3		Man	P	4.8
4	Man	S 1512.3	147-156, 2.3		Man	S	5.2
5	Cat	B (127)	(116-136)		Man	S	4.7-6.1
6	Dog	C 973	90-104, 3		Dog	S	4.2-5.6
7	Dog	P 1503	135-160, 3		Dog	S	3.7-6.8
8	Dog	S 1433	137-149, 3	Calcium <sup>6</sup>	Guinea pig	S	4.8
9	Monkey	C 183	86-109, 3		Guinea pig	S	7.2
10	Monkey	P 183	12.5-27.3		Rabbit	P	6.2
11	Monkey	P 183	138-177, 3		Rat	P	6.2
12	Rabbit	P 1364	47		Rat	S	5.8-6.3
13	Rabbit	S 141	47		Chicken	S	5.6
14	Rat	S 136	133-135		Pigeon	P	5.3
15	Chicken	B (108)	(98-124)		Man	B (3.2)	(3.0-3.7)
16	Chicken	C 163	32.3-23.3		Man	C	5.1
17	Chicken	P 154	140-175, 3		Man	P	1.8
18	Man	B 48	39-62		Cat	P	3.1
19	Man	C 953	89-101, 3		Cat	C	4.4
20	Man	S 4.2	3.6-4.8, 3		Cat	S	2.2
21	Cat	S 4.3	4.0-4.5, 3		Cattle	B	1.7
22	Dog	B (6.0)	(4.9-9.6)		Cattle	C	1.2
23	Dog	C 8.0	4.2-11.8	Magnesium	Cattle	S	2.3
24	Dog	P 4.4	3.7-5.8		Dog	B (2.7)	(2.7)
25	Guinea pig	P 6.5	6.0		Dog	C	3.7
26	Monkey	B 53	46-62		Dog	P	1.87
27	Monkey	C 113	96-131		Dog	S	1.9
28	Monkey	P 6.8	4.9-8.7		Guinea pig	B (5.6)	(5.6)
29	Rabbit	P 5.15	5.4-6.4, 5		Guinea pig	B	8.1
30	Rat	P 5.95	4.8-5.4, 5		Guinea pig	C	4.0
31	Rat	S 5.1	4.8-5.4, 5		Hamster	C	8.3
32	Canary	S 5.8	67		Hamster	P	2.1
33	Chicken	B (43)	(37-50)		Hedgehog	P	2.7
34	Chicken	C 119	106-143		Horse	B	(3.6)
35	Chicken	P 6.0	4.6-6.5				

Values in parentheses ( ) were calculated from contents in blood cells and plasma on basis of hematocrit values. /1/ To convert mEq/1000 ml to mg/100 ml, multiply mEq values by 2.3 for sodium, 3.9 for potassium, 2.0 for calcium, 1.2 for magnesium, 3.3 for chloride, 2.3 for bicarbonate, 1.6 for phosphate, 1.6 for sulfate. /2/ Determined by chemical method. /3/ Determined by flame photometry. /4/ Determined by zinc uranyl acetate (Butler and Rutledge) method. /5/ Determined by colorimetric method of Shohl and Benedict. /6/ Determined by Tisdall method or modification. /7/ Calculated from value of phosphate determined by method in footnote 12, after precipitation of magnesium as magnesium ammonium phosphate as described by Hales.

# 92. BLOOD ELECTROLYTES (Concluded)

Blood (B); R B C (C); Plasma (P); Serum (S); Celomic Fluid (F)

Constituent	Animal	Electrolytes mEq per 1000 ml <sup>1</sup>		Constituent	Animal	Electrolytes mEq per 1000 ml <sup>1</sup>	
		Value	Range			Value	Range
(A)	(B)	(C)	(D)	(A)	(B)	(C)	(D)
70	Horse	C	5.6	105	Homarus vulgaris	B	8.3
71	Horse	P	2.0		Mollusks;		
72	Horse	S	2.2	106	Doris tuberculata	B	116
73	Mouse	C	9.8		Arthropoda*		
74	Mouse	S	6.3		Honeybee larva	B	17.5
75	Pig	B	4.4	107	Limulus	B	99
76	Pig	C	8.4	108	Echinodermata.		
77	Pig	S	2.2		Echinus esculentus	F	101
78	Rabbit	B	6.8-9.2	109	Man	B	82
79	Rabbit	B	3.9-4.5	110	Man	C	52
80	Rabbit	C	5.3-11.2	111	Man	C	55
81	Rat	P	1.7-2.5	112	Man	C	101
82	Rat	B	(3.7-5.8)	113	Man	C	103
83	Rat	C	5.8-8.4	114	Man	C	(97)
84	Rat	P	7.1	115	Dog	C	65
85	Rat	S	2.0-3.7b	116	Dog	C	61-69b
86	Rat	S	2.4-2.8b	117	Dog	C	99-1108
87	Sheep	B	2.2-2.4	118	Monkey	B	83-110
88	Sheep	C	0.9-2.6	119	Monkey	C	56-73
89	Chicken	P	1.7-2.1	120	Monkey	P	103-118
90	Chicken	C	(4.0)	121	Monkey	P	1059
91	Chicken	P	7.7	122	Rabbit	P	100
92	Pigeon	B	2.3	123	Rabbit	P	122
93	Pigeon	C	(17)	124	Chicken	P	116-140
94	Pigeon	P	36	125	Man	S	28-10
95	Turtle	B	2.0	126	Man	S	25-3110
96	Turtle	C	(6.6)	127	Dog	S	24-2910
97	Turtle	C	20.1	128	Dog	P	20.5-10
98	Frog	P	2.8	129	Rabbit	P	28
99	Frog	B	(5.6)	130	Rat	P	22-11
100	Frog	C	13.2	131	Rat	P	16.1-25.3b
101	Frog	S	2.3	132	Man	S	2.1
102	Salmon	S	1.8	133	Dog	P	1.7-2.7
103	Sphenoides asculatus	S	3.4	134	Rabbit	P	1.3-2.0
104	Lepidosteus osseus	S	9.1	135	Rat	P	5.8-8.2
	Crustacea:			136	Man	P	1.0-1.5
	Maja verrucosa	B	113	137	Dog	P	2.0
				138	Dog	P	14.5
					Dog	P	12.0-19.0
					Dog	P	1.9-3.6
					Dog	P	0.1-0.2

/1/ Determined by absorption indicator method of Selfer and Kornblum and checked by method of Van Slyke and Sendroy. /2/ Determined by mercurimetric method of Schales and Schales. /10/ Determined by method of Van Slyke and Neil. /11/ Determined by micro-diffusion method.

# 93. BLOOD MINOR MINERALS

		Blood (B), R B C (C), Plasma (P), Serum (S); Leukocytes (L)									
Constituent		Animal		$\mu\text{g}/100 \text{ ml}$				Constituent		Animal	
(A)		(B)		Value	Range			(A)	(B)	$\mu\text{g}/100 \text{ ml}$	
				(C)	(D)					Value	Range
										(C)	(D)
1	Aluminum <sup>1</sup>	Man	B	15	-40	30	Iodine, precipitable	Man	S	5.0	3.8-7.1
2		Man	C	7	-17	31		Rat	S	2.2	1.5-3.4
3		Man	P	46	-88	32		Man	B	(48)	(43-52)
4	Bromine <sup>2</sup>	Man	S			33		Man	P	105	32-177
5		Man	B	943	0.7-1.0	34		Pig	P	169	91-247
6		Man	C	753	73-1153	35		Rat	P	2613	
7		Man	P	1103	49-1013	36		Man	B	29	18-49
8		Man	B		76-1433	37		Man	C	57	29-86
9		Cow	S		820-1400	38		Man	P	39	-7.8
10		Ewe	B		730-1450	39		Rabbit	B	15	
11		Hen	S	14	34-37	40		Rabbit	P	13	0-25
12	Copper	Horse	S	36	31-41	41		Man	C	19	-48
13		Ox	S	38	33-35	42		Man	P	8	-19
14		Pig	S	34	153-2593	43		Man	B	17	33-63
15		Rabbit	P	206	74-99	44		Dog	B	50	
16		Rabbit	B	85	51-86	45		Horse	B	23	
17		Rat	C	67		46		Horse	S	1298	
18		Rat	S	320		47		Rabbit	B	1188	
19		Man	B	(28)	(11-45)	48		Rabbit	P	100	
20	Fluorine	Man	C	27	11-44	49		Man	B,C	+	
21		Man	P	28	10-45	50		Man	B	22	-40
22		Dog	C	174	9-244	51		Man	C	26	-64
23		Dog	P	254	12-354	52		Man	P	4	-10
24		Man	B	7.75	3-135	53		Man	B	880	488-1272
25		Man	S	7.1	4.8-8.6	54		Man	C	1440	911-1969
26		Chicken	P	7.2		55		Man	P	300	0-613
27		Dog	S	29	14-52	56		Man	L	3.2 $\mu\text{g}/100 \text{ million cells}$	
28		Guinea pig	B	7.3		57		Man	B	22	-40
29		Mouse	P	4.5		58		Man	C	26	-64
		Rat	P	3.4	3.3-3.5			Man	P	4	-10
								Man	B	880	488-1272
								Man	C	1440	911-1969
								Man	P	300	0-613
								Man	L	3.2 $\mu\text{g}/100 \text{ million cells}$	

/1/ Determined by spectrochemical methods after ashing. These values relative to volume, derived from gravimetric methods on basis of assumed blood specific gravity of 1.056. /2/ Gold chloride added to filtrate. /3/ Determined by unpublished method of Gubler, Lahey, Ashenbrucker, Wintrobe, and Cartwright. /4/ Micrograms per 100 E. /5/ After alkaline ashing. /6/ Determined by methods of Kitzes, Elvehjem, and Schutte, and of Barkan and Walker. /7/ Heparinized blood stored in cellophane tube, unless specified otherwise. /8/ Defibrinated blood. /9/ Determined by modified dithione method.

94 BLOOD GASES MAN<sup>3</sup>

Variable		Whole Blood			ml Gas in 100 ml RBC	ml Gas in 100 ml Plasma	Blood Gas Pressure (Tension)	Factors and Constants	
		ml Gas in 100 ml Whole Blood	ml Gas in 450 ml <sup>2</sup> RBC	ml Gas in 550 ml <sup>2</sup> Plasma					
(A)		(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
Oxygen Capacity <sup>5</sup>		20.46 18.04 <sup>7</sup>			145.3 <sup>8</sup> 145.3 <sup>9</sup>				
Total Oxygen	Arterial <sup>1</sup>	20.3 <sup>10,11</sup> 17.9	20.1 17.7	0.142 0.155	44.7 44.7	0.258 0.258	94 94	A-V O <sub>2</sub> Diff	
								5.60	
	Venous <sup>12</sup>	15.3 13.7	15.2 13.6	0.060 0.068	33.9 34.4	0.110 0.113	40.3 41.3	4.20	
"Free" Oxygen	Arterial <sup>1</sup>	0.28 <sup>10</sup> 0.282	0.144 0.126	0.142 0.156	0.319 0.319	0.258 0.258	94 94	a <sub>O2</sub> Cells <sup>14</sup> a <sub>O2</sub> Plasma <sup>14</sup>	
								0.0258 <sup>14</sup>	0.02089 <sup>14</sup>
	Venous <sup>12</sup>	0.122 0.124	0.061 0.055	0.061 0.068	0.136 0.139	0.110 0.113	40.3 41.3		
Combined Oxygen (HbO <sub>2</sub> )	Arterial <sup>1</sup>	20.0 17.6	20.0 17.6	0 0	44.4 44.4	0 0	94 94	HbO <sub>2</sub> Sat. %	
								98 98	
	Venous <sup>12</sup>	15.2 13.6	15.2 13.6	0 0	33.6 34.3	0 0	40.3 41.3	73.5 74.5	
Total Carbon Dioxide	Arterial <sup>1</sup>	49.0 48.0 <sup>17</sup>	16.2 13.6	32.8 34.4	36.0 34.3	59.6 <sup>15</sup> 57.0 <sup>17</sup>	41 39.19	A-V CO <sub>2</sub> Diff <sup>16</sup>	
								1.217 1.187	4.1 3.4
	Venous <sup>12</sup>	53.1 51.4	18.0 14.9	55.1 56.5	40.1 37.7	63.8 60.4 <sup>17</sup>	46.5 <sup>19</sup> 43 <sup>19</sup>	1.201 1.175	
"Free" Carbon Dioxide	Arterial <sup>1</sup>	2.624 2.532	1.060 0.891	1.564 1.641	2.356 2.250	2.844 2.717	41 39.19	a <sub>CO2</sub> Cells <sup>14</sup> a <sub>CO2</sub> Plasma <sup>14</sup>	
								0.4399 <sup>20</sup>	0.5311 <sup>20</sup>
	Venous <sup>12</sup>	2.997 2.785	1.211 0.965	1.786 1.820	2.690 2.437	3.248 3.013	46.5 <sup>19</sup> 43 <sup>19</sup>		
Total Combined Carbon Dioxide	Arterial <sup>1</sup>	46.4 45.5	15.1 12.7	31.2 32.8	33.6 32.1	56.8 54.3	41 39	pH Cells <sup>22</sup> pH Plasma <sup>22</sup>	
								7.19 7.20	7.40 7.409
	Venous <sup>12</sup>	50.1 48.6	16.8 14.0	33.3 34.7	37.4 35.3	60.5 <sup>21</sup> 57.4	46.5 43	7.17 7.18	7.37 <sup>23</sup> 7.38 <sup>23</sup>
Carbamino Carbon Dioxide	Arterial <sup>1</sup>	2.2 <sup>20</sup> 1.9 <sup>20</sup>	1.7 1.5	0.4 0.5	3.9 3.7	0.8 0.8	41 39	"c" Cells <sup>22</sup> "c" Plasma <sup>22</sup>	
								0.115 0.116	0.014 0.014
	Venous <sup>12</sup>	3.1 2.7	2.6 2.2	0.5 0.5	5.8 5.5	0.8 0.8	46.5 43	0.154 0.156	0.014 0.014
Lithonate Carbon Dioxide	Arterial <sup>1</sup>	44.2 43.6	13.4 11.2	30.8 32.3	29.8 28.3	56.0 53.5	41 39	a <sub>N2</sub> Cells <sup>24</sup> a <sub>N2</sub> Plasma <sup>24</sup>	
								0.0146 <sup>24</sup>	0.0117 <sup>24</sup>
	Venous <sup>12</sup>	47.0 46.0	14.2 11.8	32.8 34.2	31.6 29.4	59.7 56.6	46.5 43		
Nitrogen	Arterial <sup>1</sup>	0.979 0.970	0.494 0.437	0.484 0.534	1.099 1.103	0.881 0.884	572.3 574.3		
	Venous <sup>12</sup>	0.979 0.970	0.494 0.437	0.484 0.534	1.099 1.103	0.881 0.884	572.3 574.3		

1/ The values from which this table has been synthesized are in many instances derived by calculation from basic assumptions, factors and constants, and do not have the same validity as actually measured values. Those for women are in general less well founded than for men. Values for carbamino CO<sub>2</sub> in particular, indicate order of magnitude rather than exact magnitudes. 2/ Assumed normal values, lower than hematological standards in Table 37, partially due to rise in plasma volume in recumbent posture. (vs. standing). 3/ Exact value used in calculations = 39.6 and 60.4. 4/ 100ml RBC in contact with plasma, and 100ml plasma in contact with RBC. 5/ O<sub>2</sub> capacity = grams Hb x 1.34. This factor based on hemoglobin Fe content of 0.33%. 6/ Based on assumed hemoglobin content of 15.5 g/dl. 7/ Based on assumed hemoglobin content of 15.5 g/dl. 8/ Based on assumed hemoglobin content of 15.5 g/dl. 9/ Based on assumed hemoglobin content of 15.5 g/dl. 10/ Based on assumed hemoglobin content of 15.5 g/dl. 11/ Exact value used in calculations = 39.6 and 60.4. 12/ Venous values are based on assumed venous blood gas tensions. 13/ Exact value used in calculations = 39.6 and 60.4. 14/ Based on assumed hemoglobin content of 15.5 g/dl. 15/ Based on assumed hemoglobin content of 15.5 g/dl. 16/ Based on assumed hemoglobin content of 15.5 g/dl. 17/ Exact value used in calculations = 39.6 and 60.4. 18/ Based on assumed hemoglobin content of 15.5 g/dl. 19/ Based on assumed hemoglobin content of 15.5 g/dl. 20/ Based on assumed hemoglobin content of 15.5 g/dl. 21/ Based on assumed hemoglobin content of 15.5 g/dl. 22/ Based on assumed hemoglobin content of 15.5 g/dl. 23/ Based on assumed hemoglobin content of 15.5 g/dl. 24/ Based on assumed hemoglobin content of 15.5 g/dl.

for plasma, "c" is calculated from K<sub>a</sub> carbamino (x1.2) and the mEq of plasma proteins (x17.0). 23/ Arterial N<sub>2</sub> pressure taken as equal to alveolar air H<sub>2</sub> pressure, the latter calculated by difference between 760 and sum of following: O<sub>2</sub> = 105; CO<sub>2</sub> = 40; 5% water vapor = 47. Venous N<sub>2</sub> taken as equal to arterial. Note that the total of blood gases and water vapor falls short of 760 in both arterial and venous bloods. 24/ ml N<sub>2</sub> dissolved in 100ml RBC = 0.016 x 100 x N<sub>2</sub> pressure/760. For plasma substitute 0.0117 for 0.016.

# 95. ARTERIAL BLOOD GASES, O<sub>2</sub> AND CO<sub>2</sub>, AT ALTITUDE: MAN

"Ascent" in a Low Pressure Chamber  
For a Period Not Longer Than One Hour

Altitude <sup>1</sup>	Arterial Blood Oxygen				Arterial Blood Carbon Dioxide	
	Tension <sup>2</sup> mm Hg	Content <sup>2</sup> ml/100 ml	Capacity <sup>2</sup> ml/100ml	Saturation <sup>2</sup> %	Tension <sup>3</sup> mm Hg	Content <sup>3</sup> ml/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)
1 Sea Level	94	21.1		98	41	49
2 760 mm Hg						
3 3000 ft						
4 1534 m	66	19.6		91	36.5	
5 632 mm Hg				87-95b		
6 8000 ft	60	19.1		89	37.4	
7 2456 m				84.5-93.5b		
8 564 mm Hg						
9 10000 ft	55	18.4		85.4	35.8	
10 3048 m				79-92b		
11 523 mm Hg						
12 12000 ft	52	18.3		84.9	34.8	
13 3658 m				77-92.5b		
14 483 mm Hg						
15 14000 ft	44	17.0		79.2	35.4	
16 4267 m				71-87.5b		
17 446 mm Hg						
18 16000 ft	41	16.4		76.2	33.8	
19 4877 m				65-87.5b		
20 13000 ft	36	15.3		71.2	31.8	
21 4486 m				57-85.5b		
22 379 mm Hg						
23 20000 ft	35	15.2		70.8	29.4	
24 6096 m				57.5-84b		
25 349 mm Hg						

A uniform O<sub>2</sub> capacity of 21.5 ml/100ml and therefore a uniform Hb content of 15.8 g/100ml are assumed.<sup>4</sup>

A uniform O<sub>2</sub> capacity of 21.5 ml/100ml and therefore a uniform Hb content of 15.8 g/100ml are assumed.<sup>4</sup>

"Ascent" to Altitudes (For a Period Not Longer Than One hour)  
Which Necessitate the Use of Pure Oxygen

Altitude <sup>1</sup>	Arterial Blood Oxygen				Arterial Blood Carbon Dioxide	
	Tension <sup>2</sup> mm Hg	Content <sup>2</sup> ml/100 ml	Capacity <sup>2</sup> ml/100ml	Saturation <sup>2</sup> %	Tension <sup>3</sup> mm Hg	Content <sup>3</sup> ml/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)
1 Sea Level		19.8		92		
2 760 mm Hg				84-100b		
3 3000 ft	74	20.2		94	40.6	46.3
4 1534 m						
5 632 mm Hg	57	19.1		88.7	39.4	50.0
6 8000 ft						
7 2456 m	55	18.9		86.1	35	42.7
8 564 mm Hg				81-95b	28-44b	35-50b
9 10000 ft	54	18.6		86.4	38.1	44.8
10 3048 m				85-88b		
11 523 mm Hg	49	17.8		83	40	47.1
12 12000 ft				71-95b	36-44b	45-50b
13 3658 m	42	16.9		78.5		41.5
14 483 mm Hg				65-92b		31-52b
15 14000 ft	36	15.5		72.2	33.2	44.9
16 4267 m				58-86b		
17 446 mm Hg						
18 16000 ft		14.6		68		
19 4877 m				53-83b		

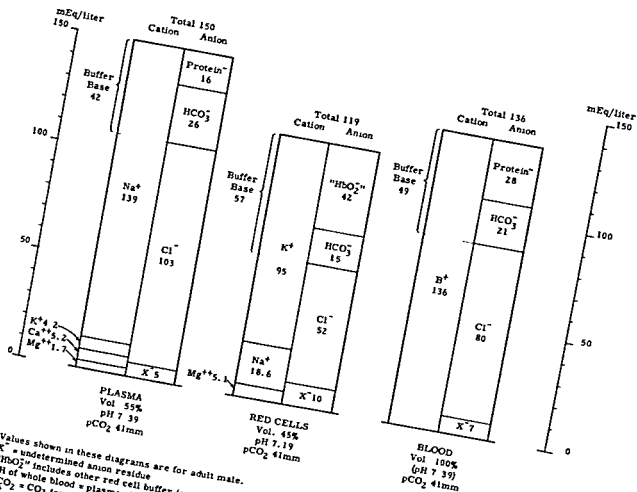


# 95. ARTERIAL BLOOD GASES, O<sub>2</sub> AND CO<sub>2</sub>, AT ALTITUDE: MAN (Concluded)

Permanent Residents in Mountainous Regions (Complete Acclimatization)							Newcomers <sup>6</sup> to Mountainous Regions (Incomplete Acclimatization)						
Altitude <sup>3</sup>	Arterial Blood Oxygen				Arterial Blood Carbon Dioxide		Altitude <sup>5</sup>	Arterial Blood Oxygen				Arterial Blood Carbon Dioxide	
	Tension <sup>1</sup> mm Hg	Content <sup>2</sup> ml/100ml	Capacity <sup>2</sup> ml/100ml	Saturation %	Tension <sup>1</sup> mm Hg	Content <sup>2</sup> ml/100ml		Tension <sup>1</sup> mm Hg	Content <sup>2</sup> ml/100ml	Capacity <sup>2</sup> ml/100ml	Saturation %	Tension <sup>1</sup> mm Hg	Content <sup>2</sup> ml/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(A)	(B)	(C)	(D)	(E)	(F)	(G)
492 ft <sup>7</sup>							11319 ft						
34, 150 m	90	20.7	21.7	95.4	41	46	46, 3450 m	55	20.5	24.1	85	31	41
746-752 mm Hg							496-496 mm Hg						
7840 ft							15421 ft						
35, 2390 m	68	21.2	23.1	91.7	37.8	41.1	47, 4700 m	44	18.7	24.1	78	29.3	38.3
36, 668-581 mm Hg		18.5-24b	19-27.5b	86.5-97b	34-42b	37-45b	429-429 mm Hg						
10300 ft	66	21.8	24.0	91.0	36.4	39.3	17521 ft	43	18.6	24.5	76.2	27.7	35.0
37, 1140 m		19-25b	22-26b	87-95b	31-42b	34.5-44b	48, 5340 m						
38, 517-511 mm Hg							2045 ft						
12238 ft	57	21.9	25.0	87.6	36.0	33-39b	6140 m	35	16.3	24.9	65.6	24.2	30.2
40, 479-482 mm Hg		18.5-25b	21.5-28.5b	84.5-91.5b			347-356 mm Hg						
14896 ft													
41, 4540 m	47	23.0	28.3	81.4	34.7	33.5							
42, 431-448 mm Hg		19.5-26.5b	24-32.5b	75.5-87b	29-40b	32-35b							
15950 ft													
43, 4860 m	46	23.4	29.0	80.7	33.0	34.0							
44, 413-429 mm Hg		20.5-26.5b	25-33b	76-85b	28-38b	31-37b							
17521 ft	43	23.0	30.2	76.2	29.3	31.8							
45, 5340 m													
387-401 mm Hg													

/1/ U.S. Standard Atmosphere. /2/ Values are for combined O<sub>2</sub> only, and do not include physically dissolved O<sub>2</sub>. /3/ Excepting IF, values represent alveolar tensions. /4/ 15.8 g Hb/100ml blood x 1.36 = ml O<sub>2</sub>. /5/ Observed altitudes. First barometric pressure is U.S. Standard for observed altitude, second is average observed barometric pressure at time of measurements. /6/ Up to 16 days. Upon exposure of only 2 hours, values in columns C,D,E tend to be lower, in F,G, tend to be higher. /7/ Approximate sea level values for comparison.

# 96. IONIC BALANCE AND BUFFER BASE, ARTERIAL BLOOD, CELLS AND PLASMA: MAN



Values shown in these diagrams are for adult male.  
 X<sup>-</sup> = undetermined anion residue  
 "HbO<sub>2</sub><sup>-</sup>" includes other red cell buffer ions, such as organic phosphate  
 pH of whole blood = plasma pH  
 pCO<sub>2</sub> = CO<sub>2</sub> tension.  
 B<sup>+</sup> = mEq total base (Na<sup>+</sup>, K<sup>+</sup>, etc.) in one liter blood, on basis of hematocrit value of 45% red cells.  
 Buffer Base = that quantity of total base equivalent in amount (in terms of mEq) to the labile portion of the total anions, i.e., proteinate, bicarbonate, oxyhemoglobinate, organic phosphate and other red cell buffer ions.

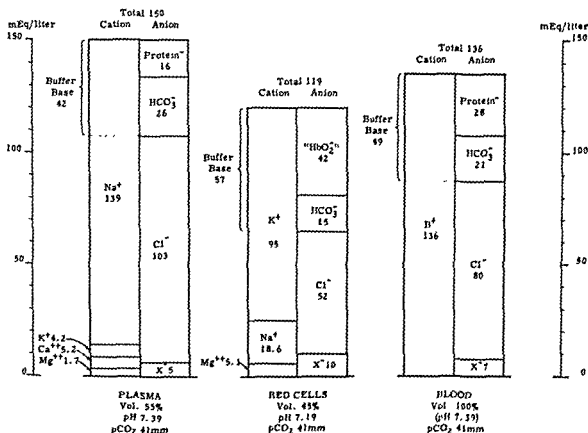
# 95. ARTERIAL BLOOD GASES, O<sub>2</sub> AND CO<sub>2</sub>, AT ALTITUDE: MAN (Concluded)

Newcomers to Mountainous Regions (Incomplete Acclimatization)									
Arterial Blood Oxygen					Arterial Blood Carbon Dioxide				
Altitude <sup>5</sup>	Tension mm Hg (B)	Content <sup>2</sup> ml/100ml (C)	Capacity <sup>2</sup> ml/100ml (D)	Saturation % (E)	Tension <sup>1</sup> mm Hg (F)	Content <sup>1</sup> ml/100ml (G)	Tension <sup>1</sup> mm Hg (H)	Content <sup>1</sup> ml/100ml (I)	Tension <sup>1</sup> mm Hg (J)
(A)									
492 ft <sup>7</sup>	90	20.7	21.7	95.4	41	46			
1450 m									
1466-1522 mm Hg									
1466 ft	68	21.2	22.1	91.7	37.8	41.1			
2490 m									
2490 ft	66	18.5-24.6	19-27.5 <sup>b</sup>	86.5-97 <sup>b</sup>	34-42 <sup>b</sup>	37-45 <sup>b</sup>			
3488-3811 mm Hg									
3488 ft	66	21.6	24.0	91.0	36.4	39.3			
10300 ft									
37140 m									
37140 ft	66	19-25 <sup>b</sup>	22-26 <sup>b</sup>	87-95 <sup>b</sup>	31-42 <sup>b</sup>	34.5-44 <sup>b</sup>			
517-531 mm Hg									
517 ft	57	21.9	25.0	87.6	36.0	33-39 <sup>b</sup>			
12238 ft									
37370 m									
37370 ft	57	18.5-25 <sup>b</sup>	21.5-28.5 <sup>b</sup>	84.5-91.5 <sup>b</sup>	34.7	33.5			
479-482 mm Hg									
479 ft	47	23.0	28.3 <sup>b</sup>	81.4	29-40 <sup>b</sup>	32-35 <sup>b</sup>			
14896 ft									
41450 m									
41450 ft	47	19.5-16.5 <sup>b</sup>	24-31 <sup>b</sup>	75.5-87 <sup>b</sup>	33.0	34.0			
431-448 mm Hg									
431 ft	46	23.4	29.0	80.7	28-38 <sup>b</sup>	31-37 <sup>b</sup>			
15950 ft									
43480 m									
43480 ft	46	20.5-26.5 <sup>b</sup>	25-35 <sup>b</sup>	76-85 <sup>b</sup>	29.3	31.8			
413-429 mm Hg									
413 ft	43	23.0	30.2	76.2					
17521 ft									
5340 m									
5340 ft	43								
387-401 mm Hg									
387 ft									

1/ U. S. Standard Atmosphere. 2/ Values are for combined O<sub>2</sub> only, and do not include physically dissolved O<sub>2</sub>. 3/ Excepting 1F, values represent alveolar tensions. 4/ 15.8 g/100ml blood x 1.36 = ml O<sub>2</sub>. 5/ Observed altitudes. 6/ Up to 16 days. Upon exposure of only 2 hours, values in columns C, D, E tend to be lower; in F, G, tend to be higher. 7/ Approximate sea level values for comparison.

Arterial Blood Oxygen  
Tension mm Hg (B)  
Content ml/100ml (C)  
Capacity ml/100ml (D)  
Saturation % (E)  
Tension mm Hg (F)  
Content ml/100ml (G)  
Capacity ml/100ml (H)  
Saturation % (I)  
Tension mm Hg (J)  
Content ml/100ml (K)  
Capacity ml/100ml (L)  
Saturation % (M)

## 96. IONIC BALANCE AND BUFFER BASE, ARTERIAL BLOOD, CELLS AND PLASMA: MAN



Values shown in these diagrams are for adult male

X<sup>-</sup> = undetermined anion residue.

"HbO<sub>2</sub>"<sup>-</sup> includes other red cell buffer ions, such as organic phosphate.

pH of whole blood = plasma pH.

pCO<sub>2</sub> = CO<sub>2</sub> tension

B<sup>+</sup> = mEq total base (Na<sup>+</sup>, K<sup>+</sup>, etc.) in one liter blood, on basis of hematocrit value of 45% red cells.

Buffer Base = that quantity of total base equivalent in amount (in terms of mEq) to the labile portion of the total anions, i.e., proteinate, bicarbonate, oxyhemoglobinate, organic phosphate and other red cell buffer ions.

997. ACID-BASE VALUES, BLOOD AND PLASMA: MAN<sup>1</sup>

[illegible][illegible]

# 98. EFFECTS OF RADIATION ON PERIPHERAL BLOOD

EFFECTS OF RADIATION ON PERIPHERAL BLOOD						
	Type Radiation (A)	Application (B)		Accumulated Dose or Exposure Time (D)	Effects (E)	
			(C)			
1	X-ray (200 KVP, 400 KVP, 1000 KVP)	Chronic, total body	15-20 r	40 r	Decreased lymphocytes	
2		Chronic, total body	5-10 r/day	200-300 r	Decreased lymphocytes	
3		Chronic, total body	5-20 r/day	88-300 r	Decreased lymphocytes, neutrophils, WBC, RBC, Hb, increased monocytes	
4		Chronic, total body		? dose. Pre- sumably greater than 0.1 r/day	Increased incidence of leukemia	
5	High energy gamma	Chronic, total body	0.2 r/week	77 weeks	Decreased WBC, absolute lymphocytes, absolute neutrophils	
6	200 KVP x-ray	Dog			Decreased absolute lymphocytes	
7		Acute, total body	20 r		Decreased: absolute lymphocytes platelets, WBC, granulocytes	
8	X-ray (250 KVP, 1000 KVP)	Acute, total body	50 r		Decreased absolute lymphocytes, reti- culocytes	
9		Acute, total body	200 r		Decreased lymphocytes	
10	X-ray (250 KVP, 1000 KVP)	Chronic, total body	0.5 r/day 6 days/week	2 years	Decreased lymphocytes, WBC, neutrophils	
11		Chronic, total body	1.0 r/day 6 days/week	1- year	Decreased absolute lymphocytes, platelets, reticulocytes	
12	X-ray (200 KVP)	Chronic, total body	3 r/day 6 days/week	1-2 mo.	Decreased absolute lymphocytes, WBC, neutrophils	
13		Chronic, total body	6 r/day 6 days/week	1-2 mo	Decreased absolute lymphocytes, platelets	
14	Fast neutrons	Chronic, total body	10 r/day 6 days/week	1 month	Decreased WBC, absolute, lympho- cytes, absolute neutrophils, platelets	
15		Chronic, total body	12.5 r/day	1262 r	Decreased absolute lymphocytes, absolute neutrophils, WBC, RBC, Hb, reticulocytes	
16	X-ray (200 KVP)	Chronic, total body	1 7 n/day 6 days/week	1 year	Aplastic anemia	
17		Chronic, total body	1 7 n/day 6 days/week	3-4 weeks	Decreased WBC, absolute lympho- cytes, absolute neutrophils, platelets, RBC	
18	Goat	Acute, total body	300 r		Decreased WBC, absolute lympho- cytes, absolute neutrophils	
19					Decreased lymphocytes, neutrophils, WBC, RBC, Hb.	

# 98. EFFECTS OF RADIATION ON PERIPHERAL BLOOD (Continued)

Type Radiation		Application		Accumulated Dose or Exposure Time	Effects
(A)		(B)	(C)	(D)	(E)
Guinea pig					
18	Gamma (radium)	Chronic, total body	0.11 r/day	approx. 1 mo.	Decreased: lymphocytes, WBC, heterophils
19		Chronic, total body	1.1 r/day	approx. 1 mo.	Decreased: lymphocytes, WBC, heterophils
20		Chronic, total body	1.1 r/day	approx. 1 year	Decreased: lymphocytes, WBC, heterophils, (platelets, in females only)
21		Chronic, total body	2.2 r/day	approx. 1 mo.	Decreased: lymphocytes, WBC, heterophils
22		Chronic, total body	2.2 r/day	approx. 1 year	Decreased: lymphocytes, WBC, heterophils, platelets
23		Chronic, total body	2.2 r/day	78 weeks	Decreased: lymphocytes, WBC, heterophils, (RBC, Hb, males only)
24		Chronic, total body	4.4 r/day	approx. 1 mo.	Decreased: lymphocytes, WBC, heterophils
25		Chronic, total body	4.4 r/day	approx. 1 year	Decreased: lymphocytes, WBC, heterophils, platelets, RBC, Hb.
26		Chronic, total body	8.8 r/day	10 days	Decreased: lymphocytes, WBC
27		Chronic, total body	8.8 r/day	approx. 1 mo	Decreased: lymphocytes, WBC, heterophils, platelets
28		Chronic, total body	8.8 r/day	19 weeks	Decreased: lymphocytes, WBC, heterophils, RBC, Hb.
Mouse					
29	X-ray (200 KVP)	Acute, total body	50 r		Decreased: lymphocytes, WBC
30	X-ray (165 KV)	Acute, total body	400 r		Decreased: lymphocytes, WBC, granulocytes, reticulocytes, RBC, Hb.
31	Fast neutrons (cyclotron)	Chronic, total body	0.07 n/day 5 days/week	30 n	Decreased: WBC
32		Chronic, total body	1.4 n/day 5 days/week	230 n	Decreased: absolute lymphocytes, WBC, RBC, Hb.
33					Increased: frequency of WBC below 15,000/mm <sup>3</sup>
34					Shift to left present in heterophils
35	Gamma (radium)	Chronic, total body	2.2 r/day	29 weeks	Decreased: WBC, absolute lymphocytes
36		Chronic, total body	4.4 r/day	29 weeks	Decreased: WBC, absolute lymphocytes
37		Chronic, total body	4.4 r/day	78 weeks	Decreased: WBC, absolute lymphocytes, (RBC, Hb, platelets, males only)
38		Chronic, total body	8.8 r/day	29 weeks	Decreased: WBC, absolute lymphocytes
39		Chronic, total body	8.8 r/day	29 weeks	Decreased: WBC, absolute lymphocytes, RBC, Hb, platelets

# 98. EFFECTS OF RADIATION ON PERIPHERAL BLOOD (Continued)

EFFECTS OF RADIATION ON PERIPHERAL BLOOD (Continued)					
Type Radiation (A)	Application (B)		Accumulated Dose or Exposure Time (D)	Effects (E)	
		(C)			
			Rabbit		
40		Achle, total body	25 r		
41	X-ray (8200 KVP)	Acute, total body	100 r		Decreased: absolute lymphocytes
42		Acute, total body	500 r		Decreased: absolute lymphocytes, WBC, platelets, reticulocytes, granulocytes
43	X-ray (250 KVP, 1000 KVP)	Chronic, total body	10 r/day 6 days/week	60-120 r	Decreased absolute lymphocytes, WBC platelets, reticulocytes, granulo- cytes, RBC, Hb.
44		Chronic, total body	10 r/day 6 days/week	720-960 r	Decreased absolute lymphocytes, absolute neutrophils, WBC
45	Fast neutrons (cyclotron)	Chronic, total body	1.7 n/day 6 days/week	15-45 n	Decreased WBC, absolute lymphocytes, absolute neutrophils, RBC, platelets
46		Chronic, total body	1.7 n/day 6 days/week	50-75 n	Decreased absolute lymphocytes, WBC
47		Acute, total body	9 n		Decreased absolute lymphocytes, absolute neutrophils, WBC
48	Fast neutrons (pile)	Acute, total body	26 n		Decreased absolute lymphocytes
49		Acute, total body	68-76 n		Decreased absolute lymphocytes and heterophils
50		Acute, total body	128 n		Decreased absolute lymphocytes, heterophils, and platelets
51		Chronic total body	1.1 r/day	approx 2 mos	Decreased absolute lymphocytes, heterophils, platelets, RBC, Hb
52		Chronic, total body	2.2 r/day	approx 2 mos	Decreased absolute lymphocytes
53		Chronic, total body	2.2 r/day	approx 3 mos	Decreased absolute lymphocytes
54	Gamma (radium) (females only)	Chronic total body	4.4 r/day	approx 2 mos	Decreased absolute lymphocytes, platelets
55		Chronic, total body	4.4 r/day	approx 9 mos	Decreased absolute lymphocytes
56		Chronic, total body	8.8 r/day	approx 2 mos	Decreased absolute lymphocytes, platelets
57		Chronic, total body	8.8 r/day	approx 9 mos	Decreased absolute lymphocytes, platelets
58					



# 98. EFFECTS OF RADIATION ON PERIPHERAL BLOOD (Concluded)

Type Radiation		Application		Accumulated Dose or Exposure Time	Effects
(A)		(B)	(C)	(D)	(E)
Rat					
59	250 KVP X-ray	Acute, total body	5 r		Decreased: absolute lymphocytes
60		Acute, total body	10 r		Decreased: absolute lymphocytes, WBC
61		Acute, total body	50 r		Decreased: neutrophils, WBC, absolute lymphocytes, reticulocytes
62		Acute, total body	100 r		Decreased absolute lymphocytes, neutrophils, WBC, reticulocytes, platelets
63		Acute, total body	300 r		Decreased absolute lymphocytes, neutrophils, WBC, reticulocytes, platelets, RBC, Hb
64	X-ray (250-1000 KVP)	Chronic, total body	0.5 r/day	2 years	Decreased absolute lymphocytes
65		Chronic, total body	1 r/day	1 year	Decreased absolute lymphocytes
66	Fast neutrons	Chronic, total body	1.7 n/day 5 days/week	1 mo.	Decreased absolute lymphocytes
67		Chronic, total body	1.7 n/day 5 days/week	1 year	Decreased absolute lymphocytes, RBC, Hb.
Chick					
68	X-ray (85 KV)	Acute, total body	360 r		Decreased lymphocytes
69	X-ray (200 KVP)	Acute, total body	600 r		Decreased lymphocytes, WBC, heterophils, thrombocytes, RBC, Hb, reticulocytes
Salmon					
70	X-ray (200 KV)	Acute, total body	750 r		Decreased nucleated cells in peripheral blood

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES

RADIATION ON HEMATOPOIETIC TISSUES						
Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>	
(A)	(B)	(C)	(D)	(E)	(F)	
Man						
1	X-ray (150-175 KV)	100 r. Acute, localized (therapeutic irradiation single dose)	Sternal marrow	Decreased erythroblasts	2 days	complete by one month
2		200 r. Acute, localized (therapeutic irradiation single dose)	Sternal marrow	Decreased myeloid cells	3 days	complete by one month
3				Decreased erythroblasts	2 days	----
4		300 r. Acute, localized (therapeutic irradiation single dose)	Sternal marrow	Decreased myeloid cells	2 days	----
5				Decreased erythroblast mitosis	2 days	4 days
6		2000 r. Acute localized (therapeutic irradiation single dose)	Sternal marrow	Decreased mitosis	1 day	1-2 months
7				Decreased erythroid cells	1 day	1-2 months
8		3000 r. or more, Acute, localized (therapeutic irradiation single dose)	Sternal marrow	Decreased myeloid cells	1 day	1-2 months
9				Transient aplasia	----	----
10				Permanent aplasia	----	----
Dog						
11	X-ray (250 KVP)	300 r. Acute, total body	Bone marrow	Decreased erythroid cells	As early as 2 days post-radiation	24-28 days
12				Decreased myeloid cells	2 days	24 days
13				Reversed myeloid-erythroid ratio	2 days	4 days
Guinea pig						
14	X-ray (200 KVP)	175 r. Acute, total body	Lymph node	Increased debris	7-9 days	40 days
15				Slight decreased cellularity	3 hrs	24 hrs.
16				Increased heterophils	3-8 hrs	by 4 days
17				3 hrs	48 hrs	

<sup>1</sup>/ Time represents an approximation. The figures given have been determined, in large part, by f

1/ Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Continued)

	Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>
	(A)	(B)	(C)	(D)	(E)	(F)
	Mouse					
18	X-ray (200 KVP)	50 r, Acute, total body	Bone marrow	Increased myeloid cells	8 hrs.	----
19				Increased myeloblasts	12 hrs.	----
20		350 r, Acute, total body	Bone marrow	Increased debris	3 hrs.	5 days
21				Decreased erythropoiesis	3 hrs.	5-9 days
22				Decreased myelopoiesis	3 hrs.	5 days
23	X-ray (186 KV)	400 r, Acute, total body	Bone marrow	Decreased lymphocytes	1 day	----
24				Decreased normoblasts	1 day	7 days
25				Decreased cellularity	1 day	7 days
26				Decreased myelopoiesis	2 days	7-14 days
27				Shift to left present in myelocytic series	10 days	----
28	Fast neutrons (pile)	117 n, Acute, total body	Bone marrow	Cell death	3 hrs.	----
29				Decreased cellularity	3 hrs.	5-9 days
30				Increased debris	3 hrs.	----
31				Cytological damage, megakaryocytes	1 day	---- <sup>2</sup>
32		96 n, Acute, total body	Bone marrow	Increased spindle cells	9 days	---- <sup>2</sup>
33	Increased debris			3 hrs.	----	
34	Decreased cellularity			3 hrs.	3 days	
35	X-ray (200 KVP)	50 r, Acute, total body	Lymph nodes and spleen	Occurrence of giant cells	4 hrs.	----
36				Increased debris in follicles	4 hrs.	----
37				Necrotic foci in germinal follicles	8-12 hrs	----

/1/ Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.  
 /2/ Not recovering at time last sample studied.

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Continued)

TISUES (Continued)							
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)
			Mouse (Continued)				
38	X-ray (185 KV)	400 r, Acute, total body	Lymph nodes	Decreased cellularity	2-3 hrs.	4 days	
39				Increased debris	2-3 hrs	----	
40				Cytological damage	2-3 hrs.	4 days	
41				Disappearance of secondary nodules	48 hrs.	7-14 days	
42	Fast neutrons (pile)	98 n, Acute, total body	Lymph nodes	Increased extramedullary myelopoiesis	3-4 weeks	----	2
43				Complete destruction of nodules	1/2 hr.	3-9 days	
44	X-ray (185 KV)	400 r, Acute, total body	Spleen	Decreased mitosis	2-3 hrs	7-10 days	
45				Cytological damage, lymphocytes	2-3 hrs.	7-10 days	
46				Increased debris	2-3 hrs	----	
47				Decreased erythropoiesis	2-3 hrs.	7-10 days	
48				Decreased megakaryocytes	2-3 hrs	10-14 days	
49				Increased extramedullary myelopoiesis	10 days	----	2
50	X-ray (200 KVP)	400 r, Acute, total body	Bone marrow	Decreased mitosis	Almost immediately after exposure	3 hrs.	
51				Abnormal mitosis	8 hrs.	14 hrs	
52				Decreased mitosis	3 hrs	----	
53				Decreased erythropoiesis	3 hrs	14 days	
54				Decreased granulocytopoiesis	1 day	before 14 days	
55				Increased debris	3 hrs	----	
56				Cessation of mitosis	1/2 hr.	10 days	
57				Decreased erythropoiesis	1/2 hr	10-14 days	
58				Increased debris	3 hrs	----	
59				Decreased myelopoiesis	1 day	14 days	
				800 r, Acute total body	Bone marrow		
Rabbit							
51	X-ray (200 KVP)	400 r, Acute, total body	Bone marrow	Decreased mitosis	Almost immediately after exposure	3 hrs.	
52				Abnormal mitosis	8 hrs.	14 hrs	
53				Decreased mitosis	3 hrs	----	
54				Decreased erythro-	3 hrs	14 days	
55				Decreased poiesis	3 hrs	before 14 days	
56				Decreased granulo-	1 day	----	
57				Increased debris	3 hrs	10 days	
58				Cessation of mitosis	1/2 hr.	10-14 days	
59				Decreased erythro-	1/2 hr	----	
				Increased debris	3 hrs	14 days	
	800 r, Acute total body	Bone marrow					

1/ Time represents an approximation. The figures given have been determined, in large part, by c-

2/ Not recovering at time last sample studied

<sup>1/1</sup> Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling  
<sup>2/2</sup> Not recovering at time last sample studied

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Continued)

Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>
(A)	(B)	(C)	(D)	(E)	(F)
Rabbit (Cont'd)					
60	X-ray (200 KVP)	800 r, Acute, total body	Bone marrow	Decreased. megalo-blasts	2 days
61		50 r, Acute, total body	Lymph nodes	Increased. debris	3 hrs.
62		100 r, Acute, total body	Lymph nodes	Increased. debris	3 hrs.
63	X-ray (200 KVP)	400 r, Acute, total body	Lymph nodes	Increased: debris	within 3 hrs.
64				Decreased. cellu-larity of nodules	3 hrs.
65				Infiltration with heterophils	1st day
66				Decreased: mitosis	3 hrs.
67				Shift to left in lymphocytes	8-16 hrs.
68	X-ray (200 KVP)	800 r, Acute, total body	Lymph nodes	Decreased: mitosis	30 min.
69				Destruction of lym-phocytes	30 min.
70				Disappearance of nodules	17-24 hrs
71				Increased debris	30 min.
72				Increased spindle cells	24 hrs.
73				Increased plasma cells	9 days
74	X-ray (200 KVP)	25 r, Acute, total body	Spleen	Increased debris	1 day
75				Cytological damage	1 day
76	X-ray (200 KVP)	50 r, Acute, total body	Spleen	Decreased mitosis	1 day
77				Cytological damage	1 day
78	X-ray (200 KVP)	600 r, Acute, total body	Spleen	Cessation of mitosis	1 hr.
79				Increased debris	1 hr.
80				Cytological damage	1 hr.
81				Decreased cellu-larity	1 hr

<sup>1</sup>/ Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Continued)

RADIATION ON HEMATOPOIETIC TISSUES (Continued)						
Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>	
(A)	(B)	(C)	(D)	(E)	(F)	
Rabbit (Cont'd)						
82	X-ray (200 KVP)	800 r, Acute, total body	Spleen	Decreased size	1 day	----
83				Cessation of mitosis	1/2 hr.	8 hrs.
84				Cytological damage (lymphocytes)	1/2 hr.	----
85				Increased debris	1/2 hr.	8 hrs.
86				Destruction of lymphocytes	3 hrs.	8 days
87				Decreased cellularity	3 hrs.	9 days
88				Increased plasma cells	9 days	----
89				Decreased erythroblasts	24 hrs.	14 da-
90				Decreased myelocytes	2 days	14 days
Rat						
91	X-ray (200 KVP)	400 r, Acute, total body	Lymph nodes	Cytological damage (lymphocytes)	within 16 hrs.	3 days
92				Degeneration of nodule	within 16 hrs	3 days
93	X-ray (200 KVP)	600 r, Acute, total body	Lymph nodes	Cytological damage (lymphocytes)	1/2 hr.	14-1/2 hrs.
94				Degenerative change (nodules)	14-1/2 hrs	21 days
95				Disappearance of nodules	24 hrs.	21 days
96				Infiltration with heterophils	8 hrs	----
97	X-ray (250 KVP)	550 r, Acute, total body	Spleen	Hemorrhage	all periods up to 31 days	---- <sup>2</sup>
98				Increased debris	30 min.	8 hrs.
99				Destruction of lymphocytes	2 hrs.	----
100				Disappearance of nodules	30 hrs.	15 days
101				Decreased cellularity	2 hrs	9 days

1/ Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.

2/ Not recovering at time last sample studied

<sup>1/</sup> Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.  
<sup>2/</sup> Not recovering at time last sample studied

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Continued)

Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>
(A)	(B)	(C)	(D)	(E)	(F)
Rat (Continued)					
102	X-ray (200 KVP)	Spleen	Decreased megakaryocytes	3 days	3 weeks
103			Cytological damage (erythroblasts)	3 hrs.	----
104			Decreased erythropoiesis	8 hrs.	9 days
105			Destruction of lymphocytes	3 hrs.	----
106			Decreased mitosis	1/2 hr	8 hrs.
107			Decreased cellularity of nodules	3 hrs.	14 days
108	X-ray (200 KVP)	Marrow	Decreased lymphocytes	15 min.	12 days
109			Decreased megakaryocytes	3 hours	14 days
110			Decreased myeloid cells	30-45 hrs.	14 days
111			Decreased erythroid cells	6-15 hrs.	5 days
112	X-ray (200 KVP)	Marrow	Increased debris	3 hrs.	----
113			Decreased erythropoiesis	3 hrs.	9 days
114			Decreased myelopoiesis	2 days	31 days
115	Fast neutrons (cyclotron)		Decreased megakaryocytes, granulocytes, erythroid cells, cellularity, mitosis	----	16 days
Chick					
116	X-ray (200 KVP)	Bone marrow	Cytological damage (lymphocytes)	----	----
117		Bone marrow	Decreased mitosis, cellularity	30 min 13 hrs	2 hrs. 12 days
118		Bone marrow	Decreased lymphocytes	7 hrs.	13-48 hrs.
119			Decreased myelocytes	7 hrs.	13-48 hrs.
120			Decreased erythroblasts	7 hrs.	13-48 hrs.

<sup>1</sup>/ Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.

# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES (Concluded)

	Radiation	Dose and Application	Tissue	Main Histological Effect	Time Post-Exposure When First Observed <sup>1</sup>	Time Post-Exposure When Definite Recovery Began <sup>1</sup>
	(A)	(B)	(C)	(D)	(E)	(F)
	Chick (Continued)					
121	X-ray (200 KVP) (Continued)	800 r, Acute, total body	Bone marrow	Decreased, mitosis	45 min.	---- <sup>2</sup>
122				Erythropoiesis abolished	1 hr.	2-4 days
123				Erythroblasts destroyed	1 hr.	2-4 days
124				Small lymphocytes destroyed	1-2 hrs.	---- <sup>2</sup>
125				Increased debris	1-2 hrs	----
126				Granulocytopoiesis destroyed	14 hrs.	---- <sup>2</sup>
127	X-ray (200 KVP)	25 r, Acute, total body	Spleen	Cytological damage (lymphocytes)	----	----
128		100 r, Acute, total body	Spleen	Debris and dead lymphocytes	1/2 hr.	18 hrs.
129		400 r, Acute, total body	Spleen	Decreased mitotic activity	1/2 hr	7 hrs.
130				Destruction of small lymphocytes	1/2 hr.	3 days
131				3/4 of small lymphocytes killed	4 hrs	3-5 days
132		800 r, Acute, total body	Spleen	Cessation of mitosis	45 min.	5 hrs.
133				Destruction of lymphocytes	45 min	9 days
134				Increased hemocytoblasts	45 min.	1-3 days
135	Increased plasma cells			2 hrs	----	
	Salmon					
136	X-ray (200 KVP)	500 r, Acute, total body	Kidney	Decreased hematopoietic cells in hematopoietic tissue	----	----

1/1 Time represents an approximation. The figures given have been determined, in large part, by frequency of sampling.  
 1/2 Not recovering at time last sample studied.



# 100. CHANGES IN STORED PRESERVED BLOOD

Variable	Preservative <sup>2</sup>	Period of Storage															
		0 days			10 days			20 days			30 days			40 days			
		Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)	
1																	
2	pH <sup>3</sup>	C	7.4			7.1			7.1						6.7		
3		CD	7.4			7.1			6.8						6.6		
4		ACD	7.1			6.7			6.6						6.5		
5	Potassium	C		100	5		75	23		63	32						
6	(mEq/1000 ml)	CD		100	5		74	20		63	25		60	28		53	34
7		ACD		90	5		75	12		68	19		80	23		53	27
8	Sodium	C		25	166		45	149		55	142						
9	(mEq/1000 ml)	CD		21	158		40	148		47	145		52	142		58	138
10		ACD		18	160		25	152		30	148		37	142		42	137
11	Inorganic Phosphorus	C		0	0		35	7									
12	(mg/100 ml)	CD		0	2		10	3		28	6		29	10		26	12
13		ACD		5	2		18	5		25	8		25	9		25	10
14	Adenosine Tri-phosphate	C		15			5			2						2	
15	(mg/100 ml)	CD		15			9			5			2			6	
16		ACD		15			14			12			9				
17	Lactic acid <sup>4</sup>	C	20			80		80									
18	(mg/100 ml)	CD	10		135			185				210					
19		ACD	20		90			130				155			170		
20	Glucose <sup>5</sup>	C	70		106												
21	(mg/100 ml)	CD	7287		6509				106			106					
22		ACD	6708		6007				5509			470					

/1/ Whole preserved (C, CD, and ACD) human blood kept at 4°C and analyzed within 20 minutes after removal from storage. /2/ Preservative C; 5 g trisodium citrate-di-H<sub>2</sub>O, in 100 ml aqueous solution, pH 7.5 (effective tonicity re blood = 141%). Ten ml added to 100 ml blood, increasing blood citrate by 10 mM/liter, and plasma citrate by 17 mM/liter. Preservative CD; 2.13 g trisodium citrate-di-H<sub>2</sub>O, plus 5 g glucose anhydrous, in 100 ml aqueous solution, pH 7.5 (effective tonicity re blood = 94%). Fifteen ml added to 100 ml blood, increasing blood citrate by 11 mM/liter, and plasma citrate by 19 mM/liter, blood glucose by 650 mg/100 ml. Preservative ACD; 1.33 g trisodium citrate-di-H<sub>2</sub>O, plus 470 mg citric acid, plus 3 g glucose anhydrous, in 100 ml aqueous solution, pH 5.03 (effective tonicity re blood = 66%). Twenty-five ml added to 100 ml blood, increasing blood citrate by 13 mM/liter, plasma citrate by 21 mM/liter, blood glucose 600 mg/100 ml, citric acid by 14.5 mEq/liter. Blood changes in 8 days with C equivalent to 18 days with CD equivalent to 33 days with ACD. /3/ pH measured at 37.5°C. Blood and plasma, same pH. /4/ Determined by modified aeration-titration method of Friedman. /5/ Determined by Nelson modification of Somogyi method. /6/ Residual non-fermentable reducing substances. /7/ Normal blood glucose plus glucose in CD. /8/ Normal blood glucose plus glucose in ACD.

# 100. CHANGES IN STORED PRESERVED BLOOD (Concluded)

Variable	Preservative	Period of Storage															
		0 days				10 days				20 days				30 days			
		Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	Blood	RBC	Plasma	
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)	(J)	(K)	(L)	(M)	(N)	(O)	(P)	(Q)	
22	(mg glucose/100 ml RBC/hr)	C	50			10.10				110							
23	(mg lactic acid/100 ml RBC/hr)	CD	40.11														
24	(mg lactic acid/100 ml RBC/hr)	ACD	17.11														
25	(mg lactic acid/100 ml RBC/hr)	C	7.69			20.11				6.11							
26	(mg lactic acid/100 ml RBC/hr)	CD	1.80			17.11				12.11							
27	(mg lactic acid/100 ml RBC/hr)	ACD	2.32			6.92				6.25							
28	(mg lactic acid/100 ml RBC/hr)	C	7.50			2.32				3.05							
29	(mg lactic acid/100 ml RBC/hr)	CD	2.08			7.33											
30	(mg lactic acid/100 ml RBC/hr)	ACD	2.33			2.30											
31	(mg lactic acid/100 ml RBC/hr)	C	7.42			2.38				6.92							
32	(mg lactic acid/100 ml RBC/hr)	CD	2.33			7.17				2.52							
33	(mg lactic acid/100 ml RBC/hr)	C	0			9.8											
34	(mg lactic acid/100 ml RBC/hr)	CD	0			30											
35	(mg lactic acid/100 ml RBC/hr)	ACD	0			61											
36	(mg lactic acid/100 ml RBC/hr)	C	0			58											
37	(mg lactic acid/100 ml RBC/hr)	CD	0			60											
38	(mg lactic acid/100 ml RBC/hr)	C	0			0											
39	(mg lactic acid/100 ml RBC/hr)	CD	0			2											
40	(mg lactic acid/100 ml RBC/hr)	ACD	0			3.6											
41	(mg lactic acid/100 ml RBC/hr)	C	0			16											
42	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
43	(mg lactic acid/100 ml RBC/hr)	C	0			0											
44	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
45	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
46	(mg lactic acid/100 ml RBC/hr)	C	0			4.2											
47	(mg lactic acid/100 ml RBC/hr)	CD	0			12.1											
48	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
49	(mg lactic acid/100 ml RBC/hr)	C	0			0											
50	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
51	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
52	(mg lactic acid/100 ml RBC/hr)	C	0			0											
53	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
54	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
55	(mg lactic acid/100 ml RBC/hr)	C	0			0											
56	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
57	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
58	(mg lactic acid/100 ml RBC/hr)	C	0			0											
59	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
60	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
61	(mg lactic acid/100 ml RBC/hr)	C	0			0											
62	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
63	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
64	(mg lactic acid/100 ml RBC/hr)	C	0			0											
65	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
66	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
67	(mg lactic acid/100 ml RBC/hr)	C	0			0											
68	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
69	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
70	(mg lactic acid/100 ml RBC/hr)	C	0			0											
71	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
72	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
73	(mg lactic acid/100 ml RBC/hr)	C	0			0											
74	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
75	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
76	(mg lactic acid/100 ml RBC/hr)	C	0			0											
77	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
78	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
79	(mg lactic acid/100 ml RBC/hr)	C	0			0											
80	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
81	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
82	(mg lactic acid/100 ml RBC/hr)	C	0			0											
83	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
84	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
85	(mg lactic acid/100 ml RBC/hr)	C	0			0											
86	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
87	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
88	(mg lactic acid/100 ml RBC/hr)	C	0			0											
89	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
90	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
91	(mg lactic acid/100 ml RBC/hr)	C	0			0											
92	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
93	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
94	(mg lactic acid/100 ml RBC/hr)	C	0			0											
95	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
96	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
97	(mg lactic acid/100 ml RBC/hr)	C	0			0											
98	(mg lactic acid/100 ml RBC/hr)	CD	0			0											
99	(mg lactic acid/100 ml RBC/hr)	ACD	0			0											
100	(mg lactic acid/100 ml RBC/hr)	C	0			0											

10/ Glycolysis measured by breakdown of glucose. 11/ Glycolysis and glucose lost. 12/ Glycolysis involves breakdown of added glucose. 13/ Determined colorimetrically. 14/ Determined colorimetrically. 15/ Determined colorimetrically. 16/ Determined colorimetrically. 17/ Determined colorimetrically. 18/ Determined colorimetrically. 19/ Determined colorimetrically. 20/ Determined colorimetrically. 21/ Determined colorimetrically. 22/ Determined colorimetrically. 23/ Determined colorimetrically. 24/ Determined colorimetrically. 25/ Determined colorimetrically. 26/ Determined colorimetrically. 27/ Determined colorimetrically. 28/ Determined colorimetrically. 29/ Determined colorimetrically. 30/ Determined colorimetrically. 31/ Determined colorimetrically. 32/ Determined colorimetrically. 33/ Determined colorimetrically. 34/ Determined colorimetrically. 35/ Determined colorimetrically. 36/ Determined colorimetrically. 37/ Determined colorimetrically. 38/ Determined colorimetrically. 39/ Determined colorimetrically. 40/ Determined colorimetrically. 41/ Determined colorimetrically. 42/ Determined colorimetrically. 43/ Determined colorimetrically. 44/ Determined colorimetrically. 45/ Determined colorimetrically. 46/ Determined colorimetrically. 47/ Determined colorimetrically. 48/ Determined colorimetrically. 49/ Determined colorimetrically. 50/ Determined colorimetrically. 51/ Determined colorimetrically. 52/ Determined colorimetrically. 53/ Determined colorimetrically. 54/ Determined colorimetrically. 55/ Determined colorimetrically. 56/ Determined colorimetrically. 57/ Determined colorimetrically. 58/ Determined colorimetrically. 59/ Determined colorimetrically. 60/ Determined colorimetrically. 61/ Determined colorimetrically. 62/ Determined colorimetrically. 63/ Determined colorimetrically. 64/ Determined colorimetrically. 65/ Determined colorimetrically. 66/ Determined colorimetrically. 67/ Determined colorimetrically. 68/ Determined colorimetrically. 69/ Determined colorimetrically. 70/ Determined colorimetrically. 71/ Determined colorimetrically. 72/ Determined colorimetrically. 73/ Determined colorimetrically. 74/ Determined colorimetrically. 75/ Determined colorimetrically. 76/ Determined colorimetrically. 77/ Determined colorimetrically. 78/ Determined colorimetrically. 79/ Determined colorimetrically. 80/ Determined colorimetrically. 81/ Determined colorimetrically. 82/ Determined colorimetrically. 83/ Determined colorimetrically. 84/ Determined colorimetrically. 85/ Determined colorimetrically. 86/ Determined colorimetrically. 87/ Determined colorimetrically. 88/ Determined colorimetrically. 89/ Determined colorimetrically. 90/ Determined colorimetrically. 91/ Determined colorimetrically. 92/ Determined colorimetrically. 93/ Determined colorimetrically. 94/ Determined colorimetrically. 95/ Determined colorimetrically. 96/ Determined colorimetrically. 97/ Determined colorimetrically. 98/ Determined colorimetrically. 99/ Determined colorimetrically. 100/ Determined colorimetrically.

1. Blood incubated 90 minutes at 37°C. measured lactic acid produced and glucose lost. 10/ Glycolysis measured by breakdown of added glucose as glucose originally present has disappeared. see Fn 5. 11/ Glycolysis involves breakdown of added glucose. see Fn 7, 8. 12/ Mean Corpuscular Volume (M.C.V.) = Hematocrit/RBC count per mm<sup>3</sup> blood. Hemocrit determined by method of Guest and Slier. Mean diameter read from stained film covered with 0.1% NaCl. Mean Thickness = V.C.V./π (Diam./2)<sup>2</sup>. 13/ Determined colorimetrically 14/ 1% sodium citrate.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
<b>I. Aliphatic Narcotics</b>								
1 Amytal, pentobarbital and secobarbital	Sedation and hypnosis	100-250 mg	Oral	1 hr	<1			
2 Barbitol	Sedation and hypnosis	16-20 mg/kg/day	Oral	1-24 hr	5			
3 Chloroform	Surgical anesthesia	1.5 vol. % in inspired air	Inhalation	About 5 min	12 <sup>1</sup>			
4 Cyclopropane	Surgical anesthesia	20-30 vol. % in inspired air	Inhalation	2-3 min	15-20			
5 Ethyl alcohol	Euphoria	About 30 ml 50% alcohol	Oral	15-30 min	30			
6	Hypnotic	About 100 ml, 50% alcohol	Oral	15-30 min	100			
7	Anesthetic (stupor)	About 300-400 ml 50% alcohol	Oral	15-30 min	300-400			
8	Generally fatal	Adults: 1000 ml, Children: 100-200 ml	Oral	5-100 hr	500			
9 Ethyl chloride	Surgical anesthesia	50% alcohol 3-4.5 vol. % in inspired air	Inhalation	2 min	20-30			
10 Ethylene	1st plane anesthesia	80-85 vol. % in inspired air	Inhalation	20 min	120-180			
11 Ethyl ether	Surgical anesthesia	6-10 vol. % in inspired air	Inhalation	15-25 min	50-130			
12 Methyl N-propyl ether (Metopryl)	Surgical anesthesia	2.5-3 vol. % in inspired air	Inhalation	10-15 min	30-60			
13 Nitrous oxide (Nitrogen monoxide)	1st plane anesthesia	85-92 vol. % in inspired air	Inhalation	2 min	23 vol. %			
14 Paraldehyde	Obstetrical amnesia and analgesia	30 ml in 60 ml olive oil	Rectal or nasal	0.5-16 hr <sup>2</sup>				
15 Phenobarbital	Sedation and hypnosis	5-10 mg/kg/day	Oral	•1-24 hr	5 or less <sup>3</sup>	11.8-24.4		
16 Thiopental sodium (Sodium pentothal)	1st & 2nd plane anesthesia	35 mg/kg/hr	Intravenous	Immediate				
17 Tribromethanol (Avertin)	Basal anesthesia	80-100 mg/kg	Rectal	30 min	6-9			
18 Vinyl Ether (Divinyl oxide, "Vinethene")	Surgical anesthesia	4 vol. % in inspired air	Inhalation	1-2 min	30-40			
<b>II. Bromides</b>								
19 Sodium bromide or other alkaline bromides	Sedation	2-3 g/day	Oral	1 hr or more	16-50			
20	Antiepileptic	3-10 g/day	Oral	Several days				
<b>III. Analgesics</b>								
21 Isonipacaine (Demerol)	Analgesic	100 mg	Intramuscular	1.5-2 hr	0.125 in 0.5 hr and 0.05 after 2 hr			

1/ May produce unconsciousness 2/ Peak level 2-4 hr. 3/ Blood level over 7 mg/100 ml indicates overdosage.  
 4/ Bromoderma and other symptoms observed.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS (Continued)

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
22 Sodium salicylate	Acute rheumatic fever	5-15 g/day	Oral	24 hr or more		8-28	157-558 µg/ml	
23 <u>IV. Cardiac Drugs</u> Digloxin	Congestive heart failure	1.2-1.6 mg	Oral	About 6 hr			20 µg/100 ml <sup>5</sup>	
24 Ouabain	Congestive heart failure	0.5 mg (divided)	Intra-venous	About 1-2 hr			5 µg/100 ml <sup>5</sup>	
25 Potassium acetate	In relation to digitalis glycosides	5 g	Oral	1 hr or more			20-27	
26 Procaine hydrochloride	Abolition of cardiac arrhythmias	200 mg	Intra-venous	25 min	0.055-0.080			
27 Quinidine sulfate	Control of various cardiac arrhythmias	0.8 g	Oral	2-10 hr		0.08-0.27 <sup>6</sup>		
28 <u>V. Hypotensive Agents</u> Potassium thiocyanate	Hypertension	0.5 g/day	Oral	1-3 hr		8-10		
29 <u>VI. Skeletal Muscle Relaxants</u> Myanesin or 3-(2'-Methylphenoxy)propane-1,2-diol (spinal cord depressant)	Adjuvant to anesthetics, treatment of convulsive poisonings (particularly strychnine)	50 mg	Intra-venous	Immediate		5		
30				20 min later		2.4		
31 d-Tubocurarine chloride (neuro-effector depressant)	Adjuvant to anesthetics, control of shock therapy in schizophrenia	0.2 mg/kg	Intra-venous	3 min		0.4		
32				15 min		0.26		
33				30 min		0.1		
34 <u>VII. Histamine and Antihistaminics</u> beta-Dimethylaminoethyl benzhydriol ether hydrochloride (Benadryl)	Allergic reactions	400 mg	Oral	1-2 hr	0.5-1.8 µg/ml			
35 Histamine phosphate	Stimulation of hydrochloric acid secretion	2 mg	Subcutaneous	30 min	>7.5 µg/100 ml			
36 Pyridyl-benzyl-dimethyl-ethylene diamine hydrochloride (Tripellenamide or pyribenzamine)	Allergic reactions	400 mg	Oral	2-3 hr	0.4 µg/ml			
37 <u>VIII. Anti-anemics</u> Ferric sodium citrate	Secondary anemia	0.75 g <sup>7</sup>	Oral	1 hr			0.15	
38				3 hr			0.25	
39				8 hr			18	
40				14 hr			<0.8	

<sup>5</sup>/Calculated, /6/1-4 hr 1 g oral 0.2 - 0.43 mg/100 ml plasma /7/insufficient therapeutic dosage.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS (Continued)

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
41 Ferrous ammonium sulfate	Secondary anemia	0.75 g <sup>7</sup>	Oral	Same as (37-40E)			Similar to (37-40 H)	
IX. Iodine and Anti-thyroid Substances								
42 Sodium iodide	Prevention of hypothyroidism	1.5 mg/day	Oral	2-4 wk	7 µg/100 ml			
43	Pre-operative treatment of thyrotoxicosis	15 mg/day	Oral	Several days	>19 µg/100 ml <sup>8</sup>			
44 Thiourea	Thyrotoxicosis	1-2 g	Oral	Variable	3-5			
45 2-Thiouracil	Thyrotoxicosis	0.8 g/day	Oral	Variable		0.3-0.5		
X. Calcium Compounds								
46 Di-calcium phosphate	Calcium deficiency	0.8 g t.i.d. (with vitamin D)	Oral	Several days	8-11			
XI. Chemotherapeutic Agents								
(I) Antibiotics								
47 Aureomycin	Typhoid fever	100 mg	Intramuscular	24 hr	0.03-0.06 µg/ml			
48	Rocky Mountain spotted fever	300 mg	Oral	24 hr	0.03 µg/ml			
49	A. aerogenes, abscess pneumonia, and meningitis	700 mg	Oral	24 hr	0.03-0.1 µg/ml			
50 Chloramphenicol (Chloromycetin)	Tularemia and other conditions	15.5 g in four days	Oral	24 hr or more	20-30 µg/ml			
51 Penicillin	Infection with penicillin-susceptible organisms	100,000 units in 24 hr	Intravenous drip			0.13 units/ml		
52		200,000 units in 24 hr				0.22 units/ml		
53		400,000 units in 24 hr				0.42 units/ml		
54 Penicillin	Respiratory infections	30,000 units every 2-4 hr	Inhalation <sup>9</sup>	2-3 hr			0.11-0.22 units/ml	
55 Penicillin-aluminum hydroxide	Mild infections	100,000 units	Oral	1-10 hr		0.03-0.3 units/ml		
56 Penicillin (Beeswax-peanut oil mixture)	Usual indications	800,000 units	Intramuscular	12 hr		0.1-1.0 units/ml		
57 Penicillin (Crystalline) without caronamide	Severe infections	1,000,000 units	Rapid Intravenous	3 hr		2 units/ml		
58 Penicillin (Crystalline) with caronamide	Severe infections	1,000,000 units	Rapid intravenous	3 hr		10 units/ml		
59 Penicillin (Procaine penicillin G in oil)	Pneumonia	300,000 units	Intramuscular	8 hr		0.34-0.66 units/ml		
60 Streptomycin	Infections with streptomycin-susceptible bacteria	0.15 g every 3 hr	Intramuscular	3 hr	14 µg/ml <sup>10</sup>			

<sup>7</sup>/Insufficient therapeutic dosage. <sup>8</sup>/Calculated. <sup>9</sup>/Through nebulizers. <sup>10</sup>/16-20 µg/ml is usually more desirable.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS (Continued)

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
81 Terramycin hydrochloride (3) Anti-Malarials	Acute pulmonary infections	750 mg or more	Oral	1-5 hr	0.7-5.0 µg/ml			
82 Chloroquine hydrochloride (Paludrine)	Suppression and termination of attacks during vivax or falciparum infection	50 mg b.i.d. 500 mg b.i.d.	Oral	4 hr		50-100 µg/1000 ml 300-500 µg/1000 ml		
84 Chloroquine diphosphate	Suppression of malaria in man	1.24 g (divided within a day)	Oral	4 hr		80-160 µg/1000 ml		
85		Priming dose 1.2 g (within 6 hr)		6-8 hr		280 µg/1000 ml		
86 Pamaquine naphthoate	Gametocidal effect of malaria organisms in man	0.01 g 5 times/day	Oral	1-2 hr		250 µg/1000 ml		
87 Pentaquine monophosphate	Gametocidal effect	60 mg/day	Oral	1 hr		30-50 µg/1000 ml		
88 Quinacrine hydrochloride (Atabrine)	Suppression of malaria in man	0.6 g/wk (divided)	Oral	1 wk			30-50 µg/1000 ml	
89	Suppression of malaria in man	1.6 (immediately)	Oral	48 hr		81-245 µg/1000 ml <sup>11</sup>		
90 Quinacrine	Severe malaria (ante mortem) or in gastrointestinal intolerance to quinacrine	1 g	Intravenous	Immediate		8-22		
91 Quinine salts	Human malaria, Class I (Decelerated increase in the parasitemia)	0.5 g once daily	Oral			<0.2		
92	Class II (Marked depression of parasitemia with clinical remission)	1-2 g/day	Oral			0.2-0.4		
93	Class III (Complete disappearance of parasitemia, no clinical symptoms)	3 g/day	Oral			0.5-0.7		
(3) Antimony Compounds								
94 Antimonialine (Antimony lithium thiomaleate)	Wuchereria bancrofti infection and kala azar	0.5 mg of Sb/kg	Intra-muscular	1 hr		0.18 µg/g		0.78 µg/g
95		0.5 mg of Sb/kg	Intra-muscular	24 hr		0.03 µg/g		0.15 µg/g
96 Monosodium antimony thio-glycollate	Same as 94B	0.5 mg of Sb/kg	Intra-muscular	1 hr		0.12 µg/g		1.1 µg/g
97 Neostibosan (Ethylstibamine)	Same as 94B	2.0 mg of Sb/kg (initial)	Intra-venous	24 hr		0.07 µg/g		0.35 µg/g
				24 hr		0.7-0.9 µg/g		0.2-0.3 µg/g

<sup>111</sup>/Each gram of quinacrine administered by mouth raises the plasma level by 25 µg/1000 ml, and there is a daily decrease of the plasma concentration by 10 per cent. <sup>112</sup>/Total dose: 58-66 mg Sb/kg in 8-10 days.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS (Continued)

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
79 Solustibosan (Diethylamino-ethanol salt of sodium antimony glyconate) (4) Aromatic Amidines	Same as 74B	3.0 mg of Sb/kg (initial) <sup>13</sup>	Intra-venous	24 hr		0.4 µg/g		0.6 µg/g
80 Stilbamidine	Trypanosomiasis, Kala azar, babesiasis, and multiple myeloma		Oral	30 min			40 µg/ml	
81			Subcutaneous	1 hr			20 µg/ml	
(5) Arsenicals								
82 Arspenamine	Syphilis (Intensive therapy)	31.7 mg/kg in 5 days t.i.d.	Intra-venous	4-6 hr	0.5-0.7 mg/100 g			
83 Clorarsen (Dichlorophenarsine hydrochloride)	Syphilis	0.067 g/day	Intra-venous	15 min		< 0.05 <sup>14</sup>		
84 Neoarsphenamine	Syphilis	0.1 g	Intra-venous	15 min		0.2-0.38 <sup>14</sup>		
85 Oxophenarsine hydrochloride (Mapharsen)	Syphilis	0.06 g	Intra-venous	10 min		< 0.05 <sup>15</sup>		
86	Syphilis (Intensive therapy)	0.84 g/day	Intra-venous	5 days	17 µg/100 ml <sup>14</sup>			
87 Tryparsamide	Neurosyphilis, trypanosoma gambiense infection	2 g	Intra-venous	24 hr		0.03-0.04 µg/ml <sup>14</sup>		
(6) Sulfonamides								
88 Sulfadiazine	Same as the other sulfonamides, particularly effective against type B Friedlaender bacillus	4 g initially followed by 1 g every 4 hr	Oral	6-12 hr	10-15			
89 Sulfaguanidine	Prophylaxis in colonic surgery, intestinal antiseptics	0.01 g/kg initially, followed by 0.05 g/kg every 4 hr	Oral	Irregular	5			
90 Sulfamerazine	Same as other sulfonamides	3-4 g initially, followed by 1 g every 8 hr	Oral	12-24 hr	10-15			
91 Sulfanilamide	Streptococcal, meningococcal, pneumococcal, staphylococcal infections, etc.	30-50 mg/kg in 24 hr <sup>15</sup>	Oral	12-24 hr	10-15			
92 Sulfapyridine	Same as 91 B (seldom used)	5 g	Oral	6-12 hr	4-6			
93 Sulfathiazole	Pneumococcal and gonococcal infections	4 g initially, followed by 0.5-1 g every 4 hr	Oral	4-8 hr	4-6			

/13/ Total dose 71-84mg Sb/kg in 8 or 9 days. /14/ Expressed as arsenic /15/ One-third of total dose should be given first day

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS (Concluded)

Therapeutic Agent	Therapeutic Use	Dosage	Method of Admin.	Time after Admin.	Blood	Plasma	Serum	RBC
					mg/100ml	mg/100ml	mg/100ml	mg/100ml
(A)	(B)	(C)	(D)	(E)	(F)	(G)	(H)	(I)
<b>XII. Miscellaneous Chemotherapeutic Agents</b>								
94 Para-aminobenzoic acid	Typhus, dengue, etc.	8 g initially followed by 3 g every 2 hr	Oral	12-24 hr	100-130			
95 p-Chloroxylenol	Urogenital infections	1-2 g	Intra-muscular	2 hr	1-3			
98 Suramin sodium (Germanin, Bayer 203)	African sleeping sickness	1 g	Intra-venous	48 hr		3		
97				9 days		0.8		
<b>XIII. Vitamins</b>								
96 Vitamin A	Hypovitaminosis A	10,000 or more units/day	Oral			80-130 I.U./100 ml		
99 Ascorbic acid	Scurvy; peptic ulcer	5 g/day	Oral	Variable		0.6-1.8		
100 Biotin			Oral		1.3-1.8 µg/100ml			
101 Carotene	Hypovitaminosis A ( seldom used)	0.3 mg/day	Oral			100-220 µg/100ml		
102 Folic acid	Nontropical sprue	10-20 mg/day	Oral		3.5 µg/100 ml			
103 Inositol	Adipotropic lateral sclerosis	1.5 g/day	Oral	3 wk		0.75		
104 Nicotinic acid	Clinical pellagra	Up to 500 mg/day	Oral		0.8 µg/100 ml			
105 Pantothenic acid			Oral		30-45 µg/100 ml			
106 Riboflavin	Cheilosis, etc.	5-10 mg/day	Oral		80			
107 Tocopherol	Habitual abortion, menstrual disorders, etc.	0.25-8 ml of wheat germ oil/day	Oral	Variable		1.2 or more		
<b>XIV. Hormones</b>								
108 Estrone sulfate (Conjugated estrogens)	Cancer of prostate, etc.	100-400 mg <sup>16</sup>	Intra-venous	Immedi-ately			18-48 µg/ml	
109				2 hr			2-18.4 µg/ml	
110				4 hr			0-18 µg/ml	
111				8 hr			0-7.8 µg/ml	
112				24 hr			0-9 µg/ml	
113 ACTH	Acute rheumatic fever, rheumatoid arthritis	100-330 mg <sup>16</sup>	Intra-venous	2 hr		10-100 µg/ml		
114 Thyroxin	Myxedema	0.3 mg	Oral	Variable	12-13 µg/100 ml of hormonal iodine			
<b>XV. Antimitotic Agents</b>								
115 Urethane	Chronic myeloid leukemia and Hodgkin's disease	1 g 6 times/day	Oral	22 hr	10-38			

<sup>16</sup>/Experimental human medication





## BIBLIOGRAPHY



# Bibliography

The system for presenting bibliographic references used here has been adapted from one in common use in geographic atlases. An item in a table is identified by two coordinates, a letter designating the column, and a number the row, in which the item appears. In a typical instance in the bibliography, the coordinates of an item are followed, in the column to the right, by a number accompanied by a small letter. The number refers to the bibliographic source, the letter to the person contributing the item. If an item in a table is based on another item from another part of the same table, the coordinates of the latter are given as source, enclosed in parentheses.

This method of presentation of bibliographic references permits identification of both the reference source of each item and the person contributing it, without the necessity of use of reference numbers on the face of the table. The resulting saving in space permits use of a larger type face in each table, with a consequent gain in readability. It is not uncommon for some value in a table to be based on more than a single reference source. The occurrence of such instances has given emphasis to the desirability of eliminating reference numbers from the tables.

Where an item is contributed as general knowledge, or from the unpublished investigations of the contributor, a letter appears in the source column without accompanying number.

It should be understood that a person listed as a contributor for any table is not responsible for portions of the table other than as indicated.

The following abbreviations are used: Fn = footnote; Calc. fr. = calculated from; Av. = average.

## 1. BLOOD SPECIFIC GRAVITY

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 2 1-3 ABCD & Fn 1 1-9 E 4, 5 A-E 6, 7 A-D 6, 7 E 8, 9 A-E* & Fn 3, 4, 5 10, 11 A-E	d, 10 7b 9b 5 8 12 1 5	12, 13 A-D* & Fn 6, 7 12, 13 E 14-19 ABCD* & Fn 8 14-19 E 20 A-D* & Fn 9 21 A-D* & Fn 10 21 E 22-27 ABCD* & Fn 11 22-27 E	3 12 13a 7 6c 2 11 13a 7	28 A-D* 28 E 29, 30 A-E 31 A-D* & Fn 12 31 E 32-36 ABCD* & Fn 13 32-36 E 37 A-E* & Fn 9, 14 38, 39 A-E	2 11 5 2 11 13a 7 c 5

\*Values in literature converted to standard reference temperature (as indicated in column D) by V P Dole

- Contributors (a) Cronkite, E P (b) Dole, V P (c) Holman, H H (d) Van Slyke, D D
- References (1) Braun, W. Am J Vet Res 145, 1946 (2) Dougherty, T F, and White, A Endocrinology 15, 1944 (3) Ederstrom, H E, and DeBoer, B, Anat Rec 94 663, 1946 (4) Holman, H H J Comp Path Therap 54 26, 1944 (5) MacLeod, J, Quart J Exp Physiol 22 275, 1932 (6) Stewart, J and Holman, H H, Vet Rec 52 157, 1940 (7) Van Slyke, D D, Phillips, R A, Dole, V P, Hamilton, P B, Archibald, R M., and Plazin, J J Biol Chem 183 349, 1950 (8) Windle, W F et al Anat Rec 78 321, 1940 (9) Phillips, R A, Van Slyke, D D, Hamilton, P B, Dole, V P, Emerson, K E, Jr., and Archibald, R M J Biol Chem 182 105, 1950 (10) Gregersen, M I, and Schiro, H, Am J Physiol 121 284, 1938 (11) Kagan, B M, J Lab Clin Med 26 168, 1941 (12) Barbour, H G, and Hamilton, W F J Biol Chem 63 625, 1926 (13) Cronkite, E P, unpublished data

## 2. RELATIVE VISCOSITIES, BLOOD, PLASMA, SERUM

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 1-3 1 A-C 1 D* E*	a 1a, 2a 3a, 4	5, 6 A-C 6 D* E*	7a 7a, 4	10 A-C 10 D* E*	7a 7a, 4
2 A-C 3 A-C 4 A-C	5a 5a 6a	7, 8 A-C 8 E*	7a 7a, 4	11 A-C E & Fn 4	8a
		9 A-C 9 E*	7a 7a, 4	12 A-C & Fn 4	9a

\* Calculated values  
 Contributors. (a) Pappenheimer, J.R. (b) Green, H.D. contributed to organization of table.

References (1) Hess, W., Arch. f. klin. Med. 94 404, 1908 (2) Determan, Z. f. klin. Med. 59 307, 1906 (3) Boile, B., "Beitrag zur Kenntnis der Viskosität des Blutes, des Serums und des Plasmas." Inaugural Dissertation. Gustav Schade, Berlin, 1909. (4) Handbook of Biological Data, Table on Plasma Proteins, Laboratory and Farm Animals (5) Hürthle, K., Pflügers Arch. 82:438, 1900. (6) Burton-Opitz, R., J. Physiol. 32:17, 1904. (7) Rhiel, Pflügers Arch. 246 709, 1943. (8) Tromsdorff, Arch. f. exp. Path. 45:82, 1900. (9) Burton-Opitz, R., Am. J. Physiol. 2 243, 1902.

## 3. BLOOD MISCELLANEOUS PHYSICAL PROPERTIES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-16 AB & Fn 1 17, 18 A-C 19-25 AB	1a 2 4	26-28 AB 29 A C & Fn 2	5 2	30-32 ABC & Fn 3 33-35 AB	5 3b

Contributors (a) Dole, V. P. (b) Mendlowitz, M.

References (1) Abramson, H. A., and Moyer, J. Gen. Physiol. 19 601, 1936 (2) Gram, H. C., Am. J. Med. Sci. 168:523, 1924. (3) Mendlowitz, M., Science 107 97, 1948. (4) Dukes, H. H., "The Physiology of Domestic Animals." Comstock Publishing Co., Ithaca, N. Y., 1937 (5) Keys, A., and Hill, R. M., J. Exp. Biol. 11 28, 1934

## 4. BLOOD COLLOIDS, OSMOTIC PRESSURE

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1, 2 A-D & Fn 1 3 A-D 4, 5 A-D 6-10 ABCD & Fn 3 11-21 ABCD	4b 3a, 6a 4b 2a, 5a, 7c 4b	22-24 ABCD 25 A-D 26 A-D 27 A-D 28-30 ABCD	2a, 5a 4b 2a, 5a 4b 2a, 5a	31 A-D 32, 33 A-D 34, 35 A-D	4a 2a, 5a 1a, 6a

Contributors (a) Hill, R. M. (b) Keys, A. (c) Walford, L. A. contributed common and accepted scientific names of elasmobranchs (6-10B) and of fish (22-33B)

References (1) Campbell, M. L., and Turner, A. H., Biol. Bull. 73 504, 1937 (2) Cardot, H., and Meyer, P., Ann. physiol. 12 665, 1936 (3) Horioka, Tôhoku J. Exp. Med. 19 233, 1932 (4) Keys, A., and Hill, R. M., J. Exp. Biol. 11 28, 1934 (5) Krogh, A., and Meyer, P., Ergeb. Physiol. 34 18, 1932 (6) Krogh, A., and Nakazawa, F., Biochem. Z. 188 241, 1927 (7) American Fisheries Society, "A List of Common and Scientific Names of the Better Known Fishes of the United States and Canada." Special Publication No. 1 Ann Arbor Michigan 1948

# 5. ERYTHROCYTE SEDIMENTATION RATE, TIME

Fal, 2	a	19 A CD	8	32 ABD & Fn 5	7
1.2 A CD	13ab	20 A-D & Fn 4	8	33 A-D	6
3.4 A CD	15a	21 A-D	10	34 ABD	10
5.6 A CD	9a	22.23 A-D	6	35.36 ABD	1
7.8 A CD	14a	24 A-D	6	37 A-D	6
9.10 A CD	11a	25 A-D	4	38 ABD	10
11.12 A CD	12a	26 ABD	10	39 A-D	6
13.14 A-D & Fn 3	9a	27 ABD	3	40 ABD	10
15 A-D	6	28 A-D	6	41 A-D	6
16 ABD	10	29 ABD	10	42 A CD & Fn 6	16a
17 A-D	6	30 A-D	6	43 ABD	2
18 ABD	10	31 ABD	10	44 A CD	5

Contributors: (a) Hirschboeck, J. S. (b) Riser, W. H., Jr.

References: (1) Creskoff, A. J., et al., p. 358 in "The Rat in Laboratory Investigation" Lippincott Co., Philadelphia, 1942. (2) Froila, G., Arch. biol. 5, 47, 1928. (3) Hammersland, H. L., et al., J. Am. Vet. Med. Assoc. 93 320, 1938. (4) Nicolle, P., and Simons, H., Sang 13, 401, 1919. (5) Rix, E., Z. ges. exp. Med. 99-178, 1936. (6) Schappes, H., Folia haematol. 59, 180, 1937. (7) Sharpe, J. C., and Bisgard, J. D., J. Lab. Clin. Med. 21, 347, 1936. (8) Simms, B. T., J. Am. Vet. Med. Assoc. 46, 77, 1940. (9) Smith, C. H., Am. J. Med. Sci. 162, 73, 1936. (10) Zott, F., Wien. tierarztl. Mschr. 18, 570, 1931. (11) Westergren, A., Am. Rev. Tuberc. 14, 94, 1926. (12) Wintrobe, M. M., and Landsberg, J. W., Am. J. Med. Sci. 189, 102, 1935. (13) Cutler, J. W., Am. J. Med. Sci. 183 251, 1936. (14) Walton, A. C. R., J. Lab. Clin. Med. 18, 711, 1933. (15) Landau, A., Am. J. Dis. Child. 45 691, 1933. (16) Linzenmeier, G., in Halban-Setz, "Biologie u. Pathologie des Weibes" 1927.

# 6. ERYTHROCYTE FRAGILITY MAN

Data C and i
1 A C-
2 A C-
3 A-H

Contributors: (a) Hendry, E. B. (b) Riser, W. H., Jr.

References: (1) Bohr, D. F., J. Lab. Clin. Med. 31, 1179, 1946. (2) Creed, E. ff., J. Path. Bact. 46 331, 1938. (3) Dacie, J. V., and Vaughan, J. M., J. Path. Bact. 46 341, 1938. (4) Daland, G. A., and Worthley, K., J. Lab. Clin. Med. 20 1122, 1935. (5) Hendry, E. B., Edin Med J 55, 427, 1948. (6) Hunter, F. T., J. Clin. Invest. 19 691, 1940. (7) Kato, K., J. Lab. Clin. Med. 26 703, 1941. (8) Olbrich, O., Edin Med. J 34 649, 1947. (9) Whithy, L. E. H., and Hynes, M., J. Path. Bact. 40, 219, 1935. (10) Wiczorowski, E., and Fishback, H. R., J. Lab. Clin. Med. 27 542, 1942.

# 7. ERYTHROCYTE FRAGILITY: VERTEBRATES OTHER THAN MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-C	11a	7 A-C	6a	14 A-C	1a
2 A-C	7a	8, 9 A-C	1a	15 A-C	8
3 A-C	3a	10 A-C	2a	16 A-C	9
4 A-C	4a	11 A-C	11a	17, 18, 19 A-C	1a
5 A-C	11a	12 A-C	6a	20 A-C	10
6 A-C	1a	13 A-C	11a	21, 22 A-C	5a

Contributors (a) Hendry, E. B.

References (1) Kato, K., J. Lab. Clin. Med. 26:703, 1941. (2) Ashby, W., Am. J. Physiol. 68 611, 1924. (3) Haeck, E., Wien. tierarztl. Mschr. 23:682, 1936. (4) Hammon, W.D., Anat. Rec. 76 259, 1940. (5) Hogan, A. G., and Parrott, E. M., J. Biol. Chem. 132:507, 1940. (6) Ponder, E., Yeager, J. F., and Charipper, H. A., Quart. J. Exp. Physiol. 19 115, 1928. (7) Ponder, E., Saslow, G., and Yeager, J. F., Biochem. J. 24:805, 1930. (8) Demmel in Dukes, H. H., "Physiology of the Domestic Animals," Comstock Publishing Co., Ithaca, N.Y., 1942. (9) Lyon, M. W., J. Infect. Dis. 22:49, 1918. (10) Hunter, F. T., J. Clin. Invest. 19 691, 1940. (11) Ponder, E., Yeager, J. F., and Charipper, H. A., Quart. J. Exp. Physiol. 19:181, 1928-29.

# 8. BLOOD COAGULATION. Theory of P. A. Owren (1952)

Footnotes	Contributors and References
Fn 1	1a, 2a
Fn 2	3a, 4a, 5a
Fn 3, 4, 5	6a, 7a
Fn 6	1a, 2a

Contributor Owren, P. A.

References (1) Nordisk med 43 977, 1950. (2) Proc. Internat. Congr. Hematology, Cambridge, 1950. (3) Bull. Acad. Suisse Sci. Med. 3 163, 1947. (4) Scand. J. Clin. Lab. Invest. 1 162, 1949. (5) Scand. J. Clin. Lab. Invest. 2 168, 1951. (6) Aarbok for det Norske Videnskapsakademi (Proc. Norwegian Acad. Sci.) 21, 1944. (7) The Coagulation of Blood, Acta Med. Scand. Suppl., 194, 1947.

# 9. BLOOD COAGULATION Theory of A. J. Quick (1952)

Footnotes	Contributors and References
Fn 1	1a
Fn 2	2a, 3a
Fn 3	1a
Fn 4	4a
Fn 5	5a

Contributor (a) Quick, A. J.

References (1) Quick, A. J., Studies on the Enigma of the Hemostatic Dysfunction of Hemophilia, Am. J. Med. Sci. 214 272, 1947. (2) Quick, A. J., and Stefanni, M., The State of Component A (Prothrombin) in Human Blood, Evidence that it is Partly Free and Partly in an Inactive or Precursor Form, J. Lab. Clin. Med. 34 1203, 1949. (3) Quick, A. J., On the Constitution of Prothrombin, Am. J. Physiol. 140 212, 1943. (4) Quick, A. J., "The Physiology and Pathology of Hemostasis," Lea and Febiger, Philadelphia, 1951. (5) Quick, A. J., and Favre-Gilly, J., Fibrin, a Factor Influencing the Consumption of Prothrombin in Coagulation, Am. J. Physiol. 158 387, 1949.

# 10. BLOOD COAGULATION Theory of W. H. Seegers (1952)

Footnotes	Contributors and References	Footnotes	Contributors and References
Fn 1	2a, 6a	Fn 4	1a, 2a
Fn 2	2a	Fn 5	2a
Fn 3	1a	Fn 6	3a, 4a, 5a

Contributor (a) Seegers, W. H.

References (1) Ware, A. G., and Seegers, W. H.. Serum Ac-globulin: Formation from plasma Ac-globulin; role in blood coagulation, partial purification, properties, and quantitative determination, *Am. J. Physiol.* 152:567, 1948.  
(2) Ware, A. G., Fahey, J. L., and Seegers, W. H.. Studies with platelet extracts: Their effect on the interaction of

in "The Enzymes," Vol. I, Part 1, edited by J. B. Sumner and K. Myrback, Academic Press, 1950. (b) Milstone, J. H.. Activation of prothrombin by platelets plus globulin, *Proc. Soc. Exp. Biol. Med.* 68:229, 1948.

# 11. BLOOD COAGULATION Theory of M. Stefanini (1952)

Footnotes	Contributors and References	Footnotes	Contributors and References
Fn 1	1a	Fn 5	
Fn 2	2a	Fn 6	5a, 6a, 7a
Fn 3	3a	Fn 7	8a
Fn 4	4a	Fn 1-7	9a

Contributor (a) Stefanini, M.

References (1) Quick, A. J., Shanberge, J. N., and Stefanini, M., *Am. J. Med. Sci.* 217:198, 1950. (2) Stefanini, M., and Crosby, W. H., *Proc. Soc. Exp. Biol. Med.* 74:370, 1950. (3) Stefanini, M., *Acta Med. Scand.* (in press) (4) Stefanini, M., and Crosby, W. H., *Am. J. Clin. Path.* 20:1026, 1950. (5) Quick, A. J., and Stefanini, M., *Proc. Soc. Exp. Biol. Med.* 67:111, 1948. (6) Stefanini, M., *Experientia* 5:330, 1949. (7) Wilson, S. J., *Arch. Int. Med.* 69:647, 1942. (8) Stefanini, M., and Zannoni, L., *Fed. Proc.* 10:132, 1951. (9) Stefanini, M., *Blood* 6:84, 1951.

# 12. BLOOD COAGULATION Theory of L. M. Tocantins (1952)

Contributor (a) Tocantins, L. M.

References (1) Tocantins, L. M.. Coagulation of Circulating and Shed Blood Results from Disruption of the Dynamic Equilibrium Existing between Anticoagulant and Coagulant Components of the Blood Responsible for the Maintenance of its Fluidity and Coagulability, *Surg. Clin. N. Amer.*, 1935, Dec 1949. (2) Tocantins, L. M., *Am. J. Physiol.* 139:265, 1941. (3) Tocantins, L. M., *Proc. Soc. Exp. Biol. Med.* 55:291, 1944. (4) Tocantins, L. M., *Fed. Proc.* 1:156, 1946. (5) Tocantins, L. M., *Am. J. Physiol.* 143:67, 1946. (6) Tocantins, L. M., *Blood* 1:156, 1946.



## 13. BLOOD COAGULATION TIME: MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-E	1a	11 A-E	2a	16 AB DE	9a
2 A-D	1c	12 A DE	4b	17 AB	10
3 A-E	2a	13 AB DE	7a	17 C-E	10a
4,5 A-E	3a	14 AB DE	6a	18 A-E	11
6-9 AB DE	4b	15 AB D	8	Fn 3	8
10 A-E	5a				

Contributors: (a) Diggs, L.W. (b) Lewis, J.H. (c) White, P.D.

References: (1) Lee, R. I., and White, P.D., *Am. J. Med. Sci.* 145:495, 1913. (2) Tocantins, L. M., *Med. Clin. N. Amer.* 30:1361, 1946. (3) Aggeler, P.M., Howard, J., and Lucia, S.P., *Blood* 1:472, 1946. (4) Lewis, J.H., unpublished data. (5) Copley, A. L., and Houlahan, Science, Dec. 1, 1944. (6) Gibbs, O.S., *Quart. J. Med.* 17:312, 1934. (7) Diggs, L.W., unpublished data. (8) Wintrobe, M. M., "Clinical Hematology," p. 205, 207, Lea & Febiger, Phila., 1946. (9) Quack, A. J., "The Hemorrhagic Diseases and the Physiology of Hemostasis." C. C. Thomas, Springfield, Ill., 1942. (10) Cheney, G., *Am. J. Med. Sci.* 201:325, 1942. (11) Nygaard, K. K., "Hemorrhagic Diseases." C. V. Mosby Co., St. Louis, Mo., 1941.

## 14. HEPARIN-RETARDED COAGULATION TIME: MAN

1,2 A-E	1a	10 A-U	J
3-9 ABC	2	10 EF	3b
3-9 DEF	2b		

Contributors: (a) de Takats, G. (b) Diggs, L.W.

References: (1) de Takats, G., *Angiology* 1:317, 1950. (2) Waugh, T.R., and Ruddick, D.W., *Canad. Med. J.* 50:547, 1944. (3) Tuft, H.S., and Rosenfield, R.E., *Am. J. Clin. Path.* 17:862, 1947.

## 15. HEPARIN TOLERANCE MAN

Data Coordinates and Footnotes	Contributors and References
1 AB	1
1 DE	1a
2 A-E	2

Contributor: (a) Diggs, L.W.

References: (1) de Takats, G., *Surg. Gyn. Obst.* 77:31, 1943. (2) Hagedorn, A., and Barker, N.W., *Am. Heart J.* 35:603, 1948.

# 16. SKIN BLEEDING TIME. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-E	1,2b	5 A-E	3a, 7b
2 A-E	3a	6 A-E	6a, 7b
3 A-E	4ab	7 A-E	8
4 A-E	5a		

**Contributors** (a) Diggs, L.W. (b) Lewis, J.H.

**References.** (1) Wintrobe, M.M., "Clinical Hematology" p 210. Lea & Febiger, Phila. 1946 (2) Duke, W.W., J. Am. Med. Assoc. 55 1185, 1910. (3) Tocantins, L.M., Med. Clin. N. Amer. 130 1361, 1946. (4) Copley, A.L. and Lalich, J.J., J. Clin. Invest. 21:145, 1942 (5) Aggeler, P.M., Howard, J., and Lucia, S.P., Blood 1 472, 1946. (6) Diggs, L.W., unpublished data. (7) Ivy, A.C., Shapiro, P.F., and Melnick P., Surg. Gyn. Obst. 60 781, 1935. (8) Ivy, A.C., Nelson, D., and Bucher, G.R., J. Lab. Clin. Med. 26 1812, 1941.

# 17. BLOOD CLOT RETRACTION, VOLUME, TIME MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-D	1a	4 CD	4a
2 A CD	4a	5 A-D	3a
3 A-D	2a		

**Contributor.** (a) Diggs, L.W.

**References** (1) Aggeler, P.M., Howard, J., and Lucia, S.P., Blood 1 472, 1946 (2) Tocantins, L.M., Med. Clin. N. Amer. 130 1361, 1946. (3) Hirschboeck, J.S., J. Lab. Clin. Med. 33 347, 1948. (4) Diggs, L.W., unpublished data.

# 18. PLASMA PROTHROMBIN TIME MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 AB	1	5 A-E	8	10 AB DE	2
1 C EF	1b	5 F	8b	10 C F	2b
2 AB	3	6 AB DE	4	11 AB	7
2 C EF	3b	6 C F	4b	11 C EF	7b
3 AB DE	5	7 AB DE	8	12 AB E	6
3 C F	5b	7 C E	8b	12 C F	6ab
4 A-F	9	8, 9 A-F	9		

**Contributors** (a) Diggs, L.W. (b) Owen, C.A.

**References** (1) Quick, A.J., Am. J. Clin. Path. 15 560, 1945 (2) Kato, K. and Poncher, H.G., J. Am. Med. Assoc. 114 749, 1940 (3) Magath, T.B. and Hurn, M., Am. J. Clin. Path. Tech. Suppl. 3 187 1939 (4) Page, R.C. and Russell, H.K., J. Lab. Clin. Med. 26 1366, 1941 (5) Witts, L.J. and Hobson, F.C.G., Brit. Med. J. 1 575, 1942 (6) Ziffren, S.E., Owen, C.A., Hoffman, G.R., and Smith, H.P., Am. J. Clin. Path. 10 13, 1940. (7) Karabin, J.E., and Anderson, E.R., J. Lab. Clin. Med. 26 723, 1941 (8) Shapiro, S., Sherwin, B., Redish, M., and Campbell, H.A., Proc. Soc. Exp. Biol. Med. 50 85, 1942 (9) Aggeler, P.M., Howard, J., Lucia, S.P., Clark, W., and Astaff, A., Blood 1 220, 1946

# 19. PLASMA PROTHROMBIN, RELATIVE LEVELS VERTEBRATES

Data Coordinates and Footnotes	Contributors and References
1-11 AB Fn 4	Calc. from lab 1

Contributors: (a) Diggs, L.W. (b) Warner, E.D.

References: (1) Warner, E.D., Brinkhous, K.M., and Smith, H.P., Am. J. Physiol. 125:296, 1939.

# 20. PLASMA ACCELERATOR GLOBULIN, RELATIVE LEVELS: VERTEBRATES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,3-6,8B 1-9 C	1c Calc fr. 2abc	Fn 3 Fn 4	1c 4c	Fn 5	2 abc, 3c

Contributors (a) Owen, C.A. (b) Seegers, W.H. (c) Stefanini, M.

References: (1) Quick, A.J., and Stefanini, M., J. Lab Clin Med. 33:819, 1948 (2) Murphy, R.C., and Seegers, W.H., Am. J. Physiol. 154:134, 1948 (3) Ware, A.G., and Seegers, W.H., Am. J. Physiol. 152:567, 1948. (4) Ware, A.G., and Seegers, W.H., J. Biol. Chem. 172:699, 1948

# 21. BLOOD PLATELET COUNT MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A 2,3 AB 4,5 A 4,5 B 6 AB	1a 1c 2a 2,3 4a	7 AB 8 AB 9 AB 10 AB 11 AB	5cd 6ab 7cd 8c 9c

Contributors (a) Brecher, G. (b) Cronkite, E.P. (c) Diggs, L.W. (d) Lewis, J.H.

References (1) Tocantins, L.M., Am. J. Med. Sci. 192:150, 1936 (2) Kristenson, A., "Akademische Abhandlung," Uppsala, 1924 (3) Sunderman, W.F., and Boerner, F., "Normal Values in Clinical Medicine" W B Saunders Co., Phila., 1949. (4) Kristenson, A., Acta Med Scand 69:227, 1928 (5) Aggeler, P.M., and Lucia, S.P., "Hemorrhagic Disorders, A Guide to Diagnosis and Treatment," Univ of Chicago Press, Chicago, 1949 (6) Brecher, G., and Cronkite, E.P., J. Applied Physiol Dec 1950. (7) Dameshek, W., Arch Int Med 50:579, 1932 (8) Fonto, A. Deut. Z. Chir 117:176, 1912. (9) Olef, L., J Lab Clin Med 20:416, 1935.

## 22. BLOOD PLATELET COUNT: LABORATORY, FARM ANIMALS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-D	7ab	10 A-D	12ab	19 A-D	3ab
2 A-D	11ab	11 A-D	13ab	20 A-D	22ab
3, 5, 9 A-D	1ab	12 A-D	14ab	21 A-D	17ab, c
4 A-D	21ab	13 A-D	15ab	22 A-D	16ab
6 A-D	15ab	14 A-D	8ab	23 A-D	13ab
7 A-D	18ab	15 A-D	10ab	24 A-D	5ab
8, 20 A-D	2ab	16 A-D	4ab	25 A-D	14ab
9 A-D		17 A-D	9ab	26 A-D	6ab
10 A-D		18 A-D	8ab	27 A-D	

Contributors: (a) Tocantins, L. M. (b) Jaques, L. B. (c) Nice, L. B.

Reference: [1] Wright, I. S. and Lichtenfeld, A. Arch. Int. Med. 57: 241, 1936. [2] Lundquist, N. Acta paediat. Supp. 2, 20, 1937. [3] Frontali, G., "Capillari nel Bambino" edit. L. Cappelli, Bologna, 1937. [4] Gothlin, G. F. Acta paediat. 20-21, 1937. [5] Diggs, L. W. unpublished data. [6] Daildorf, G. Am. J. Dis. Child. 46: 794, 1933. [7] Aggeler, P. M., Howard, J. and Lucia, S. P. Blood 1: 472, 1946.

Note: \*quoted from [19]  
\*\*quoted from Gardner, M. V., J. Franklin Inst. 243: 77, 172, 1947.

## 23. CAPILLARY FRAGILITY MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-E	6	11 ABC E	7	14 A-C	4
2-1 ABCDE	2	12 A-D	5a	15 ABC E	1
6-10 ABC E	3b	13 ABC E	5a		

Contributor: (a) Diggs, L. W. (b) Frontali, G.

References: [1] Wright, I. S. and Lichtenfeld, A. Arch. Int. Med. 57: 241, 1936. [2] Lundquist, N. Acta paediat. Supp. 2, 20, 1937. [3] Frontali, G., "Capillari nel Bambino" edit. L. Cappelli, Bologna, 1937. [4] Gothlin, G. F. Acta paediat. 20-21, 1937. [5] Diggs, L. W. unpublished data. [6] Daildorf, G. Am. J. Dis. Child. 46: 794, 1933. [7] Aggeler, P. M., Howard, J. and Lucia, S. P. Blood 1: 472, 1946.

24. THE ABO SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Allen, F.H., Wiener, A.S., and Levine, P. Frequencies for General Blood Groups from Snyder, L. H., 20,000 Americans, cited by Boyd, W.C., in *Tabulae Biologicae* 17 Part 2, 1939. Frequencies for Subgroups from Wiener's tests on 1077 whites (N.Y.C.).

25. THE DISTRIBUTION OF THE ABO BLOOD GROUPS IN VARIOUS POPULATIONS

Data contributed by Levine, P., Wiener, A.S., and Levine, V.E. Selections from Weiner, A.S., "Blood Groups and Transfusion," 3rd ed., 1943, C.C. Thomas, Springfield, Ill., from Boyd, W.C. *Tabulae Biologicae* 17 Part 2, 1939, and from Boyd, W.C., "Genetics and the Races of Man," p.223. Little, Brown & Co., Boston, 1950.

26. HEREDITY OF THE ABO BLOOD GROUPS  
Application in Disputed Paternity

Modified from Levine, P.

27. HEREDITY OF THE ABO BLOOD SUBGROUPS  
Application in Disputed Paternity

Data contributed by Riser, W.H., modified from DeGowin, E.L., Hardin, R.C., Alsever, J.B., "Blood Transfusion," W.B. Saunders Co., 1949

28. THE Rh-Hr (CDE-cde) SYSTEM OF BLOOD FACTORS  
AND THEIR FREQUENCIES

Data contributed by Wiener, A.S., Levine, P., Allen, F.H., and Haberman, S. Frequencies for U.S.A. whites (N.Y.C.) from Wiener, A.S. Frequencies for British whites from Race, R.R., et al., 2000 English bloods, in *Blood* 3:689, 1948. Frequencies in footnote /3/ from Hill, J.M., *Am J. Clin. Path.* 17 No 6, 1947.

29. THE DISTRIBUTION OF THE Rh-Hr BLOOD TYPES  
IN VARIOUS POPULATIONS

Data contributed by Levine, P. and Wiener, A.S. Selections from Wiener, A.S., *Am J. Clin. Path.* 16 477, 1946.

### 30. THE MNS SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Allen, F.H., Wiener, A.S., and Levine, P. Frequencies for 340 English bloods, Pickles, M.M., Nature 162 66, 1946.

### 31. THE DISTRIBUTION OF THE MN BLOOD FACTORS IN VARIOUS POPULATIONS

Data contributed by Levine, P., and Wiener, A.S. Frequencies from Wiener, A.S., "Blood Groups and Transfusion," 3rd ed., 1943, except for Eskimos contributed by Levine, V.E.

### 32. HEREDITY OF THE MN BLOOD FACTORS. Application in Disputed Paternity

Modified from Levine, P.

### 33. THE K ("KELL-CELLANO") SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Allen, F.H., and Levine, P. Frequencies from Levine, P. et al., Blood 4 No. 7, 669, July, 1949 for U.S.A. whites, and from Sanger, R., et al., Rev. d'hémat. 4:32, 1949, for English whites.

### 34. THE Le ("LEWIS") SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Allen, F.H. Frequencies from Sanger, R., 132 Harvard medical students and Children's Hospital personnel, 1948. Terminology from Race, R.R., et al., Brit. J. Exp. Path. 30:73, 1949.

### 35. THE Lu ("LUTHERAN") SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Haberman, S., and Allen, F.H. Nomenclature from Nature 161 580, 1949. Frequencies for 582 English bloods, Callender, S.T., and Race, R.R., Ann. Eugen. 13:102, 1946. Frequencies in footnote 1/ for 110 U.S. (Tex.) bloods, Hill, J.M., Haberman, S., and Guy, R., unpublished data.

### 36. THE P SYSTEM OF BLOOD FACTORS AND THEIR FREQUENCIES

Data contributed by Allen, F.H. Frequencies from Sanger, R., 132 Harvard medical students and Children's Hospital personnel, 1948.

# 37 BLOOD ERYTHROCYTE VALUES MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-3 ABC 4-7 ABC & Fn 1, 2 8, 9 A-C Fn 4 10-12 ABC 13 A-C & Fn 5 14 A-C 15 A-C & Fn 6 16 AB	17a, 9j 9j, 11j 17a, d, 6o, 9j 9j 17a, 9j 17a, 10j 7k 17a, 9j 7k	17 A-C & Fn 7 18 AB 19-21 ABC Fn 8 22 A-C & Fn 9 23 A-C & Fn 10 24-26 ABC 27 A-C & Fn 11 28-33 ABC & Fn 12 34 A-C	g, 9j 7k 9j 7k j, 8c 8cn 9j 3b, 9j, 18 6h 9j	35 AB 36, 37 A-C 38, 39 A-C 40, 41 A-C 42, 43 A-C 44, 45 A-C 46 A-C 47, 48 AB & Fn 14 49 A-C 50-52 AB & Fn 15 53-65 AB	17a, 9j 12l 13i 14i 15m 16m 5b 1c 2f 4e, 19b 17a, 9j, 19b

Contributors: (a) Bethell, F. H. (b) Collier, H. B. (c) Dole, V. P. (d) Drabkin, D. L. (e) Ferguson, J. H. (f) Gram, H. C. (g) Guest, G. M. (h) Heath, C. W. (i) Hirschboeck, J. S. (j) Osgood, E. E. (k) Ponder, E. (m) Riser, W. H., Jr. (n) Van Slyke, D. D. (o) Wintrobe, M. M.

References: (1) Abramson, H. A., J. Gen. Physiol. 12:711, 1929. (2) Gram, H. C., Am. J. Med. Sci. 168 521, 1924. (3) Drabkin, D., Am. J. Clin. Invest. 26 636, 1947. (4) Wintrobe, M. M., et al., J. Biol. Chem. 179 463, 1949. (5) Parpart, A. K., et al., J. Clin. Invest. 26 636, 1947. (6) Wintrobe, M. M., "Clinical Hematology," Lea & Febiger, Phila., 1946. (7) Ponder, E., "Hemolysis and Related Phenomena," Grune & Stratton, New York, 1948. (8) Van Slyke, D. D., et al., Am. J. Physiol. 128:382, 1940. (9) Osgood, E. E., Arch. Int. Med. 58 849, 1935. (10) Donelson, E. G., et al., Am. J. Physiol. 128:382, 1940. (11) Ponder, E., Am. Rev. Tuberc. 14:94, 1926. (12) Wintrobe, M. M., and Landsberg, J. W., Publishers, 1944. (13) Westergren, A., Am. Rev. Tuberc. 14:94, 1926. (14) Giffin, H. Z., and Sanford, A. H., J. Lab. Clin. Med. 4 465, 1919. (15) Bethell, F. H., J. Lab. Clin. Med. 20 1122, 1935. (16) Giffin, H. Z., and Sanford, A. H., J. Lab. Clin. Med. 4 465, 1919. (17) Bethell, F. H., "Clinical Laboratory Diagnosis and Essentials of Hematology," The Edwards Letter Shop, Ann Arbor, Michigan, 1948. (18) Grinstein, M., and Moore, C. V., J. Clin. Invest. 28 505, 1949. (19) Callender, S. T., Powell, E. O., and Witts, L. J., J. Path. Bact. 57 129, 1945. (20) Gregersen, M. I., and Schiro, H., Am. J. Physiol. 121:284, 1918.

## 38 BLOOD ERYTHROCYTE, HEMATOCRIT, HEMOGLOBIN AND THROMBOCYTE VALUES, BIRTH TO MATURITY MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
*1-18 BC 20-22 BC 1, 3-12 DE 22 DE *1-18 F 20-22 F *1-18 G	7bi, 8bi, d, 14e, 15e, 1f, 2j 9g 4h 10g d, 14e, 15e, 2j 9g d, 2j	20 G 21 G 22 G 1-18 IH 1-18 J	Calc fr g, (20F)x 10/(20B) Calc fr g, (21F)x 10/(21B) Calc fr g, (22F)x 10/(22B) d, 14e, 15e, 1f, 3f, 5f, 2j Calc fr (i)-(18H) x 10/(1-18H)	1-18 K 20-22 HI 20-22 J 20-22 K *1, 3-12 LM 22 LM Fn 4 Fn 7	Calc fr (1-18H) x 100/(1-18F) 9g 9g 9g, 16a 1f 6, 11c, 12 13 7bi, 8bi

\*Note These values are smoothed means and sample ranges from plotted curves based on averages of the data referred to.

Contributors: (a) Bethell, F. H. (b) DeMarsh, Q. B. (c) Diggs, L. W. (d) Glaser, K. (e) Guest, G. M. (f) Mayerson, H. S. (g) Osgood, E. E. (h) Washburn, A. H. (i) Windle, W. F. (j) Wintrobe, M. M.

References: (1) Merritt, K. K., and Davidson, L. T., Am. J. Dis. Child 46 991, 1900, 1001, 1008, 1933. (2) Wintrobe, M. M., "Clinical Hematology," 2nd ed., p. 73, Lea & Febiger, Philadelphia, 1946. (3) Williamson quoted by Merritt, K. K., and Davidson, L. T., Am. J. Dis. Child 46 991, 1933. (4) Washburn, A. H., Am. J. Dis. Child 62 530, 1941. (5) Elvehjem, C. A., et al., Am. J. Dis. Child 46 105, 1933. (6) Rees, H. M., and Ecker, E. E., J. Am. Med. Assoc 116:2568, 1941. (7) DeMarsh, Q. B., Alt, H. L., Windle, W. F., and Hillis, D. S., J. Am. Med. Assoc 116:2568, 1941. (8) DeMarsh, Q. B., Alt, H. L., Windle, W. F., Am. J. Dis. Child 75 850, 1948. (9) Osgood, E. E., Arch. Int. Med. 58 849, 1935.

(Continued on the next page)

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 C-E	1a	7 C-E	7a	13 C-E	13a
2 C-E	2a	8 C-E	8a	14 C-E	14a
3 C-E	3a	9 C-E	9a	15 C-E	15a
4 C-E	4a	10 C-E	10a	16 C-E	16a
5 C-E	5a	11 C-E	11a	17 C-E	17a
6 C-E	6a	12 C-E	12a	18 C-E	18a

References: (1) Osgood, E. E., Arch. Int. Med. 37 685, 1926. (2) Osgood, E. E., Arch. Int. Med. 56 849, 1935 (3) Waintrub, M. M., and Miller, M. W., Arch. Int. Med. 43 96, 1929 (4) Foster, F. C., and Johnson, J. R., Proc. Soc. Exp. Biol. Med. 28 929, 1931. (5) Walters, O. S., J. Lab. Clin. Med. 19 851, 1934 (6) Sachs, A., Levine, V. E., and Fishbein, A. D., Arch. Int. Med. 56 849, 1935 (7) Hurler, E. R., and Stoker, R., Folia haemat. 55 333, 1937. (8) Compt. rend. Soc. Biol. 108 133, 1931. (9) Heilmeyer, L., and Hansold, L., Arch. 1928. (10) Hamre, C. J., and Au, M. H. J., Indian J. Med. Res. 23 305, 1935. (11) Sokhey, S. S., Gokhale, S. K., Malandar, M. A., and Bilimoria, H. S., Indian J. Med. Res. 25 505, 1937 (12) Jervell, O., and Waaler, J. H. M., Norsk mag. lægevid. 35 1141, 1934. (13) Hurtado, A., Pons, M. J., and Merino, C., "La Anemia de la enfermedad de Carrion" Libreria e Imprenta Gil, Lima, Peru, 1938 (14) Hurtado, A., Merino, C., and Delgado, E., Arch. Int. Med. 75 284 1945

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 C-F	1a	7 C-F	7a	13 C-F	10a
2 C-F	2a	8 C-F	8a	14 C-F	9a
3 C-F	3a	9 C-F	9a	15 C-F	12a
4 C-F	4a	10 C-F	9a	16 C-F	9a
5 C-F	5a	11 C-F	10a	17 C-F	9a
6 C-F	6a	12 C-F	11a	18 C-F	13a

References: 20 581, Wschr. Quart. 2 Terán, I Delgado Dill, D. and Fonio, G. A., Rev. Soc. argent. biof. 1935 (4) Burgi, Kuno, Schweiz. med. Wochenschr. 1936 (6) Lukatsky, I., 1937 (7) Gill, J. R., and Hurtado, A. Merino, C. and Talbot, J. H., 1950 (12) Talbot, J. H., 1951 (13) Talbot, J. H., 1952 (14) Talbot, J. H., 1953 (15) Talbot, J. H., 1954 (16) Talbot, J. H., 1955 (17) Talbot, J. H., 1956 (18) Talbot, J. H., 1957 (19) Talbot, J. H., 1958 (20) Talbot, J. H., 1959 (21) Talbot, J. H., 1960 (22) Talbot, J. H., 1961 (23) Talbot, J. H., 1962 (24) Talbot, J. H., 1963 (25) Talbot, J. H., 1964 (26) Talbot, J. H., 1965 (27) Talbot, J. H., 1966 (28) Talbot, J. H., 1967 (29) Talbot, J. H., 1968 (30) Talbot, J. H., 1969 (31) Talbot, J. H., 1970 (32) Talbot, J. H., 1971 (33) Talbot, J. H., 1972 (34) Talbot, J. H., 1973 (35) Talbot, J. H., 1974 (36) Talbot, J. H., 1975 (37) Talbot, J. H., 1976 (38) Talbot, J. H., 1977 (39) Talbot, J. H., 1978 (40) Talbot, J. H., 1979 (41) Talbot, J. H., 1980 (42) Talbot, J. H., 1981 (43) Talbot, J. H., 1982 (44) Talbot, J. H., 1983 (45) Talbot, J. H., 1984 (46) Talbot, J. H., 1985 (47) Talbot, J. H., 1986 (48) Talbot, J. H., 1987 (49) Talbot, J. H., 1988 (50) Talbot, J. H., 1989 (51) Talbot, J. H., 1990 (52) Talbot, J. H., 1991 (53) Talbot, J. H., 1992 (54) Talbot, J. H., 1993 (55) Talbot, J. H., 1994 (56) Talbot, J. H., 1995 (57) Talbot, J. H., 1996 (58) Talbot, J. H., 1997 (59) Talbot, J. H., 1998 (60) Talbot, J. H., 1999 (61) Talbot, J. H., 2000 (62) Talbot, J. H., 2001 (63) Talbot, J. H., 2002 (64) Talbot, J. H., 2003 (65) Talbot, J. H., 2004 (66) Talbot, J. H., 2005 (67) Talbot, J. H., 2006 (68) Talbot, J. H., 2007 (69) Talbot, J. H., 2008 (70) Talbot, J. H., 2009 (71) Talbot, J. H., 2010 (72) Talbot, J. H., 2011 (73) Talbot, J. H., 2012 (74) Talbot, J. H., 2013 (75) Talbot, J. H., 2014 (76) Talbot, J. H., 2015 (77) Talbot, J. H., 2016 (78) Talbot, J. H., 2017 (79) Talbot, J. H., 2018 (80) Talbot, J. H., 2019 (81) Talbot, J. H., 2020 (82) Talbot, J. H., 2021 (83) Talbot, J. H., 2022 (84) Talbot, J. H., 2023 (85) Talbot, J. H., 2024 (86) Talbot, J. H., 2025 (87) Talbot, J. H., 2026 (88) Talbot, J. H., 2027 (89) Talbot, J. H., 2028 (90) Talbot, J. H., 2029 (91) Talbot, J. H., 2030 (92) Talbot, J. H., 2031 (93) Talbot, J. H., 2032 (94) Talbot, J. H., 2033 (95) Talbot, J. H., 2034 (96) Talbot, J. H., 2035 (97) Talbot, J. H., 2036 (98) Talbot, J. H., 2037 (99) Talbot, J. H., 2038 (100) Talbot, J. H., 2039 (101) Talbot, J. H., 2040 (102) Talbot, J. H., 2041 (103) Talbot, J. H., 2042 (104) Talbot, J. H., 2043 (105) Talbot, J. H., 2044 (106) Talbot, J. H., 2045 (107) Talbot, J. H., 2046 (108) Talbot, J. H., 2047 (109) Talbot, J. H., 2048 (110) Talbot, J. H., 2049 (111) Talbot, J. H., 2050 (112) Talbot, J. H., 2051 (113) Talbot, J. H., 2052 (114) Talbot, J. H., 2053 (115) Talbot, J. H., 2054 (116) Talbot, J. H., 2055 (117) Talbot, J. H., 2056 (118) Talbot, J. H., 2057 (119) Talbot, J. H., 2058 (120) Talbot, J. H., 2059 (121) Talbot, J. H., 2060 (122) Talbot, J. H., 2061 (123) Talbot, J. H., 2062 (124) Talbot, J. H., 2063 (125) Talbot, J. H., 2064 (126) Talbot, J. H., 2065 (127) Talbot, J. H., 2066 (128) Talbot, J. H., 2067 (129) Talbot, J. H., 2068 (130) Talbot, J. H., 2069 (131) Talbot, J. H., 2070 (132) Talbot, J. H., 2071 (133) Talbot, J. H., 2072 (134) Talbot, J. H., 2073 (135) Talbot, J. H., 2074 (136) Talbot, J. H., 2075 (137) Talbot, J. H., 2076 (138) Talbot, J. H., 2077 (139) Talbot, J. H., 2078 (140) Talbot, J. H., 2079 (141) Talbot, J. H., 2080 (142) Talbot, J. H., 2081 (143) Talbot, J. H., 2082 (144) Talbot, J. H., 2083 (145) Talbot, J. H., 2084 (146) Talbot, J. H., 2085 (147) Talbot, J. H., 2086 (148) Talbot, J. H., 2087 (149) Talbot, J. H., 2088 (150) Talbot, J. H., 2089 (151) Talbot, J. H., 2090 (152) Talbot, J. H., 2091 (153) Talbot, J. H., 2092 (154) Talbot, J. H., 2093 (155) Talbot, J. H., 2094 (156) Talbot, J. H., 2095 (157) Talbot, J. H., 2096 (158) Talbot, J. H., 2097 (159) Talbot, J. H., 2098 (160) Talbot, J. H., 2099 (161) Talbot, J. H., 2100 (162) Talbot, J. H., 2101 (163) Talbot, J. H., 2102 (164) Talbot, J. H., 2103 (165) Talbot, J. H., 2104 (166) Talbot, J. H., 2105 (167) Talbot, J. H., 2106 (168) Talbot, J. H., 2107 (169) Talbot, J. H., 2108 (170) Talbot, J. H., 2109 (171) Talbot, J. H., 2110 (172) Talbot, J. H., 2111 (173) Talbot, J. H., 2112 (174) Talbot, J. H., 2113 (175) Talbot, J. H., 2114 (176) Talbot, J. H., 2115 (177) Talbot, J. H., 2116 (178) Talbot, J. H., 2117 (179) Talbot, J. H., 2118 (180) Talbot, J. H., 2119 (181) Talbot, J. H., 2120 (182) Talbot, J. H., 2121 (183) Talbot, J. H., 2122 (184) Talbot, J. H., 2123 (185) Talbot, J. H., 2124 (186) Talbot, J. H., 2125 (187) Talbot, J. H., 2126 (188) Talbot, J. H., 2127 (189) Talbot, J. H., 2128 (190) Talbot, J. H., 2129 (191) Talbot, J. H., 2130 (192) Talbot, J. H., 2131 (193) Talbot, J. H., 2132 (194) Talbot, J. H., 2133 (195) Talbot, J. H., 2134 (196) Talbot, J. H., 2135 (197) Talbot, J. H., 2136 (198) Talbot, J. H., 2137 (199) Talbot, J. H., 2138 (200) Talbot, J. H., 2139 (201) Talbot, J. H., 2140 (202) Talbot, J. H., 2141 (203) Talbot, J. H., 2142 (204) Talbot, J. H., 2143 (205) Talbot, J. H., 2144 (206) Talbot, J. H., 2145 (207) Talbot, J. H., 2146 (208) Talbot, J. H., 2147 (209) Talbot, J. H., 2148 (210) Talbot, J. H., 2149 (211) Talbot, J. H., 2150 (212) Talbot, J. H., 2151 (213) Talbot, J. H., 2152 (214) Talbot, J. H., 2153 (215) Talbot, J. H., 2154 (216) Talbot, J. H., 2155 (217) Talbot, J. H., 2156 (218) Talbot, J. H., 2157 (219) Talbot, J. H., 2158 (220) Talbot, J. H., 2159 (221) Talbot, J. H., 2160 (222) Talbot, J. H., 2161 (223) Talbot, J. H., 2162 (224) Talbot, J. H., 2163 (225) Talbot, J. H., 2164 (226) Talbot, J. H., 2165 (227) Talbot, J. H., 2166 (228) Talbot, J. H., 2167 (229) Talbot, J. H., 2168 (230) Talbot, J. H., 2169 (231) Talbot, J. H., 2170 (232) Talbot, J. H., 2171 (233) Talbot, J. H., 2172 (234) Talbot, J. H., 2173 (235) Talbot, J. H., 2174 (236) Talbot, J. H., 2175 (237) Talbot, J. H., 2176 (238) Talbot, J. H., 2177 (239) Talbot, J. H., 2178 (240) Talbot, J. H., 2179 (241) Talbot, J. H., 2180 (242) Talbot, J. H., 2181 (243) Talbot, J. H., 2182 (244) Talbot, J. H., 2183 (245) Talbot, J. H., 2184 (246) Talbot, J. H., 2185 (247) Talbot, J. H., 2186 (248) Talbot, J. H., 2187 (249) Talbot, J. H., 2188 (250) Talbot, J. H., 21

Data contributed by Bethell, F. H., from Bethell, F. H., Gardner, S. H. and MacKinnon, F. Ann Int Med 13:91, 1939.



# 42. BLOOD ERYTHROCYTTIC VALUES: MAMMALS, LABORATORY

Data Coordinates	Contributors and References	Data Coordinates	Contributors and References	Data Coordinates	Contributors and References
Cat 1B	Av. of means from 4,8,15, 34,36,44,46,49,50,82,94,95, 115,128,129,147,162,198, 199,200,201,230,235	2J	Av. of means from 7,8,21a, 22a, 27, 66, 84a, 85, 137, 139, 160, 193, 242	Rabbit 5B	Av. of means from 1,5,14, 15,16,17,19,20,28,29,30,31, 34,39,40,43,51,52,56,59,60, 61,62,63,67,68,76,77,78,82, 86,90,91,97,100,101,107, 102,104,109,112,113,114, 120,121,132,133,134,140, 143,144,151,153,156,165, 166,167,168,169,170,171, 173,175,177,179,180,181, 188,182,191,194,195,196, 204,207,216,217,218,219, 220,221,222,227,228,233, 232,235,241,242,252
1C	Av. of ranges from 34,36, 49,82,95,115,198,199,201	2K	Av. of ranges from 7, 242	5C	Av. of ranges from 1,5,16, 17,19,20,28,29,30,31,43,51, 52,59,60,61,62,67,76,78,82, 86,90,91,97,101,102,103,109, 113,114,120,121,132,133, 140,143,168,169,170,171, 175,179,180,188,195,204, 207,216,217,218,221,233,252
1D	116,119	2L	Av. of means from 27,192d	5D	Av. of means from 82,198, 221
1F	Av. of means from 9,13,15, 82,93,94,115,147,148,183, 202,203,236	2M	Calc. from (2L)x100/(2J)	5E	221
1G	Av. of ranges from 9,15,94, 203,236	2N	Av. of ranges from 7,27, 137,160,242	5F	189
1H	129	2O	Calc. from (2L)x10/(2B)	5G	c
1I	c	2P	Av. of ranges from 7,27, 137,160,161,242	5H	Av. of means from 189,242
1J	129	2Q		5I	Av. of means from 43,104, 196,220,242
1K	Calc. from 129	Guinea Pig 3B	Av. of means from 12,15, 23,33,34,38,42,44,56,73,82, 86,87,89,94,98,115,122,149, 153,156,163,173,177,178, 210,211,212,232,244	5J	Calc. from 242
1L	Av. of means from 129,50, 201,225,230,248	3C	Av. of ranges from 12,23, 33,34,38,82,86,87,88,115, 122,149,156,178,210,232	5K	Av. of means from 40,58, 91,104,220,242
1M	Calc. from (1L)x100/(1J)	3D	Av. of means from 3,10,12, 87,88,211	5L	Calc. from 242
1N	c	3E	Av. of ranges from 12,87,88	5M	Calc. from (5L)x100/(5J)
1O	Calc. from (1L)x10/(1B)	3F	Av. of means from 15,33, 44,71,82,93,94	5N	Range interval fr. c
1P	Range interval fr. c	3G	Av. of means from 56,244	5O	Calc. from (5L)x10/(5B)
1Q		3H	Av. of means from 56,87, 117,193,244	5P	c
Dog 2B	Av. of means from 2,6,7, 18,20,24,25,26,27,28,29,32, 34,35,37,44,47,49,51,52,53, 54,55,57,70,82,83,94,96,97, 110,115,123,124,125,126, 128,130,131,132,133,134, 135,136,137,149 through 161,174,175,176,179,182, 184,185,186,190,191,192c, 200,204,205,206,207,208, 227,231,232,235,237,238, 239,240,241,242,246,249, 251	3I	Av. of means from 56,87, 87	5Q	Calc. from (5L)x10/(5B)
2C	Av. of ranges from 2,6,18, 20,25,28,29,32,34,35,37,44, 49,51,52,53,54,55,70,82,83, 96,97,110,123,124,125,126, 128,131,132,133,135,136, 149,152,154,157,158,159, 174,175,179,182,184,185, 186,190,192,204,206,207, 208,231,232,238,239,240, 246	3J	Av. of means from 56,87, 212,244	5R	Av. of means from 41,45, 48,58,105,106,127,146,197, 229,245,250,254
2D	Av. of means from 116,118, 192d	3K	Av. of means from 56,87, 212,244	6C	Av. of ranges from 105, 146,250
2E	192d	3L	Calc. from (3L)x100/(3J)	6D	Av. of means from 27,45, 48,223,224,229,255,256
2F	Av. of means from 13,44, 53,71,72,82,93,94,108,115, 137,138,148,162,183,202, 203,204,209,226,233,243	3M	Range interval fr. c	6E	234
2G	Av. of ranges from 44, 53, 72, 84, 115, 138, 202, 203, 209	3N	Calc. from (3L)x10/(3B)	6F	189
2H	Av. of means from 7,27 137, 160, 242	3O	Range interval fr. c	6G	c
2I	Range interval fr. 7,27,137 160, 242	3P	Range interval fr. c	6H	242
		3Q		6I	Av. of means from 11,48, 63,69,80,107,117,127,172, 242,253
		Mouse 4B	Av. of means from 45,56, 58,74,81,82,111,115,118, 141,142,153,187,213,244	6J	242
		4C	Av. of ranges from 74,75, 115	6K	Av. of means from 11,45, 48,58,99,106,172,197,229, 245
		4D	Av. of means from 62,202	6L	Calc. from 172
		4F	Av. of means from 56,187, 244	6M	Calc. from (6L)x100/(6J)
		4H	c	6N	c
		4I	Av. of means from 56,187, 183	6O	Calc. from (6L)x10/(6B)
		4J	Av. of means from 56,58, 79,80,89,99,214,111,215	6P	Range interval fr. c
		4L	Calc. from 89	6Q	
		4M	Calc. from (4L)x100/(4J)		
		4N	Range interval fr. c		
		4O	Calc. from (4L)x10/(4D)		
		4P	Range interval fr. c		
		4Q			

(Continued on the next page)





verdächtigter Flecke in Criminalfällen." Leipzig, 1848. \* /203/ Schmidt, P., "Experimentelle Beiträge zur Pathologie

Physiologie der Haussäugethiere." 2. I, 163-218, 1890. \* /227/ Tallqvist, T.W., Ztschr. f. klin. Med. 61: 427-532, 1907.  
 \* /228/ Taussig, O., Arch. f. exper. Path. u. Pharmacol. 39: 161-79, 1892. /229/ Totter et al, J. Biol. Chem. 178,  
 847, 1949. \*\*\* /230/ Vaulont, H., "Untersuchungen des Katzen- und Pferdeblutes," Ztschr. f. Biol. 85: 241, 1935.  
 \* /231/ Vierordt, K., Arch. f. physiol. Heilk. 11: 1852, (quoted by Turowski). \* /232/ Voss, G. von, Deutsches Arch.  
 f. klin. Med. 58: 489-522, 1897. \* /233/ Vierordt, K., Arch. f. physiol. Heilk. 262, 1854 (quoted by Storch). \* /234/  
 Vollmer, E.F., Gordon, A.S., Levenstein, L., and Charipper, H.A., Endocrinology 23: 879, 1935. \* /235/ Wells, J.J.,  
 and Sutton, J.E., Am. J. Physiol. 39: 31-8, 1915. \* /236/ Welcker, H., Ztschr. f. rat. Med. 20, 1882. \* /237/ Welcker,  
 H., Prag Vrtljschr. 44, 11, 1854, Also Ztschr. f. rat. Med. 4: 145, 1858. \* /238/ Wetzel, J., Ztschr. f. Tiermed. 14:  
 1-47, 1910. \* /239/ Whipple, G.H. and Hooper, C.W., Am. J. Physiol. 42: 343-59, 1916. \* /240/ Whipple, G.H., Hooper,

## 43. HEMATOCRIT VALUES AND HEMOGLOBIN CONCENTRATIONS OF BLOOD. VERTEBRATES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B D	39d	7 F	Calc. from (7D x 100/7B)	13 B	1, 3, 12
1 F	19d, 40b	8 B	Av. of 1, 12-18	13 D	1, 36
2 B D	1	8 C	Calc. from 13	13 F	Calc. from (13D x 100/13B)
2 F	Calc. from (2D x 100/2B)	8 D	Av. of 14-24	14 B	3, 12
3 B	2, 3, 4	8 E	17	14 D	35
3 C	Calc. from 4	8 F	Calc. from (8D x 100/8B)	14 F	Calc. from (14D x 100/14B)
3 D	Av. of 2, 5, 6, 7	9 B	Av. of c, 3, 12, 25, 26	15 B	3, 12
3 E	Calc. from 2	9 C	Calc. from c	15 D	1
3 F	Calc. from (3D x 100/3B)	9 D	Av. of c, 25-27	15 F	Calc. from (15D x 100/15B)
4 B	c	9 E	Calc. from c	16 B-F	36a
4 C	Calc. from c	9 F	Calc. from (9D x 100/9B)	17 B D	1, 37
4 D	Av. of c, 1, 8	10 B	Av. of 1, 11, 28	17 F	Calc. from (17D x 100/17B)
4 E	Calc. from c	10 C-E	Calc. from 11	18 B-F	36a
4 F	Calc. from (4D x 100/4B)	10 F	Calc. from (10D x 100/10B)	19 B	1, 12
5 B-E	9	11 B	1, 12, 29, 30	19 D	1
5 F	Calc. from (5D x 100/5B)	11 D & F n 6	1, 31, 32	19 F	Calc. from (19D x 100/19B)
6 B	Av. of 3, 8, 10, 11, 12	11 F	Calc. from (11D x 100/11B)	20 E	38a
6 C	Calc. from 11	12 B	3, 12	21 B D	1
6 D	Av. of 10, 11	12 D	Av. of 31, 33	21 F	Calc. from (21D x 100/21B)
6 E	Calc. from 11	12 E	Calc. from 33	22 B-F	8
6 F	Calc. from (6D x 100/6B)	12 F	Calc. from (12D x 100/12B)		
7 B D	10				

(Continued on the next page)

Contributors: (a) Atlans, F.D. (b) Bethell, F.H. (c) Cronkite, E.P. (d) Osgood, E.E.

References: (1) Wintrobe, M.M., *Folia haematol.* 51:32, 1933. (2) Brody, S., et al., *Univ. Missouri Agr. Exp. Sta. Res. Bull.* 433, 1949. (3) Kerr, S.E., and Daoud, L., *J. Biol. Chem.* 109:301, 1935. (4) Ashley, A., and Guest, G.M., *J. Clin. Invest.* 13:219, 1934. (5) Allcroft, W.M., *J. Agr. Sci.* 31:320, 1941. (6) Brooks, H.J., and Hughes, J.S., *J. Nutrition* 5:35, 1932. (7) McCay, C.M., *J. Dairy Sci.* 14:373, 1931. (8) Drastisch, L., *Pflügers Arch.* 219:227, 1928. (9) Cruz & Ubatuba, *Proc. Soc. Exp. Biol. Med.* 65:321, 1947. (10) MacLeod, J., and Ponder, E., *Science* 103:73, 1946. (11) Holman, H.H., *Proc. Roy. Soc. Med.* 40:186, 1947. (12) Rapoport, S., and Guest, G.M., *J. Biol. Chem.* 138:269, 1941. (13) Overman, R.R., and Feldman, H.A., *J. Clin. Invest.* 26:1049, 1947. (14) Shukers, C.F., et al., *Folia haematol.* 60:416, 1938. (15) Bussabarger, R.A., and Ivy, A.C., *Proc. Soc. Exp. Biol. Med.* 34:151, 1936. (16) Langston, W.C., *J. Exp. Med.* 68:923, 1938. (17) Majumder, D.N., and Das Gupta, C.P., *Indian J. Med. Res.* 32:101, 1944. (18) Tyslowitz, (19) Billmoria, H.S., quoted by Jones, et al., *Blood* 2:15 45, 1945. (21) Day, P.L., et al., *Proc. Soc. Exp. Biol.* (23) Rao, quoted by Jones, et al., *Blood* 2:154, 1947. (24) 212, 1942. (25) Goetz, R., *Zschr. Konstitutionslehre* 9:217, 1924. (26) Wintrobe, M.M., Samter, M., and Lisco, H., *Bull. Johns Hopkins Hosp.* 64:399, 1939. (27) Oglesby, W.T., et al., *Iowa State College J. Sci.* 6:227, 1932. (30) Cartwright, G.E., (32) Fritsch, G., *Pflügers* (34) Werner, H., quoted Riddle, O., and Braucher, *Biol. Chem.* 148:261, 1943. (37) Kisch, B., *Exp. Med. Surg.* 7:318, 1949. (38) Steamer, S.P., *Argonne National Lab.*, #108, p.42, 1948. (39) Osgood, E.E., *Arch. Int. Med.* 55:849, 1935. (40) Bethell, F.H., "Clinical Laboratory Diagnosis and Essentials of Hematology," The Edwards Letter Shop, Ann Arbor, Michigan, 1948.

#### 44. BLOOD ERYTHROCYTE VALUES: FISH

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 1	d	9 A-F	7d	15 F	d
1 AB D	4d	10 A-F	3b	16 AB DE	2ad
1 C	5a	11 A-F	3ab	16 C	d
2, 3 A-D F	1ac	12 AB DE	2ad	16 F	6d
4-6 ABCDEF	3b	12 F	d	17-22 ABCDEF	3b
7 A-F	7d	13, 14 A-C	5a	23 AB DE	2d
8 A-F	3b	15 AB DE	2ad	24, 25 A-D F	1ac

Contributors: (a) Hart, J.S. (b) Kisch, B. (c) McCutcheon, F.H. (d) Root, R.W.

References: (1) Field, J.B., et al., *J. Biol. Chem.* 148:261, 1943. (2) Kisch, B., *Exp. Med. Surg.* 7:318, 1949. (3) Kisch, B., *Exp. Med. Surg.* 9:125, 1951. (4) McCay, C.M., N.Y. State Cons. Dept. Suppl. 16th Ann. Report, p. 140, 1928. (5) McCay, C.M., *J. Biol. Chem.* 90:497, 1931. (6) Root, R.W., *Biol. Bull.* 51:427, 1931. (7) Wintrobe, M.M., *Folia haematol.* 51:32, 1933.

#### 45. ERYTHROCYTE DIAMETERS: MAMMALS

Data Coordinates and Footnotes	Contributors and References
1 AB	2b
2-116 AB & Fn 1,2	1c, a

Contributors: (a) Handley, C. O., Jr.: contributed common and accepted scientific names. (b) Osgood, E. E. (c) Ponder, E.

References: (1) Ponder, E., "Hemolysis and Related Phenomena," Grune & Stratton, N. Y., 1948. Quoted from Gulliver, G., *Proc. Zool. Soc.*, London, p. 474, 1875. (2) Donelson, E. G., Leitchearing, J. M., and Wall, L. M., *Am. J. Physiol.* 128:382, 1940.

# 46. BLOOD ERYTHROCYTE VALUES IN FETUS, NEWBORN, AND IN ADULT FEMALE: MAMMALS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-5BC, 14-17BC, 28-29BC, 38-41BC, 50-51BC	11ad	2-9FG, 12FG, 14-21FG, 24FG, 25-33FG, 38FG, 39-43FG, 48FG, 50-57FG, 60FG	9ad	7-9NO, 19N, 20NO, 10-12NO, 22, 23NO, 24N	5ad 13ad
10BC, 14BC, 48BC, 52BC	3ad	14H, 16H, 17-19H, 20H, 21H, 22H, 24H, 26H, 29-31H, 32H, 32H, 34H, 36H, Fh 8, 10	2ad	29-32NO, 34, 35NO, 36N, 43N, 44NO, 46, 47NO, 48N, 55N, 58NO, 59, 59NO, 60N, Fh 11, 12	6ad 13ad
12BC, 24BC, 48BC, 60BC	7bc	1-8JK, 10, 11JK, 12J, 13-20JK, 22, 23JK, 24J, 26-33JK, 36JK, 38-44JK, 48, 49JK, 48J, 49-56JK, 58, 59JK, 60J, Fh 7, 8	12ad, 13ad	3PQ, 5P, 7P, 8PQ, 15PQ, 17P, 19P, 20PQ, 39P, 41P, 43P, 44PQ, 51PQ, 53P, 55P, 56PQ, 12P, 24P, 43P, 60P	d 8ad 1ad
22BC, 36BC	10ad				
Fh 3	4				
Fh 2, 5	14ad				
5-TDE, 10DE, 12D, 17-19DE, 22DE, 24D, 28-31DE, 34DE, 34D, 41-43DE, 48DE, 49D, 53-55DE, 58DE, 60D, Fh 4	ad 12ad, 13ad				

Contributors: (a) Barron, D. H. (b) Bethell, F. H. (c) Osgood, E. E. (d) Young, I. M.

References: (1) Barcroft, J., "Researches on Prenatal Life," Blackwell, Oxford, 1947. (2) Elliott, R. H., Hall, F. G., and Huggett, A. St. G., J. Physiol. 82: 160, 1934. (3) Guest, G. M., Brown, E. W., and Wing, M., Am. J. Dis. Child. 56: 319, 1938. (4) Handbook of Biological Data, Table on Blood Erythrocyte Values: Man. (5) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (6) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (7) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (8) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (9) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (10) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (11) Kindred, J. E., and Corey, E. L., Arch. Path. 47: 200, 1943. (12) Wint, G. R., and Sherry, A. D., Am. J. A. 11: 1, 1950.

# 47. ERYTHROCYTE OXYGEN CONSUMPTION

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A CDE	8ab	8 A CDE	5ab	17 AB DE	4ab
2 A-E	2ab	9-12 ABCDE	7a	18 A CDE	8ab
3 A CDE	1ab	13 A-E	3ab	19 AB DE	6ab
4 A CDE	5ab	14 A-E	8ab	20 A CDE	5ab
5, 6 A-E	7ab	15 AB DE	6ab	21, 22 A-E	7a
7 AB DE	6ab	16 A-E	7ab	23 A-E	5ab
				24, 25, 26 A-E	7a

Contributors: (a) Hunter, F. R. (b) Ponder, E.

References: (1) Darnall, W. D., and Barron, E. S. G., J. Exp. Med. 48: 107, 1928. (2) Darnall, W. D., and Barron, E. S. G., J. Exp. Med. 48: 107, 1928. (3) Darnall, W. D., and Barron, E. S. G., J. Exp. Med. 48: 107, 1928. (4) Darnall, W. D., and Barron, E. S. G., J. Exp. Med. 48: 107, 1928. (5) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (6) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (7) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (8) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (9) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (10) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (11) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (12) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (13) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (14) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (15) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (16) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (17) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (18) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (19) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (20) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (21) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (22) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (23) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (24) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (25) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (26) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (27) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (28) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (29) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (30) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (31) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (32) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (33) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (34) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (35) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (36) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (37) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (38) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (39) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (40) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (41) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (42) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (43) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (44) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (45) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (46) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (47) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (48) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (49) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (50) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (51) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (52) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (53) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (54) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (55) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (56) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (57) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (58) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (59) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (60) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (61) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (62) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (63) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (64) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (65) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (66) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (67) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (68) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (69) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (70) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (71) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (72) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (73) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (74) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (75) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (76) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (77) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (78) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (79) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (80) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (81) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (82) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (83) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (84) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (85) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (86) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (87) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (88) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (89) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (90) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (91) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (92) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (93) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (94) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (95) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (96) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (97) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (98) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (99) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909. (100) Ramsey, R., and Warren, C., J. Physiol. 31: 413, 1909.

# 48. ERYTHROCYTE CARBOHYDRATE METABOLISM

Data contributed by Spicer, S. S., from Spicer, S. S., and Clark, A. M., J. Biol. Chem. 173: 987, 1949, and Spicer, S. S., J. Pharm. Exp. Therap. 99: 185, 1950.

49. BLOOD LEUKOCYTE VALUES, BIRTH TO MATURITY: MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 3	7,8,9	#2-46 F	(##1-45 B and ##1-45 F)	#1-45 N	44
Fn 6	7	#1-45 H	39 and (##1-45 D)	#2-45 N	(##1-45 B and ##1-45N)
1-46 B	Sum of (1-46 D J L N P) checked against I	#2-46 H	(##1-45 B and ##1-45 I)	#1-45 O	46
#1-45 C	28	#1-45 J	40	#2-46 O	(##1-45 B and ##1-45 O)
Fn 8	34	#2-46 J	(##1-45 B and ##1-45 J)	27-38 G	25 through 27
#1-45 D	37	#1-45 K	41	44, 46 G	5
#2-46 D	(##1-45 B and ##1-45D)	#1-45 L	42	#1-45 I	(##27-45 E and ##27-45 G)
#1-45 E	38	#2-46 L	(##1-45 B and ##1-45 L)	#28-42 I	25 through 27
2,4,6 E	7,8	#1-45 M	43	44, 46 I	25 through 27,5
8 E	8				
#1-45 F	39 and (##1-45 D)				

Note # Even numbers only  
\* Odd numbers only

Note # Even numbers only  
# Odd numbers only

Note: \* quoted by reference 28  
 \*\* quoted by reference 24  
 \*\*\* quoted by reference 29  
 @ quoted by reference 7  
 @@ quoted by reference 30  
 @@@ quoted by reference 35

[illegible]

# 50. BLOOD LEUKOCYTE VALUES AT ALTITUDE MAN

Data contributed by Diggs, L. W., from Hurtado, A., Merino, C., and Delgado, E., Arch. Int. Med. 25 284, 1945

# 51. BLOOD LEUKOCYTE VALUES IN PREGNANCY AND POSTPARTUM MAN

Data contributed by Bethell, F. H., from Sturgis, C. C., and Bethell, F. H., Physiol. Rev. 23 279, 1943.

# 52 BLOOD LEUKOCYTE COUNTS, ABSOLUTE AND DIFFERENTIAL: MAMMALS, LABORATORY

Data Coordinates	Contributors and References	Data Coordinates	Contributors and References	Data Coordinates	Contributors and References
1A	Smoothed av. of means fr 1 through 14	7A	Smoothed av. of means fr. 5,6,7,8,10,11,13, 26 through 91,92b	12D	49,50,53,54,55,60,61,62,63, 71,72,73,76,89,90,92b,94
2B	Calc. & smoothed fr. (2D x 1B)	8B	Calc. & smoothed fr. (8D x 7B)	8E	Smoothed av. of means fr. 3,4,8,33,34,35,36,39,44,50, 53,54,55,60,62,63,71,72, 73,76,89,90,92b,94
3B	Calc. & smoothed fr. (3D x 1B)	9B	Calc. & smoothed fr. (9D x 7B)	9E	Smoothed av. of ranges fr. 3,4,5,30,33,35,39,43,44,60, 61,66,72,76,78,92b
4B	Calc. & smoothed fr. (4D x 1B)	10B	Calc. & smoothed fr. (10D x 7B)	10E	Smoothed av. of ranges fr. 3,4,5,30,33,35,38,39,44,60, 61,66,72,76,78,92b
5B	Calc. & smoothed fr. (5D x 1B)	11B	Calc. & smoothed fr. (11D x 7B)	11E	Smoothed av. of ranges fr. 3,4,33,35,39,44,60,61,72, 76,78,92b
6B	Calc. & smoothed fr. (6D x 1B)	12B	Calc. & smoothed fr. (12D x 7B)	12E	Smoothed av. of ranges fr. 3,4,33,35,39,44,60,61,72, 76,78,92b
1C	Smoothed av. of ranges fr. 1,2,3,4,5,8,10,12,15,16,17, 18,19,20,21	7C	Smoothed av. of ranges fr. 4,5,8,10,11,26,27,30,32,33, 34,35,36,39,40,41,42,43,44, 46,47,48,49,50,51,52,53,54, 55,58,59,60,61,64,65,66,72, 73,74,75,76,78,83,84,82b, 93	Rabbit 13B	Smoothed av. of means fr. 3,4,5,7,8,10,11,23,30,33, 37,47,49,51,52,56,67,68,69, 72,73,74,83,94,96 through 150,152 through 159,224, 225,226
2C	Calc. & smoothed fr. (2E), (1C), (1B)	8C	Calc. & smoothed fr. (8E), (7C), (7B)	14B	Calc. & smoothed fr (14D x 13B)
3C	Calc. & smoothed fr. (3E), (1C), (1B)	9C	Calc. & smoothed fr (9E), (7C), (7B)	15B	Calc. & smoothed fr (15D x 13B)
4C	Calc. & smoothed fr (4E), (1C), (1B)	10C	Calc. & smoothed fr (10E), (7C), (7B)	16B	Calc. & smoothed fr (16D x 13B)
5C	Calc. & smoothed fr. (5E), (1C), (1B)	11C	Calc. & smoothed fr (11E), (7C), (7B)	17B	Calc. & smoothed fr. (17D x 13B)
6C	Calc. & smoothed fr. (6E), (1C), (1B)	12C	Calc. & smoothed fr (12E), (7C), (7B)	18B	Calc. & smoothed fr (18D x 13B)
2D	Smoothed av. of means fr. 1,2,4,5,6,8,11,12,17,19,22, 23,24	8D	Smoothed av. of means fr. 3,4,5,6,8,11,13,29,30,33, 34,35,36,38,39,43,44,49,50, 51,53,54,55,60,61,62,63,71, 72,73,75,76,78,83,89,90, 92b,93,94	13C	Smoothed av. of ranges fr. 3,4,5,8,11,20,33,37,47,51, 53,67,69,74,77,78,93,96, 102,103,104,105,106,108, 109,110,111,113,114,115, 116,118,119,120,121,122, 123,126,127,129,130,131, 132,135,136,137,138,140, 141,142,143,144,145,146, 147,149,151,156,157,224, 225,226
3D	Smoothed av. of means fr. 1,2,4,5,6,8,11,12,17,19,22, 23,24	9D	Smoothed av. of means fr. 3,4,5,6,8,11,13,29,30,33,34, 35,36,39,44,48,49,53,54,55, 60,61,62,63,71,72,73,76,89, 90,92b,94		
4D	Smoothed av. of means fr. 1,2,4,5,6,8,11,17,19,22, 23,24	10D	Smoothed av. of means fr. 4,8,34,35,39,44,50,53,66, 92b		
5D	Smoothed av. of means fr. 1,2,4,5,6,8,11,17,19,22, 23,24	11D	Smoothed av. of means fr. 3,4,6,29,33,34,35,36,39,44, 92b		
6D	Smoothed av. of means fr. 1,2,4,8,12,17,19,22, 23,24				
2E	Smoothed av. of ranges fr. 5,8,16,18				
3E	Smoothed av. of ranges fr. 8,16,18				
4E	Calc. fr. range fr 25				
5E	Smoothed av. of ranges fr. 5,8,16,18				
6E	Smoothed av. of ranges fr. 8,16,220				

(Continued on the next page)



Data Coordinates	Contributors and References	Data Coordinates	Contributors and References	Data Coordinates	Contributors and References
14C	Calc. & smoothed fr. (14E), (13C), (13B)	4F	Calc. & smoothed fr. (4H x 1F)		190,191,192,193,194,195, 196,197,198,199,200,201, 202,203,204,205,206,207, 208,209,210,211,212
15C	Calc. & smoothed fr. (15E), (13C), (13B)	5F	Calc. & smoothed fr. (5H x 1F)	14F	Calc. & smoothed fr. (14H x 13F)
16C	Calc. & smoothed fr. (16E), (13C), (13B)	6F	Calc. & smoothed fr. (6H x 1F)	15F	Calc. & smoothed fr. (15H x 13F)
17C	Calc. & smoothed fr. (17E), (13C), (13B)	1G	95% range calc. fr. 175a	16F	Calc. & smoothed fr. (16H x 13F)
18C	Calc. & smoothed fr. (18E), (13C), (13B)	2-6G	Smoothed av. based on ranges fr. 53,175a,181, 183,184,185	17F	Calc. & smoothed fr. (17H x 13F)
14D	Smoothed av. of means fr. 4,5,8,11,23,33,44,51,53,55, 56,93,94,96,97,100,104, 105,106,107,108,119,123, 125,130,131,134,135,136, 141,142,148,151,155,156, 160,224,227	2H	Smoothed weighted av. of means fr. 159,170,174,175a, 177,178,179,180,182	18F	Calc. & smoothed fr. (18H x 13F)
15D	Smoothed av. of means fr. 4,5,8,11,33,56,94,100,104, 112,113,116,119,123,130, 131,134,135,141,148,151, 155,156,160,224,225,227	3H	Smoothed weighted av. of means fr. 174,175,180,182	13G	Smoothed av. of ranges fr. 3,4,8,74,123,187,188,189, 190,184,200,201,202,203, 204,207,208,209,210
16D	Smoothed av. of means fr. 4,5,8,33,94,96,100,105,108, 112,113,116,119,123,148, 151,155,156,225,227	4H	Smoothed weighted av. of means fr. 170,174,180,182	14G	Calc. & smoothed fr. (14I), (13G), (13F)
17D	Smoothed av. of means fr. 3,4,5,8,23,24,33,53,55,56, 78,94,96,100,104,107,108, 112,113,119,125,130,131, 134,135,141,142,148,151, 155,156,158,159,160,224, 225,227	5H	Smoothed weighted av. of means fr. 159,170,174,175, 178,180,182	15G	Calc. & smoothed fr. (15I), (13G), (13F)
18D	Smoothed av. of means fr. 3,8,33,53,55,94,96,100,107, 112,113,119,130,131,134, 135,141,148,151,155,156, 160,224,225,227	6H	Smoothed weighted av. of means fr. 174,181,182	16G	Calc. & smoothed fr. (16I), (13G), (13F)
14E	Smoothed av. of ranges fr. 8,11,24,51,67,78,130,135, 137,145,146,161,162,163, 224	1-3I	95% range calc. fr. 183	17G	Calc. & smoothed fr. (17J), (13G), (13F)
15E	Smoothed av. of ranges fr. 3,4,5,8,11,33,78,102,108, 113,115,119,130,135,137, 162,163,224	4I	184	18G	Calc. & smoothed fr. (18I), (13G), (13F)
16E	Smoothed av. of ranges fr. 3,4,5,8,11,24,33,102,106, 108,113,115,119,130,135, 145,146,162,163	5-6I	95% range calc. fr. 183	14H	Smoothed av. of means fr. 3,5,8,23,24,55,123,163,180, 181,182,194,188,189,202, 204,205,206,208,210,211, 212,213,214
17E	Smoothed av. of ranges fr. 3,4,5,8,24,33,78,102,106, 108,113,115,119,130,137, 145,162,224	Moose		15G	Smoothed av. of means fr. 3,5,8,23,55,56,186,190,191, 192,197,198,199,201,202, 204,205,210,212,214,215, 216
18E	Smoothed av. of ranges fr. 8,24,78,101,102,106,107, 113,115,119,130,135,137, 145,146,162,224	7F	Smoothed av. of means fr. 4,5,8,55,69,164,165,166	16H	Smoothed av. of means fr. 3,5,8,23,55,56,123,163,186, 187,190,191,192,198,199, 202,204,205,210,212
19E	Smoothed av. of means fr. 159,170,171,172,173,174, 175a,176,177,178,179,180, 181	8F	Calc. & smoothed fr. (8H x 7F)	17H	Smoothed av. of means fr. 3,5,8,23,24,55,56,123,163, 186,190,191,192,194,198, 199,202,204,205,208,210, 211,212,214,229
20E	Calc. & smoothed fr. (2H x 1F)	9F	Calc. & smoothed fr. (9H x 7F)	18H	Smoothed av. of means fr. 3,8,55,56,186,190,192,198, 199,202,204,205,208,210, 212
3F	Calc. & smoothed fr. (3H x 1F)	10F	Calc. & smoothed fr. (10H x 7F)	14I	Smoothed av. of ranges fr. 3,4,5,8,56,78,187,188,189, 193,197,198,212,217,218, 219,230
		11F	Calc. & smoothed fr. (11H x 7F)	15I	Smoothed av. of ranges fr. 3,4,5,8,123,163,187,190, 193,188,201,202,212,215, 217,230
		12F	Calc. & smoothed fr. (12H x 7F)	16I	Smoothed av. of ranges fr. 3,4,5,8,78,187,188,190,193, 197,198,212,217,230
		7G	Smoothed av. of ranges fr. 4,8,164	18I	Smoothed av. of ranges fr. 3,4,8,78,187,188,190,193, 197,198,212,217,230
		8G	Calc. & smoothed fr. (8I), (7G), (7F)		
		9G	Calc. & smoothed fr. (9I), (7G), (7F)		
		10G	Calc. & smoothed fr. (10I), (7G), (7F)		
		11G	Calc. & smoothed fr. (11I), (7G), (7F)		
		12G	Calc. & smoothed fr. (12I), (7G), (7F)		
		8H	Smoothed av. of means fr. 4,5,8,55,165,167,168,169		
		9H	Smoothed av. of means fr. 4,5,8,55,167,169		
		10H	Smoothed av. of means fr. 4,5,167,169		
		11H	Smoothed av. of means fr. 4,5,8,24,55,165,167,168, 169		
		12H	Smoothed av. of means fr. 4,5,55,166,169		
		8-12I	Smoothed av. of ranges fr. 169,228		
		Guinea Pig			
		13F	Smoothed av. of means fr. 4,5,7,8,55,68,74,93,99,123, 133,165,186,187,188,189,		

Contributors. (a) Endicott, K. M., (b) Rekers, P. E.

Note:

- \* = quoted by (220)
- \*\* = quoted by (215)
- \*\*\* = quoted by (222)
- @ = quoted by (2)
- / = quoted by (223)

#### Reference:

- 6 [1] Busch and Van Bergen, [2] Sawitsky and Meyer, J. Lab. Clin. Med. 37: 1947. \* [3] Burnett, S. H., "The Clinical Pathology of the Blood of Domesticated Animals," N. Y., 1917. \* [4] Corsy, F., "Les elements figures du sang chez les animaux de laboratoire" Cobaye, Lapin, Rat, Chien, et Chat." Montpellier, 1911. \* [5] Goodall, A., J. Path. Bact. 14: 195-98, 1910. \* [6] Goodall, A., Gulland, G. L., and Paton, D. N., J. Physiol. 30 1-8, 1904. \* [7] Hayem, G., "Du sang et de ses alterations anatomiques," Paris, 1889, 1890. \* [8] Klieneberger, C., and Carl, W., "Die Blut-Morphologie der Laboratoriums-Tiere," Leipzig, 1912. \* [9] Mikrúkov, Compt. rend. Charkow Vet. Inst., 1881, Abst. Jahresber. vet. Med. 11: 33, 1891. \* [10] Drinker, C. K., Drinker, K. R., and Lund, C. C., Am. J. Physiol. 42: 1-92, 1922. \* [11] Paton, D. N., Gulland, G. L., and Fowler, J. S., J. Physiol. 28: 83-106, 1902. \* [12] Shaw, H. B., J. Path. Bact. 9, 435-42, 1903. \* [13] Sherrington, C. S., Proc. Roy. Soc. Lond. 33: 161-207, 1894. \* [14] Wells, J. B., and Sutton, J. E., Am. J. Physiol. 33: 31-f.
- \* [21]:  
\* [24]:  
Yale J.  
München, 1884; Also, Deutsches Arch. f. klin. Med. 35. 217-53, 1884. \* [27] Asher, L., and Vogel, H., Biochem. Z. 43: 388-409, 1912. \* [28] Athanasia, J., Dict. de Physiol. de Richet 3: 328-43, 1898. \* [29] Biedl, A., and von Decastello, A., Arch. f. exp. Med. u. phys. Anat. 22: 9-24, 1894.
- 32:  
\* [2]  
Arc  
Bur  
McC  
Car  
Dri

dat.  
d'ar  
Mo.  
50,  
Pat  
190  
A.,  
\* /  
Pha  
Blu  
(qui  
by

- u. Pharmacol. 58: 319-42, 1906. \* /109/ Downs, A. W., and Eddy, N. B., *Am. J. Physiol.* 51: 279-88, 1920. \* /110/ Ewing, J., *New York Med. J.* 61: 257-63, 1895. \* /111/ Freytag, F., *Arch. f. d. ges. Physiol.* 120: 517-64, 1907. \* /112/ Fritsch, G., *Arch. f. d. ges. Physiol.* 161: 78-105, 1920. \* /113/ Furno, A., *Folia haematol.* 11: 219-52, 1911. \* /114/ Gruber, G. B., *Arch. f. exp. Path. u. Chir.* 58: 1901. \* /115/ Gruber, G. B., *Arch. f. exp. Path. u. Chir.* 58: 1901. \* /117/ Heinecke, H., *Mitt. rchow's Arch. f. path. Anat.* 168: 485-500, 1902. \* /120/ Jones, C. P., *J. Path. Bact.* 16: 48-70,

- Anat. u. z. allg. Path.* 51: 576-631, 1911. \* /142/ Simon, P., and Spillman, L., *Compt. rend. Soc. de biol.* 60: 833-35, 1906. \* /143/ Taussig, O., *Arch. f. exp. Path. u. Pharmacol.* 30: 161-79, 1892. \* /144/ Webb, Williams, W. W., and Basinger, A. F., *Tr. Nat. Tuberc. Assoc.* 5: 279-307, 1910. \* /145/ Wells, C. W., *J. Infect. Dis.* 20: 219-31, 1917. \* /146/ Ziegler, K., "Experimentelle und klinische Untersuchungen über die Histogenese der myeloiden Leukaemia." Jena, 1906. \* /147/ Zinsser, H., and Tsien, E., *J. Immunol.* 2: 247-68, 1917. \* /148/ Casey, A. E., Rossahn, P. D., Hu,

(Concluded from preceding page)

\*\* /214/ Lyons and Van de Carr. \* /215/ Ople, E. L., Am. J. Med. Sci. 127, 477-83, 1904. \*\* /216/ Stäubli. \* /217/ Schilling, V., Folia haematol. 7, 225-32, 1907. \*\* /218/ Hunter. \*\* /219/ Jolly and Aouna. /220/ Scarborough, R.A.,

### 53. BLOOD LEUKOCYTE RANGES. FARM ANIMALS

Contributor (a) Craig, A.H., Jr.

### 54. MORPHOLOGY OF BLOOD LEUKOCYTES AND PLATELETS

Data contributed by Rebuck, J.W.

### 55. PROPERTIES AND FUNCTIONS OF WHITE BLOOD CELLS AND RELATED FORMS

Data contributed by Rebuck, J.W. Bibliographic references on file in the office of the Handbook of Biological Data, National Research Council, Washington, D. C.

### 56. GENEALOGY OF THE FORMED ELEMENTS OF BLOOD Monophyletic Theory, William Bloom, (1952)

Contributor (a) Bloom, W.

References (1) Heller, M., McLean, F. C., and Bloom, W., Am. J. Anat. 87, 315, 1950. (2) Bloom, W., in chapters 5 and 11 on embryogenesis of lymphocyte and monocyte, in "Handbook of Hematology," edited by Downey, H., Paul B. Hober, Inc., New York, 1938.

### 57. GENEALOGY OF THE FORMED ELEMENTS OF BLOOD Modified Neo-Unitarian (Monophyletic) Theory (of Downey), Oliver P. Jones, (1952)

Footnotes	Contributors and References
Fn 4	1a
Fn 5	2a
Fn 6	3a
Fn 7	4a

Contributor (a) Jones, O. P.

References (1) Downey, H. Monogenic Leukemia and Leukemic Refractory Erythrocytosis. In "Handbook of Hematology" edited under H. P. Hober, Inc., New York, 1938.

58. GENEALOGY OF THE FORMED ELEMENTS OF BLOOD  
Polyphyletic (Dualist) Theory, Charles A. Doan, (1952)

Contributor (a) Doan, C. A.

References (1) Doan, C. A., Cunningham, R. S., and Sabin, F. R., Experimental Studies on the Origin and Maturation of Avian and Mammalian Red Blood Cells, Contributions to Embryol., No. 83, Publication 361, Carnegie Institution of Washington, Jan. 1925, pp. 163-226. (2) "Lymphocytes and Monocytes from a 351 of Carnegie Institution of Washington Experimental Hematology", Medicinal Studies of Blood Cells, Bull. N. Y. Its Physiology and Pathology, J. Lab. Clin. Med. 49: 69-101, 1940

59. NOMENCLATURE RECOMMENDED FOR CELLS OF BLOOD AND BLOOD-FORMING ORGANS

Contributors (a) Bethell, F. H., (b) Osgood, E. E., (c) Endicott, K. M., Blood # 89, 1949.

60. BONE MARROW (STERNAL) DIFFERENTIAL CELL COUNT. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-31 B	1a	1-31 FG	5a	1-31 KL	8a
1-31 C	2a	1-31 H	6	1-30 MN	*
1-31 D	3a	1-31 J	7a	31 MN	**
1-31 E	4				

Contributors (a) Diggs, L. W. (b) Osgood, E. E.

References (1) Israels, M. C. G., "An Atlas of Bone Marrow Pathology," Wm. Heinemann, London, 1948 (2) Diggs, L. W., in "Textbook of Clinical Pathology," Parker, F. P., Editor, Williams & Wilkins Co., 1948. (3) Whitby, L. E. H., and Britton, C. J. C., "Disorders of the Blood," 6th ed., Blakiston Co., Philadelphia, 1950. (4) Leitner, S. M., "Die Intravital Knochenmarkuntersuchung," Benno Schwabe & Co., Verlag Basel, 1945. (5) Lucia, S. P., and Hunt, M. L., Am. J. Med. Sci. 213 686, 1947 (6) Berman, L., Blood 4 511, 1949. (7) Wintrobe, M. M., "Clinical Hematology," 2nd ed., Lea & Febiger, Philadelphia, 1946. (8) Vaughan, S. L., and Brockmyre, F., Blood, Spl. Iss., 1 54-59, 1947. (9) Osgood, E. E., and Seaman, A. J., Physiol. Rev. 24 46, 1944. (10) Napier, L. E., and Sen Gupta, P. C., Indian Med. Gaz. 73 1, 1938. (11) Pitts, H. H., and Packham, E., Arch. Int. Med. 64 471, 1939 (12) Young, R. H., and Osgood, E. E., Arch. Int. Med. 55 186, 1935.

\*Smoothed weighted mean and 95% range calc. by b from 11 and 12 and published in 9.

\*\*Smoothed weighted mean and 95% range calc. by b from 10 and 11 and published in 9

61. BONE MARROW (STERNAL) DIFFERENTIAL CELL COUNT. SUMMARY MAN

References (1) Scott, R. B., Quart. J. Med. 8 127, 1939. (2) Forsett, J., J. Acta Med. Scandinav. Suppl. 101 1, 1939 (3) Plum, C. M., Acta Med. Scandinav. 107 50, 1941. (4) Mendell, T. H., Ann. Int. Med. 16 1180, 1942 (5) Vogel, P., et al., Am. J. Clin. Path. 7 436, 1937 (6) Hansen, T. S., Nord. Med. (Hospitalsid) 11 2167, 1941. (7) Segerdahl, E., Acta Med. Scandinav. Suppl. 64 1, 1935 (8) Lucia, S. P., and Hunt, M. L., Am. J. Med. Sci. 213 686, 1947. (9) Jacobson, K. M., Acta Med. Scandinav. 106 428, 1941. (10) Henning, N., and Kellback, H., Ergebn. d. inn. Med. u. Kinderh. 56 372, 1939. (11) Manning, J. H., Am. J. Med. Sci. 214 469, 1947. (12) Young, R. H., and Osgood, E. E., Arch. Int. Med. 64 471, 1939 (13) Vaughan, S. L., and Brockmyre, F., Blood, Spl. Iss. 1 54-59, 1947. (14) Pitts, H. H., and Packham, E. A., Arch. Int. Med. 64 471, 1939. (15) Osgood, E. E., and Seaman, A. J., Physiol. Rev. 24:46, 1944. (16) Israels, M. C. G., "An Atlas of Bone Marrow Pathology," Wm. Heinemann, London, 1948 (17) Diggs, L. W., "Textbook of Clinical Pathology," Parker, F. P., Editor, Williams & Wilkins Co., Baltimore, 1948. (18) Whitby, L. E. H., and Britton, C. J. C., "Disorders of the Blood," 6th ed., Blakiston Co., Philadelphia, 1950. (19) Leitner, S. M., "Die Intravital Knochenmarkuntersuchung," Benno Schwabe & Co., Verlag Basel, 1945 (20) Wintrobe, M. M., "Clinical Hematology," 2nd ed., Lea & Febiger, Philadelphia, 1946 (21) Osgood, E. E., and Ashworth, C. M., "Atlas of Hematology," J. W. Stacey, San Francisco, 1937

## 62. BONE MARROW (STERNAL) DIFFERENTIAL CELL COUNT IN PREGNANCY. MAN

All data from Pitts, H. H., &amp; Packham, E. A., Arch Int. Med. 54: 471-482, 1939.

## 63. BONE MARROW (RIB) DIFFERENTIAL CELL COUNT: DOG

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-30 BCD	1	1-30 GHI	3	1-30 MNO	5
1-30 EF	2	1-30 JKL	4a		

Contributor: (a) Rekers, P. E.

References: (1) Stasney, J., & Higgins, G. M., Am. J. Med. Sci. 193:462, 1937. (2) Van Loon, E. J., J. Lab. Clin. Med. 28:1575, 1943. (3) Mulligan, R. M., Anat. Rec. 79:101, 1941. (4) Rekers, P. E., & Coulter, M., AEC, Oak Ridge, Tenn., 5-16-49--850--A2111. (5) Mulligan, R. M., Anat. Rec. 91:161, 1945.

## 64. BONE MARROW (RIB) DIFFERENTIAL CELL COUNT, SUMMARY DOG

References: (1) Stasney, J., & Higgins, G. M., Am. J. Med. Sci. 193:462, 1937. (2) Van Loon, E. J., J. Lab. Clin. Med. 28:1575, 1943. (3) Mulligan, R. M., Anat. Rec. 79:101, 1941. (4) Rekers, P. E., & Coulter, M., AEC, Oak Ridge, Tenn., 5-16-49--850--A2111. (5) Mulligan, R. M., Anat. Rec. 91:161, 1945.

## 65. MORPHOLOGY OF FORMED ELEMENTS OF BONE MARROW. MAN

Data contributed by Rebuck, J. W.

## 66. HISTOCHEMICAL PROPERTIES OF BLOOD AND BONE MARROW CELLS. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-21 B & Fn 1	13ab	1 E	11ab, 12ab	20 F	5g
22 B & Fn 16	8f, 9f	3 E	11ab, 12ab	21-22 F	12ab
1-3 C	13ab	4 E	2ab 11ab, 12ab, 15ab	1 G	17ab, 3g, 14h
5-22 C	13ab	5 E	11ab, 12ab, 16	2 G	17a, 14h
13 C	16	6 E	2ab, 11ab, 12ab, 15ab	3-4 G	17a, 3g
Fn 7	6c	7 E	11ab, 12ab	5 G	17a, 14h
Fn 8	13ab	9 E	11ab, 12ab	6-7 G	17a
1 D	11ab, 12ab, 8f	10 E	2ab, 11ab, 12ab	8 G	17a
2 D	12ab	11 E	12ab	9 G	17h
3 D	11ab	11 E & Fn 9	11	9 G	17a
4 D	12ab	13 E	11	11 G	17a
5 D	11ab, 12ab	13 E	11	11 G & Fn 13	3g, 14h
6 D	12ab, 2	13 E & Fn 8	12	12 G	3g, 14h
7 D	12ab	15-19 E	11ab, 12ab	13 G	17a
8 D	12ab	20 E	11	14 G	17a, 3g, 14h
11-12 D	11ab, 12ab	20 E & Fn 6	12	15-19 G	17a, 3g, 14h
15-16 D	12ab	22 E	8f, 15	20 G	3g
19-20 D	11ab, 12ab	1-7 F	12ab	21 G & Fn 15	6c
21 D	12ab	8-12 F	12ab	22 G	17a
22 D	8f, 10f, 16f	13 F	12ab, 16	22 G & Fn 17	7c
Fn 14	4d	14-19 F	12ab	2-21 H	1e
				22 H	8f

(Continued on the next page)

Contributors: (a) Wislocki, G. B. (b) Bloom, M. L. (c) Lillie, R. D. (d) Gomori, G. (e) Firminger, H. E. (f) Montagna, W. (g) Stowell, R. E. (h) Bacchus, H.

(Continued from preceding page)

References: (1) Armitage, F. L., J. Path. Bact. 49, 579, 1939. (2) Deane, H. W., Am. J. Anat. 80: 321, 1947. (3) Gibb, R. P., and Stowell, R. E., Blood 4: 589, 1949. (4) Gomori, G., Stain Tech. 25: 81, 1950. (5) Gomori, G., Unpublished data, 1950. (6) Lillie, R. D., Unpublished data, 1950. (7) Lillie, R. D., Anat. Rec. 108: 239, 1950. (8) Montagna, W., and Noback, C. R., Anat. Rec. 100: 535, 1948. (9) Montagna, W., and Hamilton, J. B., Anat. Rec. 108: 535, 1951. (10) Noback, C. R., and Paff, G. H., Anat. Rec. 109: 71, 1951. (11) Rabinovitch, M., and Andreucci, D., Blood 4: 580, 1949. (12) Rheinhold, J. J., and Wislocki, G. B., Blood 3: 641, 1948. (13) Rheinhold, J. J., and Wislocki, G. B., Blood 3: 79, 1950. (14) Wachstein, M., Blood 4: 54, 1949. (15) Wislocki, G. B., and Dempsey, E. W., Anat. Rec. 86: 249, 1946. (16) Wislocki, G. B., Bunting, H., and Dempsey, E. W., Anat. Rec. 98: 527, 1947. (17) Wislocki, G. B., Rheinhold, J. J., and Dempsey, E. W., Blood 4: 562, 1949.

# 67. BLOOD WATER AND SOLIDS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B-E 2 B-E 3 B-E 4 BC 5-7 B D 8 BC	a 1 2 1 3 Calc. fr. (9.10 BC)	9,10 BC 11-13 B D 14 B-E 15 BC 16 BC 17 B D	3 a a b 3	18-28 B D 29 B-E 30 B-E 31 B D 32-34 B D	j a b 4c j

Contributors: (a) Manery, J. F. (b) Allison, J. B. (c) Bacchus, H.

References: (1) Hald, P. M., Tulin, M., Danowski, T. S., Lavietes, P. H., and Peters, J. F., Am. J. Physiol. 149: 340, 1947. (2) Miller, A. T., Jr., J. Biol. Chem. 143 65, 1942. (3) Dukes, H. C., "Physiology of Domestic Animals," Comstock Publishing Co., Ithaca, N. Y., 1942. (4) Bacchus, H., Toompas, C. A., and Heifer, M. H., unpublished data, 1951.

# 68. BLOOD CARBOHYDRATES AND RELATED SUBSTANCES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,2 B-D & Fn 1,2 3,4 BCD & Fn 3,4 5 BD & Fn 5,6 6 BD & Fn 7 7,8 BC & Fn 3,4 9,10 BC 11 B-D & Fn 3,8 12,13 BC & Fn 3,4,7 14,15 B-D & Fn 9 16 B-D & Fn 5,6 17,18 B-D & Fn 3,4 19 B-D 20,21 BCD & Fn 5,10 22,23 BC & Fn 3,4 24 BCD & Fn 9 25 BCD 26,27 BC & Fn 3,4 28,29 B-D & Fn 3,6	60a 58a 45a 27a 59a 32a 7a 59a 3a 7a 59a 57a 25a 59a 6a 57a 59a 22a	30,31 B-D & Fn 3,8 32-35 BC & Fn 3,4 36 BCD & Fn 5,11 37,38 BCD & Fn 3,4 39,40 BCD & Fn 5,10 41 BCD & Fn 5,10 42,43 BCD & Fn 3,4 44 BC & Fn 12 45 BCD & Fn 5,10 46 BCD & Fn 5,11 47 BD & Fn 5,10 48 BCD & Fn 5,11 49 BCD & Fn 3 13 50-52 BCD 53 BCD 54,55 BD 56 BC & Fn 12 57,58 BC & Fn 3,4,10,14	2a 59a 8a 59a 11a 49a 58a 4a 50a 41a 43a 61a 12a 46a 36a 19a 33a 14a	59 BD & Fn 3,4,10,14 60 BD & Fn 3,4,10,14 61,62 BC & Fn 3,4,10,14 63 BD & Fn 3,4,10,14 64 BC & Fn 3,4,10,14 65,66 BCD & Fn 3,4,10,14 67 BC & Fn 4,5 68 BD 69 BD & Fn 16 70-74 BC & Fn 5,17 75-77 BD & Fn 3,13,18 78,79 BCD & Fn 10,19 80,81 BC & Fn 10,19	14a 16a 16a 15a 16a 16a 54a 27a 4a 33a 62a 62a

(Continued on the next page)

(Continued from preceding page)

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
82 BCD & Fn 10,19	63a	116 BCD & Fn 29	38a	155,156 BCD & Fn 35	34a
83 BC & Fn 10,19	63a	117,118 BC & Fn 29	38a	157 BC & Fn 35	28a
84-87 BCD & Fn 20	55a	119 BCD & Fn 31	28a	158 BCD & Fn 36	28a
88 BCD & Fn 21, 22	64a	120,121 BCD & Fn 31	9a	159-169 BC & Fn 37	29a
89 BCD & Fn 22	53a	122 BC & Fn 22	47a	170-174 BC & Fn 38	47a
90-93 BCD & Fn 22	55a	123 BC & Fn 30	28a	175-177 BC & Fn 39	26a
94 BCD & Fn 22	56a	124 BC & Fn 33	47a	178 BCD & Fn 40	24a
95 BCD & Fn 23, 24, 25	21a	125 BC & Fn 30	28a	179,180 BCD & Fn 39	5a
96,97 BCD & Fn 23,24	20a	126,127 BC & Fn 33	47a	181 BCD & Fn 39	44a
98 BC & Fn 23,24	21a	128 BCD & Fn 30	28a	182 BC & Fn 39	18a
99 BCD & Fn 23, 24	20a	129,130 BC & Fn 33	47a	183 BCD & Fn 39	52a
100 BCD & Fn 26, 27	48a	131 BC & Fn 30	28a	184 BC & Fn 41	10a
101,102 BC & Fn 26	19a	132 BC & Fn 33	47a	185 BCD & Fn 42	51a
103 BCD & Fn 26	13a	133 BCD & Fn 30	28a	186-189 BC & Fn 42	51a
104 BC & Fn 26	13a	134 BC & Fn 33	47a	189 BCD & Fn 43	17a
105,106 BCD & Fn 26	13a	135 BC & Fn 30	28a	190 BCD & Fn 43	6a
107 BCD	13a	136 BC & Fn 33	47a	191 BCD & Fn 43	66a
108 BCD & Fn 28	Calc. fr (106,109 BCD)	137 BC & Fn 30	28a	192-194 BC & Fn 43	35a
109 BCD & Fn 28	38a	138,139 BC & Fn 33	47a	195 BC & Fn 41	10a
110 BCD & Fn 28	37a	140 BC & Fn 34	1a	196 BC & Fn 44	30a
111,112 BC & Fn 28	38a	141 BC & Fn 30	47a	197,198 BCD & Fn 45	42a
113 BCD & Fn 29	38a	142 BC & Fn 33	28a	199 B D & Fn 45	42a
114 BC & Fn 29	37a	143 BC & Fn 30	47a	200,201 BCD & Fn 46	63a
115 BC & Fn 29	38a	144,145 BC & Fn 33	1a	202 BCD & Fn 47	24a
	40a	146-148 BC & Fn 34	47a	203 BC & Fn 48	39a
		149,150 BC & Fn 33		204 BC	31a
		151-154 BC & Fn 34			

Contributors (a) Mirsky, I. A.

## References

- /1/ Albaum, H. C., and Lipschitz, R. Arch. Biochem. 27 102, 1950. /2/ Anderson, D. Austral. J. Exper. Biol. Med. Sci. 15 17, 1937. /3/ Ayar, M. A. R., and Nayar, K. N. G., Ind. Vet. J. 17 259, 1941. (Quoted by (49)). /4/ Bacon, J. S. D., and Bell, D. J. Biochem. J. 42 397, 1948. /5/ Balas, A., Bugnard, L., and Rogeon, J. Bull. Soc. Chem. Biol. 20 51, 1938. /6/ Allibone, E. C., and Finch, E. Arch. Dis. Childhood 21 165, 1946. /7/ de Bodo, R. C., Co. Tui, F. W., & Biagaglia, A. E. J. Pharm. Exp. Therap. 61 48, 1937. /8/ Broth-Kahn, R. H., Simkin, B., and Mirsky, I. A., Arch. Biochem. 25 157, 1950. /9/ Buell, M. V. J. Biol. Chem. 103 273, 1935. /10/ Cavallini, D., Frontali, A., and Toschi, G., Nature 164 792, 1949. /11/ Cole, V. V., and Harned, B. K., Endocrinology 22 318, 1933. /12/ Coulson, R. A., Hernandez, T., and Brada, F., Proc. Soc. Exp. Biol. Med. 73 201, 1950. /13/ Deleham, W. B., and Dierker, M., J. Biol. Chem. 132 484, 1939. /14/ Florkin, M., Biochemical Soc. Chem. Biol. 19 890, 1937. /15/ Florkin, M., and Dierker, M., J. Biol. Chem. 147 415, 1943. /16/ Friedemann, T. E., and Kendall, A. L., J. Biol. Chem. 82 23, 1929. /17/ Friedemann, T. E., and Bielechowski, M., J. Path. Bact. 61 101, 1949. /18/ Green, H. B., and Stoner, H. B., J. Biol. Chem. 82 23, 1929. /19/ Goss, S. K., J. Chosen Med. Assoc. 29 1925, 1939. (Chem. Abstr. 24 1404, 1940.) /20/ Green, H. N., Stoner, H. B., and Wilson, J. W., J. Biol. Chem. 157 461, 1945. /21/ Mercer, S. E., and Sandcock, B. K., Proc. Soc. Exp. Biol. Med. 130 1225, 1949. /22/ Mirskand, W. A., Hadley, M. F., Mercer, S. E., and Sandcock, B. K., Proc. Soc. Exp. Biol. Med. 130 1225, 1949. /23/ Hubbard, R. S., and Brock, R. J., J. Biol. Chem. 110 411, 1935. /24/ Hummel, J. P., J. Biol. Chem. 135 177, 1940. /25/ Johnson, D. D., Endocrinology 45 133, 1950. /26/ Johnson, R. E., Edwards, H. T., Dill, D. B., and Wilson, J. W., J. Biol. Chem. 157 461, 1945. /27/ Karvonen, M. J. Acta Paediatr. 27 68, 1949. (Chem. Abstr. 44 1953, 1950.) /28/ Kerr, S. E., and Daoud, L. J. Biol. Chem. 105 301, 1935. /29/ Klein, J. R., 1949. (Chem. Abstr. 44 1953, 1950.) /30/ Krebs, H. A., Biochem. J. 32 108, 1938. /31/ Krebs, H. A., Ann. Rev. Biochem. 18 409, 1949. /32/ Lazarewicz, G. I., Uchenyey Zapiski Kazan Gosudarst. Zootekhn. Inst. Im. Daumana 50 3, 1939. (Chem. Abstr. 35 3236, 1942.) /33/ Levenhock, C., Nature 160 465, 1947. /34/ Levitas, N., Robinson, J., Rosen, F., Huff, E., and Peritzweig, W. A. J. Biol. Chem. 167 169, 1947. /35/ Lu, G. D., Biochem. J. 33 774, 1939. /36/ Lundberg, E., and Thyssell-Lundberg, S., Acta Med. Scand. Suppl. 170 513, 1946. /37/ Mandel, P., and Metals, P., Compt. rend. Soc. Biol. 142 241, 1948. /38/ Mandel, P., and Metals, P., Compt. rend. Soc. Biol. 142 1022, 1948. /39/ Marshall, L. M., Orten, J. M., and Smith, A. H., J. Biol. Chem. 172 1127, 1949. /40/ Metals, P., Compt. rend. Soc. Biol. 144 277, 1950. /41/ Mirsky, I. A., Proc. Soc. Exp. Biol. Med. 32 35, 1943. /42/ Natelson, S., Pincus, J., and Lugovoy, J. K. J. Clin. Invest. 27 446, 1948. /43/ Nelson, N., Elgart, S., and Mirsky, I. A., Endocrinology 31

(Continued on the next page)



(Continued from preceding page)

118, 1942. /44/ Newman, E. V., Am. J. Physiol. 122: 359, 1938. /45/ Pedersen, J., Acta Paediatrica, Suppl. 71: 201, 1948. /46/ Prado, J. L., Mem. Inst. Butantan, 19: 59, 1946 (Chem. Abs. 41: 5988, 1947). /47/ Rappoport, S., and Guest, C. M., J. Biol. Chem. 138: 269, 1941. /48/ Ratish, M. D., and Bullock, J. G. M., Arch. Biochem. 2: 381, 1943. /49/ Reid, R. L., Austral. J. Agric. Res. 1: 182, 1950. /50/ Rodbard, S., Am. J. Physiol. 150: 67, 1947. /51/ Rosenthal, S. M., J. Biol. Chem. 179: 123, 1949. /52/ Seibert, F. B., and Alno, J., Biochem. 18: 279, 1948. /53/ Shetler, 69: 507, 1948. /54/ Somogyi, M., J. Biol. Chem. 179: 123, 1949.

## 69. BLOOD LIPIDS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B-D & Fn 3	Calc. fr. (2,3 B-D)	45 B-D	4	77 B-D	Calc. fr. (77,78B-D)
2 B-D & Fn 1, 2	1a	46 B-D	Calc. fr. (47,48B-D)	78 B-D	4
3-7 BCD	2a	47 B-D	6	79, 80 B-D	3a
8 B-D	b	48 B-D	Calc. fr. (25B-D)	81 B-D	2, 3a
8-11 BCD	2a	49 B-D	and Fn 5	82 B-D	b
12, 13 B-D	1a	50 B-D	b	83-85 BCD	b
14-18 BCD	2a	51 B-D	10	86 B-D	2a, 2
19 B-D	Calc. fr. (20,21B-D)	52 B-D	b	87 B-D	b
20 B-D	4	53 B-D	Calc. fr. (52,53B-D)	88 B-D	b
21, 22 B-D	2a	54 B-D	4	89 B-D	Calc. fr. (91,92B-D)
23 B-D	Calc. fr. (24,25B-D)	55 B-D	b	90 B-D	4
24 B-D & Fn 4	5a	56 B-D	Calc. fr. (56,57BC)	91 B-D	2, 2a
26-28 BCD	2a	57 BC	6	92, 93 B-D	3, 3a
29 BC	b	58-60 BC	Calc. fr. (26B-D)	94 B-D	a
30, 31 B-D	2a	61 B-D	and Fn 5	95 B-D	2, 2a
32 B-D	Calc. fr. (33,34B-D)	62 B-D	b	96-99 BCD	b
33 B-D	4	63 B-D	Calc. fr. (82,83B-D)	100 B-D	2, 2a
34, 35 B-D	2a	64 B-D	4	101 B-D	2, 2a
36 B-D	9	65 B-D	5	102 BC	Calc. fr. (104,105 B-D)
37 B-D	Calc. fr. (38,39B-D)	66-70 BCD	3a, 3	103 B-D	4
38 B-D	4	71, 72 B-D	2a	104 BC	2, 2a
39 B-D & Fn 5	Calc. fr. (25 B-D)	73 BC	b	105, 106 B-D	107-118BCD & Fn 7
40 B-D	11	74 B-D	2a	119-125 BCD	2a
41 B-D	b	75, 76 B-D & Fn 5	b	126-128 BCD	a
42 B-D	2a		7, 7a		
43, 44 B-D	b				

Contributors: (a) Best, C. H. (b) Handler, P. (c) Kirk, J. E.

(3) Brun,  
des, F. F.  
(7) Lea,  
G.,  
Jott, J.

## 70. BLOOD PROTEINS. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B	Sum of (2B, 3B), (4B)	2 FG	g	5 FG	Av. of means fr. 1bd, 2g, 3bd, 4bd, 5bd, 6g
1 D	Sum of (2D, 3D)	3 B	Calc. fr. (3D)		Calc. fr. (8F)
1 F	Sum of (3F, 8F)	3 D	7	6 B	12f
2 B	Calc. fr. 9a, c, 10h, 8e	4 B	Calc. fr. (4F)	6 FG	8e, 11e, 9a, c, 10h, 8e
2 DE	Calc. fr. 9a, 8e	4 F	Sum of (5F, 8F)	Fn 1	
		5 B	Calc. fr. (5F)		

(Continued on the next page)

Contributors: (a) Bethell, F. H. (b) Diggs, L. W. (c) Drabkin, D. L. (d) Laki, K. (e) Osgood, E. E. (f) Peters, J. P. (g) Stefanini, M. (h) Wintrobe, M. M.

(Continued from preceding page)

References: (1) Gilligan, D. R. and Ernestine, A. C., *Am. J. Med. Sci.* **187**: 552, 1934. (2) Foster, D. P. and Whipple, G. H., *Am. J. Physiol.* **58**: 355, 1922. (3) Gram, H. C., *Acta med. scand.* **58**: 107, 1922. (4) Ham, T. H. and Curtis, F., *Medicine* **17**: 413, 1938. (5) Starlinger, W. & Winands, E., *Ztschr. f. ges. exper. Med.* **60**: 130, 1924. (6) Stefanini, M., *Arch. Sci. Med.* **69**: 177, 1940. (7) Williams, H. H., et al., *Quart. Rev. Biol.* **16**: 80, 1941. (8) Osgood, E. E., *Arch. Int. Med.* **59**: 849, 1935. (9) Bethell, F. H., "Clinical Laboratory Diagnosis and Essentials of Hematology," The Edwards Letter Shop, Ann Arbor, Mich. 1940. (10) Wintrobe, M. M., "Clinical Hematology," Year Book Publishers, 1944. (11) Ponder, E., p. 597 in "Medical Physics," Otto Glasser, Editor, Chicago 1947. (12) Milne, J., *J. Biol. Chem.* **189**: 593, 1947.

## 71. PLASMA PROTEINS MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-13 B 1 C 1-11 E 1 F 1-9 H 1 I 1-9 K	Calc. fr. (1-13C) Sum of (2C, 3C) Calc. fr. (1-11F) Sum of (2F, 3F) Calc. fr. (1-9I) Sum of (2I, 3I) Calc. fr. (1-9L)	1-9 L & Fb 3 2 CD FG  2 IL 3-13 CD 3-11 FG	2b Av. of means fr. 3ae, 4g, 5seg. 6ae, 6ac, 8g  2b 1c 7f	3l 4-9 LJ 14 C 14 F 14 LJ 14 L Fn 1 Fn 2	10d Calc. fr. 10d Calc. fr. (14C 5C) Calc. fr. (4F 5F) 10d Calc. fr. (4L 5L) 11 12,7

Contributors: (a) Diggs, L. W. (b) Edsall, J. T. (c) Gutman, A. B. (d) Keys, A. (e) Laki, K. (f) Peters, J. P. (g) Stefanini, M.

References: (1) Gutman, A. B., Moore, D. H., Gutman, E. B., McClellan, V., and Kabat, E. A., *J. Clin. Invest.* **20**: 755, 1943. (2) Armstrong, S. H., Jr., Budka, M. J. E., and Morrison, K. C., *J. Am. Chem. Soc.* **69**: 418, 1947. (3) Gilligan, D. R. and Ernestine, A. C., *Am. J. Med. Sci.* **187**: 552, 1934. (4) Foster, D. P. and Whipple, G. H., *Am. J. Physiol.* **58**: 365, 1922. (5) Gram, H. C., *Acta Med. Scand.* **58**: 107, 1922. (6) Ham, T. H. and Curtis, F., *Medicine* **17**: 413, 1938. (7) Milne, J., *J. Biol. Chem.* **189**: 595, 1947. (8) Starlinger, W. and Winands, E., *Ztschr. f. ges. exper. Med.* **60**: 130, 1924. (9) Stefanini, M., *Arch. Sci. Med.* **69**: 177, 1940. (10) Taylor, H. L., Mickleisen, O., and Keys, A., *J. Clin. Invest.* **28**: 273, 1949. (11) Howe, P. E., *J. Biol. Chem.* **49**: 93, 1921. (12) Major, C. L. H., *J. Biol. Chem.* **169**: 587, 1947.

## 72. PLASMA PROTEINS. ANIMALS OTHER THAN MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 F H J *1 G I K 2 F H J *2 G *3 B 3 C 3 D 3 F ***3 H J *4 B *4 C *4 D *4 E 4 F	1d 1d 2c 3 2c, 4b 4b 2c 2c (3B) minus (3D) 2c a, 2, 5, 6, 7, 8 9 2c, 10c 10f (4B) minus (4D)	+4 G *4 H J *4 I K 5 B 5 C ***5 D ***5 H J 6 B 6 D F H J 7 D 7 E ***7 F ***7 H J	6 2c a (5D) plus (5F) 2c *9 G *9 C *10 F *10 H J 11 B 11 D *11 E 11 F H J *11 G I K 12 B D F H J 13 F H J	8 B 8 DE ***8 F 9 F H J *9 C *10 F *10 H J 11 B 11 D *11 E 11 F H J *11 G I K 12 B D F H J 13 F H J	(8D) plus (8F) f 8 12d 13d 2c, 13d 2c, 13d (11D) plus (11F) 14 14 15de 15de 2c 2c

Note:

- \*Calculated from standard deviation from reference(s) indicated.
- \*\*Average of means from reference(s) indicated.
- \*\*\*Calculated from A/G ratio from reference(s) indicated.
- \*\*\*\*Range interval from reference(s) indicated.
- \*\*\*\*\*Calculated from average A/G ratio from reference(s) indicated.

(Continued on the next page)

Contributors. (a) Allison, J. B. (b) Brody, S. (c) Howe, P. E. (d) Leatham, J. H. (e) Seibert, F. B. (f) Stefanini, M.

(Continued from preceding page)

- References: (1) Levin, L., Leatham, J. H., and Crafts, R. C., *Am. J. Physiol.* **136**, 776, 1942. (2) Howe, P. E., *Physiol. Rev.* **5**, 439, 1925. (3) Herrmann, G. R., *Proc. Soc. Exp. Biol. Med.* **61**: 229, 1946. (4) Brody, S., et al., *Univ. Missouri Agr. Exp. Sta. Res. Bull.* **433**, 1949. (5) Elman, R., Sachar, L. A., Horvitz, A., and Wolf, H., *J. Lab. and Clin. Med.* **27**: 1183, 1942. (6) Little, J. M., and Dameron, J. T., *Am. J. Physiol.* **140**, 636, 1945. (7) Danowski, T. S., Winkler, A. W., and Elkinton, J. R., *J. Clin. Invest.* **28**, 887, 1947. (8) Drinker, C. K., and Voffey, J. M., "Lymphatics, Lymph, and Lymphoid Tissue," Harvard University Press, Cambridge, Mass., 1941. (9) Helm, J. W., *Am. J. Physiol.* **103**, 553, 1933. (10) Foster, D. F., and Whipple, G. H., *Am. J. Physiol.* **58**, 365, 1922. (11) Frederick, *Récherches sur la coagulation du sang*, Gand, Paris, Leipzig, 1878. (12) Dougherty, T. F., and White, A., *Endocrinology* **33**: 370, 1946. (13) White, A., and Dougherty, T. F., *Endocrinology* **33**: 207, 1945. (14) Henriques, S. B., Henriques, O. B., and Selye, H., *Proc. Soc. Exp. Biol. Med.* **71**: 82, 1949. (15) Moore, D. H., Levin, L., and Smelser, G. K., *J. Biol. Chem.* **157**, 723, 1945.

### 73. PLASMA PROTEINS, CERTAIN PROPERTIES AND REACTIONS. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-28AB 7-11,17-25C 2-14,19-26,28D	c c, 1, 2 1	1-10,17-21,25E 26E Fn 1 Fn 2,3	c 3b, 4b, 5b, 6b, 7b a

Contributors: (a) Edsall, J. T. (b) Gurd, F. R. N. and Brown, R. K. (c) Strong, L. E.

- References: (1) Cohn, E. J., et al., *J. Am. Chem. Soc.* **72**, 465, 1950. (2) Krebs, H. A., *Ann. Rev. Biochem.* **19**, 409, 1950. (3) Shinowara, G. Y., Jones, L. M., and Reinhard, H. L., *J. Biol. Chem.* **142**, 921, 1942. (4) Somogyi, M., *J. Biol. Chem.* **125**: 399, 1938. (5) Ambard, L., *Bull. Soc. Clin. Biol.* **3**: 51, 1921. (6) Adams, D. H., and Whittaker, V. P., *Biochem. J.* **44**, 67, 1949. (7) Talalay, P., Fishman, W. H., and Huggins, C., *J. Biol. Chem.* **186**, 757, 1946.

### 74. PLASMA PROTEINS, PHYSICAL PROPERTIES. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B 2,3 B 4,5 B 6 B 7 B 8,9,11-16 B 10 B 17 B 18 B 19-25 B 26 B 27 B 28,29 B 30-32 B 2 C 3 C 4,5 C 6 C 7 C	3 3:2 quoted by 4a 3:2 quoted by 3 & by 4a 3 3:2 quoted by 3 2 2 quoted by 3 2 quoted by 3 & by 4a 3 1 3 3,2b 2,3 2b 3	8,9 C 11,12 C 13 C 14 C 17 C 18 C 20 C 21 C 22 C 26,27 C 28 C 29 C 30-32 C 2 D 3 D 4-10 D 11,12 D 17,18 D 20-22 D 26,27 D 29 D	1 1 1,3 1 1,3 1 1,3 8 quoted by 1 1 10 quoted by 1 1,3 1 5 quoted by 1 1 1,3 1 3 1 1,3 1 1,3 1	32 D 3 E 4,5 E 7 E 18 E 26,27 E 3 F 4,5,7-9,17,18,20F 21 F 26 F 27,30,31 F 32 F 4,5 G 7,18,26,27 G 3 H 4 H 7,18,26,27 H 3,4,18,26,27 I 3,4,7,18,26,27 JK Fn 2	1 1 quoted by 4a 2 quoted by 4a 2 quoted by 4a 2 quoted by 3 & by 4a 2 quoted by 3 2 quoted by 3 1 6 quoted by 1 7 quoted by 1 1 11a 2 quoted by 4a 2 quoted by 3 2 quoted by 4a 2 quoted by 3 & by 4a 2 quoted by 3 2 quoted by 4a 2 quoted by 4a 11a

Contributors (a) Edsall, J. T. (b) Strong, L. E.

- References: (1) Cohn, E. J., et al., *J. Am. Chem. Soc.* **72**, 465, 1950. (2) Oncley, J. L., Scatchard, G., and Brown, A. J., *Phys. Colloid. Chem.* **51**, 184, 1947. (3) Cohn, E. J., *Experientia*, **3**, 184, 1947. (4) Brand, E., and Edsall, J. T., *Ann. Rev. Biochem.* **16**, 223, 1947. (5) Morrison, P. R., Edsall, J. T., and Miller, S. G., *J. Am. Chem. Soc.* **70**, 3101, 1948. (6) Surgenor, D. M., et al., *J. Am. Chem. Soc.* **71**, 1223, 1949. (7) Cohn, E. J., *Bull. N.Y. Acad. Med.* **72**, 639, 1939. (8) Surgenor, D. M., and Ellis, D., *Acta Chem. Scand.* **1**, 944, 1947. (9) Holmberg, C. G., and Laurell, C. B., *Abstracts First Internat. Cong. Biochem. Camb., Eng.* No. 291-B, *Acta Chem. Scand.* **1**, 944, 1947. (10) Hughes, W. L., Jr., *Spring Harbor Symposia Quart. Biol.* **14**, 79, 1949. (11) Schmid, K. J., *J. Am. Chem. Soc.* **72**, 2816, 1950.

### 75. BLOOD FREE AMINO ACIDS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B-H & Fn 4	6a, c	26 B GH	d, l	51 B GH	d, l
2 B GH	9e	27 B-H	21c	52 B H	3f
3 B-D	10a	28 B-D GH	12a	53 B GH	9e
4 B-H	11ac	29 B GH	9e	54 B GH	21c
5 B-H	11a, 12a	30 B GH	d, l	55 B-H	21c
6 B GH	4e	31 B-H	21c	56 B-D GH	12a
7 B GH	1b	32 B-D GH	12a	57 B GH	9e
8 B H	3f	33 B GH	9c	58 B GH	d, l
9 10 B H	8a, f	34 B GH	d, l	59 B-H	1bac
11 BC E G	13a	35 B-GH	12a	60 B GH	17a
12 B-H	21c	36 B GH	12ad	61 B-D GH	12a
13 B GH	12a	37 B GH	9e	62 B G	19g
14 B G	13a	38 B GH	d, l	63 B GH	18a, 9e
15 B H	14a	39 B-H	21c	64 B GH	18a
16 B H	14a	40 B GH	12a	65 B-D GH	d, l
17 B K	14a	41 D GH	20g	66 B GH	18a
18 B GH & Fn 7, 8	15a, f	42 B GH	9e	67 B-G	21c
19 B G	2	43 B GH	d, l	68 B-D GH	12a
20 B G	8a	44 B-H	21c	69 B GH	20g
21 B GH	9e	45 B-D GH	12a	70 B GH	9e
22 B E-H	6a	46 B GH	9e	71 B GH	1d
23 B D	4f, 5f, 6a, 7f	47 B GH	d, l	72 B-H & Fn 20	d, l
23 B G	6a	48 B-H	21c	73 B-D GH	12a
24 B GH	9e	49 B-D GH	11a, 12a	74 B GH	9e
25 B G	6a	50 B GH	9c	75 B GH	d, l

Contributors (a) Deerstecher, E., Jr., and Spangler, S (b) Williams, J. N., Jr., (c) Bergöim, O. (d) Hier, S. W. (e) Steele, B. F. (f) Hamilton, P. B. (g) Schweigert, B. S.

References: (1) Henderson, L. M., et al., *J. Biol. Chem.* **177** 815, 1949 (2) Brassman et al. *J. Biol. Chem.* **217**, 1948, (3) Steele, B. F., Reynolds, M. C., et al., *J. Biol. Chem.* **168** 1, 1947, (4) Lynch, E. L., *J. Biol. Chem.* **168** 1, 1947, (5) Gutman, G. E., Landwehr, G., *J. Biol. Chem.* **154** 643, 1944, (6) Steele, L. (10) von Euler, H., *Acta Chem. Scand.* **10** 46, 1946, (11) Sutton, T. S., and Lill, G. C., *J. Dairy Sci.* **27** 181, 1948 (12) Schweigert, B. S., et al., *J. Biol. Chem.* **164** 213, 1946 (13) Schweigert, B. S., Pearson, P. B., and Wickersham, M. C., *Arch. Biochem.* **12** 139, 1947, (14) Schweigert, B. S., unpublished data (15) Johnson, C. A., and Bergman, O., *J. Biol. Chem.* **188** 883, 1951

## 76

Data Coordinates and Footnotes	Contributors and Dates			
1 B-D				
2 B-D				
3 B-D				
4-6 BCD & Fn 6, 7, 8				
7 BC	10 BCD & Fn 12	14b, 15b	32-33 BC	5
8 B-D	13, 14		34 BC	6
9, 10 B-D & Fn 9, 10	10c 25 B-D	2	35 B-D	7
	14b, 15b 26 B-D	3	36 BC	1

Contributors (a) Beerstecher, E., Jr., and Spangler, S., (b) Looney, J. M. (c) Williams, J. N.

References: (1) Heller, *Endocrinology* **42** 161, 1948 (2) Krebs, H. A. *Ann. Rev. Biochem.* **17** 1, 1948 (3) Heller, *J. Lab. Clin. Med.* **30** 706, 1945 (4) Per-

Med. **32** 1385 1947 (5) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (6) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (7) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (8) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (9) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (10) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (11) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (12) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (13) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (14) Heller, *J. Lab. Clin. Med.* **32** 1415, 1948 (15) Jellinek, *J. Biol. Chem.* **53** 321, 1924 (16) Jellinek, *J. Biol. Chem.* **88** 85, 1930

## 77. NITROGEN CONTENT OF BLOOD NITROGEN COMPOUNDS

References: All values are calculated from molecular weights and formulae as presented in "Handbook of Chemistry and Physics," Hodgman, C. D., Editor, Chemical Rubber Publishing Co., Cleveland, 1948, except those for items 9, 11, 21, 42, 46, 50, and 51 which were calculated from data in "Human Biochemistry," Kleiner, I. S., C. V. Mosby and Co., St. Louis, 1948.

## 78. BLOOD PHOSPHORUS, INORGANIC

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-3 ABC & Fn 2	6	10 A-C & Fn 4	2a	17 A-C & Fn 4	2
4 AB & Fn 3	3	11 A-C & Fn 4	2	18 A-C & Fn 4	2
5 A-C & Fn 4	2a	12 AB & Fn 3	3	19 A-C & Fn 6	5a
6 A-C	2a	13 AB & Fn 3	3a	20 AB & Fn 3	3
7 A-C	5a	14 AB & Fn 3	3	21 AB & Fn 3	3a
8 AB & Fn 3	3	15 AB	3	22-24 A-C & Fn 3	5a
9 A-C & Fn 4	a	16 A-C & Fn 4	2	25-40 AB & Fn 3	3

Contributor: (a) Freeman, S.

References: (1) Brain, Kay and Marshall, *Biochem. J.* 22:628, 1928. (2) Kay, J. *Physiol.* 65 374, 1928. (3) Guest, G. M. and Rapoport, S., *J. Biol. Chem.* 138 269, 1941. (4) Rapoport, S. and Guest, G. M., *J. Biol. Chem.* 126 749, 1938. (5) Freeman, S. and Farmer, C. J., *Am. J. Physiol.* 113:200, 1935. (6) Helve, *Acta Med. Scand.* 125: 505, 1946.

## 79. BLOOD PHOSPHORUS, ORGANIC ACID-SOLUBLE

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,2 ABC & Fn 2	1a	15,16 AB & Fn 3	3	30 AB & Fn 3	3
3,4 AB & Fn 3	3	17-19 AB & Fn 4	2	31,32 AB & Fn 4	2
5 ABC	3a	20 AB & Fn 4	2a	33-36 AB & Fn 3	3
6-8 AB & Fn 3	3a	21-24 AB & Fn 4	2	37-40 AB & Fn 4	2
9,10 AB & Fn 4	2	25,26 AB & Fn 3	3	41-44 AB & Fn 3	3
11,12 AB & Fn 3	3	27,28 AB & Fn 3	2	45,46 AB & Fn 4	2
13,14 AB	3a	29 AB & Fn 3	3a	47,48 AB & Fn 3	3
				49-60 AB & Fn 4	2

Contributor: (a) Freeman, S.

References: (1) Helve, *Acta Med. Scand.* 125:505, 1946. (2) Rapoport, S., & Guest, G. M., *J. Biol. Chem.* 138 269, 1941. (3) Kerr, S. E., & Daoud, L., *J. Biol. Chem.* 109:301, 1935.

## 80. BLOOD ADENOSINE TRIPHOSPHATE PHOSPHORUS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,2 A-C & Fn 2	1	25,26 AB	1	39-42 ABC & Fn 2	1
3,4 AB & Fn 2	1	27,28 AB & Fn 2	1	43,44 ABC & Fn 2	1
5-8 AB & Fn 2	1a	29,30 AB & Fn 2	1	45,46 ABC & Fn 3	2
9,10 AB & Fn 3	2	31,32 AB & Fn 3	2	47,48 AB & Fn 2	1
11,12 AB & Fn 3	1	33,34 AB & Fn 2	2	49-52 AB & Fn 3	2
13,14 AB	2	35,36 AB & Fn 2	1	53-56 ABC & Fn 3	2
15,16 AB & Fn 2	1	37,38 AB	2	57-60 AB & Fn 3	2
17-24 AB & Fn 3	2				

Contributor: (a) Freeman, S.

References: (1) Kerr, S. E., & Daoud, L., *J. Biol. Chem.* 109 301, 1935. (2) Guest, G. M., & Rapoport, S., *J. Biol. Chem.* 138 269, 1941.

# 81. BLOOD DIPHOSPHOGLYCERATE PHOSPHORUS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,2 A-C & Fn 3	1a	13,14 AB & Fn 4	1	20 A-C	
3-5 AB & Fn 4	2	15 AB & Fn 4	2	21-27 AB & Fn 4	2
6 A-C & Fn 4	2	16 A-C & Fn 4	2	28 A-C & Fn 4	2
7-12 AB & Fn 4	2	17-19 AB & Fn 4	2	29-36 AB & Fn 4	a

Contributor: (a) Freeman, S.

References: (1) Helve, Acta Med. Scand. 125 505, 1946. (2) Guest, G.M., & Rapoport, S., J. Biol. Chem. 138 269, 1941.

# 82. BLOOD NUCLEOTIDE PHOSPHORUS

All data from Kerr, S. E. and Daoud, L., J. Biol. Chem. 109 301, 1935.

# 83. BLOOD LIPID PHOSPHORUS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1,2 AB & Fn 1	1b	5 AB & Fn 1	3	7 A-C & Fn 1	4a
3 A-C	2	6 A-C & Fn 2	6a	8 A-C & Fn 1	5a
4 AB & Fn 1	1b	6 AB & Fn 3	7a		

Contributors (a) Flock, E.V. (b) Stearns, G

References (1) Stearns, G. & Warweg, E., J. Biol. Chem. 102 749, 1933. (2) Page, Kirk, Lewis, Thompson & Van Slyke, D.D., J. Biol. Chem. 111 613, 1935. (3) Sinclair, R.G., J. Biol. Chem. 174 343, 1948. (4) Bollman, J.L. & Flock, E.V., J. Lab. Clin. Med. 31:478, 1946. (5) Flock, E.V. & Bollman, J.L., J. Biol. Chem. 144 571, 1942. (6) Baumann, E.J., & Holly, O.M., J. Biol. Chem. 55 457 1923. (7) Artom, C., & Freeman, J.A., J. Biol. Chem. 135 59, 1940.

# 84. BLOOD HEXOSE PHOSPHATE PHOSPHORUS

All data from Helve, Acta Med. Scand. 125.505, 1946.

# 85. BLOOD PHYTIC ACID PHOSPHORUS

All data from Guest, G M and Rapoport, S., J Biol Chem 138 209, 1941

# 86. BLOOD SULFUR

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-5 BCD	3a	10 BC	1	14 B-D & Fn 1	a, 1a
6,7 BC	2b	11,12 B-D	3a	15-17 BC	2b
8 BC	1	13 BC	2b	18-20 BCD	3a
9 BC	2b				

Contributors. (a) Lewis, H B. (b) Ponder, E

References (1) Power, M.H., & Wakefield, E.G., J Biol Chem 123 669, 1938. (2) Reed, L., & Denis, W., J. Biol. Chem. 73 191,623, 1927. (3) Brown, B.H. & Lewis, H B, J. Biol. Chem. 138 705, 1941

## 87. BLOOD SULFUR COMPOUNDS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-3 BCD	7a	16 BC	6c	27 BC	Calc.fr.(26 BC)
4, 5 BC	Calc.fr. 1	17 BC	Calc.fr.(18, 19 BC)	29 B-D	11a
6 B-D	7a		4b, 5b	30 B-D	Calc.fr.(29 BC)
7 BC	7a	18, 19 BC	10a	31 BC	5b
8, 9 B-D & Fn 1	7a	20 BC	Calc.fr.(20 BC)	32 BC	Calc.fr.(33, 34 BC)
11, 12 B-D & Fn 3	a	21 BC	6c	33 BC	4b, 5b
13 BC	6c	22 BC	2c	35 BC	10a
14 B-D	Calc.fr.(15, 16 B-D)	23 B-D	4b, 5b	36 BC	Calc.fr.(35 BC)
15 B-D	9a	24, 25 BC	10a	38-42 BCD	3c
		26 BC			

Contributors (a) Lewis, H. B. (b) Looney, J. M. (c) Wood, J. L.

1932,  
nek,  
H. and  
B.,

## 88 BLOOD VITAMINS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B-D GH	22a, 26a, 71a	43 B H & Fn 3	30a	75 B-D & Fn 6, 7, 13	1a, 41a, 50a
1-15 E & Fn 2	55a	44 B G & Fn 3	30a	76 B C & Fn 6, 7	50a
2 B-D GH	4a, 32a, 34a, 37a, 40a	45 B C & Fn 3	52a	77 B C & Fn 6, 7	1a, 41a, 50a
3 B-D GH	2a, 5a	46 B G & Fn 3	30a	78 B-D & Fn 6	63a
4 B-D GH	49a	47 B-D FH & Fn 1	30a, 58a	79 B-D & Fn 6, 7	1a, 41a, 48a, 50a
5 B-D GH	21a	48 B GH & Fn 1	30a	80 B-D & Fn 4, 6, 7	23
6 B-D GH	20a, 81a	49 B H & Fn 1	31a	81 B-D & Fn 6	44a, 62a, 66a
7 B-D GH	17a	50 B H & Fn 2	25a	81 B EF & Fn 6	42a
8 B-D G	45a	51-52 B G & Fn 1	30a	81 B GH & Fn 6	42a, 59a, 83a
8 H	24a	53 B G & Fn 1	25a	82 B-H & Fn 6	83a
9 B-D G	82a	54 B H & Fn 1	25a	83 B-H & Fn 6	a
10 B D GH	26a	55 B G H & Fn 1	30a, 31a	84 B-H & Fn 6	a
11 B-D G	34a	56 B G H & Fn 2, 3	76a	85 B-H & Fn 6	a
11 H	40a	57 B E G & Fn 3	74a	86 B-H & Fn 6	42a, 48a
12 B-D GH	49a	58 B G & Fn 2, 3	75a	87 B-D & Fn 6	16a, 43a
13 BC	21a	59-61 B GH & Fn 1	75a	88-89 BC & Fn 1	52a
14 B-D GH	45a	62 B GH & Fn 1	75a	90 BC & Fn 1, 2	60a
15 B-D G	82a	63 B GH & Fn 1, 2	46a, 67a, 71a	90 B D G & Fn 1, 2	10a
15 H	11a, 82a	64 B GH & Fn 1	13a, 14a, 46a		
16 B-D	39b	65 B GH & Fn 1	61a		
16 E	56a	66 B GH & Fn 1	79a		
16 GH & Fn 3	33a, 56a	67 B H & Fn 1	55a		
17 B-D GH & Fn 3	3a, 34a	68 B GH & Fn 1	79a		
18 B-D GH & Fn 3	35a, 49a	69 B GH & Fn 5	65a		
19 B-D GH & Fn 3	21a	70 BC & Fn 5	52a		
20 B-D GH & Fn 3	35a, 84a	71 B-D & Fn 6-9	28a, 47a, 54a		
21 B-D GH & Fn 3	70a	71 B E & Fn 6, 7, 10	27a		
22 B C G	49a	71 B GH & Fn 6, 7	8a, 29a, 38a		
22 B D H	57a	72 B C & Fn 6, 7	80a		
23-37 B-D & Fn 4, 7	9	73 B-E & Fn 6, 7, 10	1a, 41a, 50a		
38 B-D GH & Fn 3	10a	73 D-E & Fn 6	27a		
39-40 BC & Fn 3	52a	74 B D & Fn 6, 12	1a		
41 B H & Fn 3	30a				
42 B D & Fn 3	77a				
42 B G & Fn 3	30a				

(Continued on the next page)







(Continued from preceding page)

- (7) Dille, R. S., and Watkins, C. H., *J. Lab. Clin. Med.* **33**: 480, 1948. (8) Fishman, W. H., Springer, and Brunetti, J., *Biol. Chem.* **173**: 449, 1948. (9) Guest, M. M., Ware, A. G., and Seegers, W. H., *Am. J. Physiol.* **150**: 561, 1947. (10) Gutman, E. B., and Gutman, A. B., *J. Biol. Chem.* **136**: 201, 1940. (11) Hall, G. E., and Langer, C. J., *Pharmacol.* **61**: 10, 1937. (12) Hawkins, R. D., and Mendel, B., *Proc. Nat. Acad. Sci. U.S.A.* **42**: 103, 1946. (13) Huggins, S. R., *Canad. Med. Assoc. J.* **31**: 376, 1934. (14) Kochakian, C., *J. Biol. Chem.* **148**: 264, 1935. (15) Kochakian, C. D., *J. Biol. Chem.* **148**: 264, 1935. (16) Meister, A., *Science* **106**: 157, 1943. (17) Meister, A., *J. Nat. Cancer Inst.* **8**: 169, 1944. (18) Meister, A., *J. Am. J. Physiol.* **158**: 158, 1944. (19) Lehninger, A. L., *J. Biol. Chem.* **165**: 1941, 1941; Somogyi, M., *J. Biol. Chem.* **145**: 399, 1938. (20) Werle, E., and Effkemann, G., *Klin. Woch.* **18**: 717, 1940. (21) Wolff, R., Drouet, L., and Karlin, R., *Science* **109**: 612, 1949. (22) Comfort, M. W., and Osterberg, A. E., *Med. Clin. N. Amer.* **24**: 1137, 1940. (23) Warburg, O., and Christian, W., *Biochem. Z.* **242**: 206, 1931; *Biochem. Z.* **266**: 377, 1933. (24) Kiese, M., *Biochem. Z.* **315**: 264, 1944.

## 91. BLOOD COENZYMES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-E	2	5-17 ABCDE	a	20 A-E	a
2-4 ABCDE	1c	& Fn 2		21-23 ABCDE	4b
& Fn 1		18, 19 A-E	3b	& Fn 3	

Contributors (a) Beerstecher, E., and Spangler, S. (b) Granick, S. (c) Lehninger, A.

References (1) Goodhart, R. S., and Sinclair, H. M., *Biochem. J.* **31**: 1099, 1939. (2) Kaplan, N. O., and Lipmann, F., *J. Biol. Chem.* **174**: 37, 1948. (3) Klein, J. R., and Kohn, H. I., *J. Biol. Chem.* **135**: 177, 1940. (4) Levitas, N., Robinson, J., Rosen, F., Hult, J. W., and Perlzweig, W. A., *J. Biol. Chem.* **167**: 169, 1947.

## 92. BLOOD ELECTROLYTES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 BCD	41f, 42f	39 BCD	6	76 BCD	20e
2, 3 BCD & Fn 2, 3	15h, 14h, 15h	40 BC	19e, 33e	79 BCD	10c, 12c
4 BCD & Fn 2, 3	21	41 BCD	33e	80 BCD	31e
5 BCD	Calc. fr 6, 7 BCD	42 BCD	7d, 8d	81 BCD	Calc. fr 82, 83 BCD
6 BCD & Fn 3	9d	43, 44 BCD	37e	82 BCD	10c
7 BCD & Fn 3	11g, 25g, 35g	45, 46 BCD	4b	83 BCD	5e
8 BCD & Fn 3	3d, 7d, 8d, 9d	47 BCD	31d	84 BCD	31d
9-11 BCD & Fn 3	11g, 25g, 35g	48 BCD	24e	85 BCD	20e
12 BC & Fn 4	4b	49 BC	25e	86 BCD	10c, 12c
13 BC	40f	50 BCD	Calc. fr 51, 52 BCD	87 BCD	20e
14 BCD	31d	51 BC	43c	88 BC	Calc. fr 89, 90 BC
15 BCD	Calc. fr 16, 17 BCD	52 BCD	10a	89, 90 BC	3b
16, 17 BCD & Fn 3	g	53 BC	Calc. fr 54, 55 BCD	91 BC	Calc. fr 92, 93 BC
18 BCD	41f, 42f	54, 55 BC	7c, 12c	92, 93 BC	28b
19, 20 BCD & Fn 3	16b, 17h	56 BCD	10a	94 BC	Calc. fr 95, 96 BC
21 BCD	6d	57, 58 BCD	10c, 12c	95, 96, 98, 99 BC	28b
22 BCD	Calc. fr 23, 24 BCD	59 BC	Calc. fr 60, 61 BCD	97 BC	Calc. fr 98, 99 BC
23 BCD	9d	60 BC	7c, 12c	100 BC	2c, 22c
24 BCD	11g, 25g, 35g	61 BCD & Fn 7	4b	101 BCD	27f
25 BC	14e	62 BCD	7d, 8d	102, 103 BC	32f
26-28 BCD	11g, 25g, 35g	63 BC	Calc. fr 64, 65 BC	104-109 BC	1i
29 BC & Fn 3	4b, 4e	64 BC	10c, 10i	110 BCD	42f, 44f
30 BCD & Fn 3	5	65 BC	10c, 10i	111, 112 BC	15h
31 BCD	31d	66, 67 BC	28f	113, 114 BCD	15h
32 BC	36e	68 BC	38f	115 BCD	Calc. fr 116, 117 BCD
33 BCD	Calc. fr 34, 35 BCD	69 BC	Calc. fr 70, 71 BC	116 BCD	9d
34, 35 BCD	34g	70, 72, 76 BCD	10c, 12c	117 BCD & Fn 8	30a
36 BC	36e	71, 73, 74 BC	10c	118-120 BCD	25g
37 BC	26e	75 BCD	20e	121 BC & Fn 9	4b
38 BCD	18c	77 BCD	10c, 12c		

(Continued on the next page)

(Continued from preceding page)

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
122 BC	40d	127 BC	4b	132 BC	46e
123 BCD	g	128 BC & Fn 11	5	133 BCD	5
124, 125 BCD & Fn 10	15h	129 BCD	6d	134 B D	39
126 BCD & Fn 10	30a	130 BCD	29e	135 BC	30g
		131 BCD	45e	136-138 BCD	g

Contributors: (a) Allison, J. B. (b) Cole, W. H. (c) Collove, E. (d) Eichelberger, L. (e) Levine, V. E. (f) Manery, J. F. (g) Overman, R. R. (h) Peters, J. P. (i) Platner, W. S.

References: (1) Bethé, A., and Berger, E., Arch. ges. Physiol., (Pflügers) 227 571, 1931. (2) Fenn, W. O., Physiol. Rev. 16 450, 1936. (3) Childs, and Eichelberger, L., Am. J. Physiol., 137, 384, 1942. (4) Cole, W. H., Allison, J. B., Murray, T. J., Boyden, A. A., Anderson, J. A., and Leatham, J. H., Am. J. Physiol., 141:165, 1944. (5) Conway, E. J., and Hingerty, Biochem. J., 40:563, 1946. (6) Darrow, D. C., et al., J. Biol. Chem., 130 487, 1939. (7) Eichelberger, L., and McLean, F. C., J. Biol. Chem., 142:467, 1942. (8) Eichelberger, L., Eisele, and Wertzler, J. Biol. Chem., 151 177, 1943. (9) Eichelberger, L., and Roma, J., Am. J. Physiol., 159 57, 1949. (10) Eveleth, D. F., J. Biol. Chem., 119 289, 1937. (11) Flanagan, J. B., Davis, A. E., and Overman, R. R., Am. J. Physiol., 160 89, 1950. (12) Greenberg, D. M., et al., J. Biol. Chem., 100 139, calculated from Abderhalden, E., Z. Physiol. Chem., 25 65, 1898. (13) Hald, P. M., J. Biol. Chem., 103 471, 1933. (14) Hald, P. M., Heinsen, A. J., and Peters, J. P., J. Biol. Chem., 118 275, 1937. (15) Hald, P. M., Heinsen, A. J., and Peters, J. P., J. Clin. Invest., 26 983, 1947, and unpublished data. (16) Hald, P. M., and Eisenmann, O. J., J. Biol. Chem., 118 275, 1937. (17) Hald, P. M., unpublished data. (18) Kramer, H., and Tisdall, F. K., J. Biol. Chem., 53:241, 1922. (19) Kinsell, L. W., and Zwemer, R. L., J. Lab. & Clin. Med., 27:206, 1941-42. (20) Kunkel, Pearson, P. B., and Schweigert, B. S., J. Lab. & Clin. Med., 32 1038, 1947. (21) Levin, L., Leatham, J. H., and Crafts, R. C., Am. J. Physiol., 136 776, 1942. (22) Macallum, A. B., Physiol. Revs., 6:316, 1936. (23) MacDonald, M. R., and Riddle, O., J. Biol. Chem., 159 445, 1945. (24) Marlow, H. W., and Richards, D., Endocrinology, 27:274, 1940. (25) Overman, R. R., Am. J. Physiol., 152:113, 1948. (26) Pereira, R. S., J. Biol. Chem., 160 617, 1945. (27) Platner, W. S., Univ. Microfilm, (Ann Arbor, Mich.,) Publ. No. 1231, pp289, Microfilm Abstract, No. 2, 20, 1949. (28) Platner, W. S., Univ. Microfilm, (Ann Arbor, Mich.,) Publ. No. 1231, pp289, 1949. (29) Robertson, J. D., Lancet II 97, 1941. (30) Root, W. S., Allison, J. B., Cole, W. H., Holmes, L. H., Walcott, W. W., and Gregersen, M. I., 142 52, 1947. (31) Smith, P. K., and Smith, A. H., J. Biol. Chem., 107 673, 1934. (32) Smith, H. W., J. Biol. Chem., 81 407, 1921. (33) Sobel, A. C., Kraus, G., and Kramer, B., J. Biol. Chem., 140 511, 1946. (34) Stern, T. N., Cole, V. V., Bass, A. C., and Tomlinson, J. H., Jr., Fed. Proc., 9 1, 1950. (35) Stern, T. N., Cole, V. V., Bass, A. C., and Overman, R. R., Am. J. Physiol. in press. (36) Velick, S. F., and Scudder, J., Am. J. Hygiene, (Sec C) 31:32, 1940. (37) Snyder, L. H., and Tweedy, W. D., Proc. Soc. Exper. Biol. & Med., 47:234, 1941. (38) Soumalainen, P., Nature, 141 471, 1938. (39) Ham, T. H., New England J. Med., 241:487, 1949. (40) Manery, J. F., et al., J. Biol. Chem., 124 359, 1938. (41) Hald, P. M., J. Biol. Chem. 163 429, 1946. (42) Hald, P. M., Heinsen, A. J., and Peters, J. P., Am. J. Physiol. 152 77, 1948. (43) Haury, V. G., J. Lab. Clin. Med. 27:1361, 1942. (44) Dill, D. B., Talbott, J. H., and Edwards, H. T., J. Physiol. 69 267, 1930. (45) Allison, J. B., Cole, W. H., Holmes, J. H., and Root, W. S., Am. J. Physiol. 149 422, 1947. (46) Behrndt, H., Am. J. Dis. Child, 64:789, 1942.

## 93. BLOOD MINOR MINERALS

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-3 BCD	11e	21, 22 B-D	21b	34 B-D	2a
4 B D	5	23 B-D	3, 4, 14, 15, 16, 25b	35 BC	8a
5-7 BCD	7a	24 B-D	19c	36-38 BCD	11e
8, 9 B D	1b	25, 26 B-D	22b	39, 40 BC	23
10 BC	14b	27 BC	10b	41-43 BCD	11e
11-13 BCD	20b	28, 29 B-D	22b	44 BC	21b
14 B-D	2a	30 B-D	9, 18d	45 B-D	12b
15, 16 B-D	20b	31 B-D	17c	46 BC	26b
17 BC	6b	32 B-D	19	47, 48 BC	12b
18-20 BCD	13e	33 B-D	8a	49, 50 BC	26b
				51-54 BCD	11e
				55-58 BCD	24

Contributors: (a) Cartwright, G. E. (b) Levine, V. E. (c) Peters, J. H. (d) Peters, J. P. (e) Largent, E. J.

(Continued on the next page)

(Continued from preceding page)

- References. (1) Beck, A., Australian J. Exper. Biol. and Med. Sci., 19-20: 145, 1940-42. (2) Cartwright, G. E., and Wintrobe, M. M., J. Biol. Chem., 126: 571, 1948. (3) Davis, C. B., Curtis, G. M., and Cole, V. V., J. Lab. and Clin. Med., 19: 818, 1934. (4) Fowweather, F. S., Brit. J. Exp. Path., 11: 400, 1930. (5) Greenstein, J. P., and Thompson, J. W., J. Nat. Cancer Inst., 1941. (6) Greenstein, J. P., and Thompson, J. W., unpublished data. (7) Hamilton, G. B., unpublished data. (8) Heinemann, M., Johnson, C. L. and Man, E. D., J. C. R. V., J. Nutrition, 19: 579, 1940. (9) King, E. J., unpublished data. (10) Hinton, J. E., Eckerson, B. and Bruger, M., Ann. Surg., 115: 206, 1941. (11) Litch, S., M. State J. Med., Obst. and Gyn. and Steenbock 1948. (12) Ta (24) Valler, B. Med 167: 186

#### 94. BLOOD GASES MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 1,2 1,2 A & Fn 5 1,2 B Fn 6 1,2 E Fn 8 3-6 ABCDEF	m 1n,2n, 13e k, 4k 3, 4k (1B) k (7-10 ABCDEF & 11-14 ABCDEF) Av. of 5d, 8h, 11h, 12h, 13e, 16ehm, 18h, 27j, 28j 3,4 G Av. of 5d, 6ch, 7ch, 8h, 9ch, 10ch, 11h, 12h, 13el Av. of 47f, 48f, 49f (15, 14H) & 50hk Av. of 5d, 13el, 11h (7-10 EF) (7-10 GH) (3,4 G) (5,6G) 11,12 A-C 11,12 D 11,12 E 11,12 F 11,12 G 11,12 H 13,14 A-C 13,14 D 13,14 E 13,14 F	13,14 G 13,14 H 15 AB 16AB & Fn 17 15,16 C 15,16 D 15,16 E 15,16 F & Fn 15 15 G 16G Fn 18 Fn 19 15,16 H 15,16 I 17,18 AB 17,18 C 17,18 D 17,18 E 17,18 F 17,18 G 17,18 H 18-22 ABCD 19-22 E 19-22 F 19-22 G	(5,6G) (1,2B,13,14B) L 51hk Av. of 5d, 25g, 31g, 27j, 28j, 17j, 21g, 19a, 45g, 46g, 26g (15 AB) & 25k (15,16B & 15,16D) (15,16F) (15,16C) (15,16B & 15,16H) Av. of 5d, 25g, 31g, 6h, 7h, 32g, 6h, 34h, 27j, 11h, 12h, 28j, k, 10j, 19a, 21g, 45g, 46g, 26g (16F & 24f & Fn 18) 32m, 25m 33m 22m, 23m, 24m 30m (3,4f & Fn 16) (15,16B & 15,16I) (17,18B, 17,18D) (17,18F) (17,18C) (17,18B & 17,18H) (17,18F & 25, 26f & Fn 19) (15,16H) (19-22E, F) (19-22G, H) (19-22G, I) (15-18G)	18-22 HI & Fn 20 23-26 ABCDEF 23-26 G 23-26 H 23 I 24 I 25 I & Fn 21 26 I 27-30 AB 27-30 C 27-30 D 27-30 E 27-30 F 27-30 G 27-30 HI & Fn 22 31-34 ABCDEF 35-38 AB 35-38 C 35-38 D 35-38 E 35-38 F 35-38 G & Fn 23 35-38 HI & Fn 24	20m, 21m (15-18 ABCDEF & 19-22 ABCDEF) (15-18G) 21k Av. of 5d, 8h, 12h, 18h, 21gm, 25g, 26g, 27j, 28jm, 39m, 40m, 42m, 43m, 45g, 46g, m (23f & Fn 8) 25g k (26F & 26 G & Fn 19) k (27-30C & 27-30D) (27-30E) (27-30F) (27-30H & 23-26E) (27-30I & 23-26F) (15-18G) Calc. by k based on 21k, 35k, 36k, 37k, 38k, (23-26 ABCDEF & 27-30 ABCDEF) (31-38C & 35-38D) (35-38E) (35-38F) (35-38G, 35H) (35-38G & 35I) Calc. by k based on 7h, 11h, 12h 14m, 44m

Contributors (a) Barron, D. H. (b) Bing, R. J. (c) Comroe, J. H., Jr. (d) Courmand, A. (e) Drabkin, D. L. (f) Hickam, J. B. (g) Kirk, J. E. (h) Lambertsen, C. J., & Kough, R. H. (i) Otis, A. B. (j) Penrod, K. E. (k) Singer, R. B. (m) Singer, R. B. & Hastings, A. B. (n) Van Slyke, D. D. (o) Wood, E.

References (1) Bernhart, F. W. & Skeggs, L., J. Biol. Chem., 147: 19, 1943 (2) Van Slyke, D. D., Hiller, A., Weisiger, J. R., & Cruz, W. O., J. Biol. Chem., 166: 121, 1948 (3) Table, Erythrocytic Values (4) Maxfield, M. E., Bassett, H. C., Chambers, C. C., Am. J. Physiol., 133: 128, 1941 (5) Courmand, A., unpublished data, 1949. (6) Lilienthal, J. L., Jr.,

(Continued on the next page)

Riley, R. L., Proemmel, D. D. & Franke R. E. *Am. J. Physiol.* 147: 100, 1954. (9) Aviado, D. M., Jr., & Pontius, unpublished data. (10) Comroe, J. I., unpublished data. (11) La, unpublished data. (12) Drabkin, J., Jr., Dillon, R. T., & Van Slyke, D. D., *Physiol.*, 152: 365, 1948. (13) Henderson, L. J., *J. Biol. Chem.*, 151: 567, 1949. (14) Henderson, L. J., *J. Biol. Chem.*, 78: 765, 1928. (15) Hastings, A. B., Sendroy, J. W., and Cullen, G. E., *J. Biol. Chem.*, 106: 7, 1934. (16) Rossier, et al., *Rev. Med. Suisse Romande*, 60: 633, 1940. (17) Shock, N. W., and Hastings, A. B., *J. Biol. Chem.*, 104: 585, 1934. (18) Peters, J. P., Barr, D. P., and Rule, F. D., *J. Biol. Chem.*, 144: 325, 1942. (19) Courmand, A., Riley, R. L., Breed, E. S., Baldwin, E. deF., and Richards, D. W., Jr., *J. Clin. Invest.*, 24: 106, 1945. (20) d'Elseaux, F. C., Blackwood, F. C., Palmer, L. E., and Sloman, K. G., *J. Biol. Chem.*, 144: 529, 1942. (21) Van Slyke, D. D., Dillon, R. T., and Margaria, R., *J. Biol. Chem.*, 105: 571, 1934. (22) Dill, D. B., Coulaert, C. V., Hurxthal, L. M., C. W., Proc. Respiratory 1944.

Arbeitsphysiologie, 10: 251, 1938. (23) Gibbs, E. L., Lennox, W. G., Nims, L. F., and Gibbs, F. A., *J. Biol. Chem.*, 144: 325, 1942. (24) Courmand, A., Riley, R. L., Breed, E. S., Baldwin, E. deF., and Richards, D. W., Jr., *J. Clin. Invest.*, 24: 106, 1945. (25) d'Elseaux, F. C., Blackwood, F. C., Palmer, L. E., and Sloman, K. G., *J. Biol. Chem.*, 144: 529, 1942. (26) Van Slyke, D. D., Dillon, R. T., and Margaria, R., *J. Biol. Chem.*, 105: 571, 1934. (27) Dill, D. B., Coulaert, C. V., Hurxthal, L. M., C. W., Proc. Respiratory 1944.

#### 95. ARTERIAL BLOOD GASES, O<sub>2</sub> AND CO<sub>2</sub>, AT ALTITUDE: MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 B E-G	See Table 94	24 C	2	37 B	1b
1 C	2	25 E	4b	37 C-G	6ab
2, 4, 6, 8, 10, 12,	1b	26 B E-G	4b	38 C-G	6b
14, 16 B		26 C	2	39 B	1b
2-17 C	2	27 E	Av. of 1, 3, 4	39 C-E G	6ab
2-17 E	3b	27 FG	4b	40 C-E G	6b
2-17 F	1b	28 B E G	4b	41 B	1b
18 C	2	29 E	Av. of 1, 3, 4	41 C-G	6ab
18, 19 E	3b	29 G	4b	42 C-G	6b
20 B E-G	4b	30 B E-G	4b	43 B	1b
20 C	2	30 C	2	43 C-G	6ab
21 B E-G	5b	31 E	Av. of 1, 3, 4	44 C-G	6b
21 C	2	32 C	2	45 B-G	7b
22 B E-G	4b	32 E	1	46 B-G	9ab
22 C	2	33 E	Av. of 1, 3	47 B-E G	7ab
23 E	Av. of 1, 3, 4	34 B-G	1b	47-49 CD	8
23 FG	4b	35 B	6a	48 B E-G	7ab
24 B E-G	4b	35 C-G	6ab	49 B E-G	7ab
		36 C-G	6b	Fn 6	6ab

Contributors. (a) Adler, H. F., and Luft, U. C. (b) Penrod, K. E.

References. (1) National Research Council CAM Report "Handbook of Respiratory Data in Aviation," Charts A-1, B-1,

## 96. IONIC BALANCE AND BUFFER BASE, ARTERIAL BLOOD, CELLS AND PLASMA: MAN

Diagram contributed by Singer, Richard B.

## 97. ACID-BASE VALUES, BLOOD AND PLASMA MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
Fn 1	a, 1a	3 P	23a, 24a	12 FG	1a, 15a, 16a, 19a, 20a
Fn 2	a, 47a, 48a, 49a	6 B-1	40a		
Fn 3	a, 30a, 32a	6 PQ	23a, 24a, 25a	12 HK	1a, 15a, 16a, 17a, 18a, 45a
Fn 4	a, 50a, 51a, 52a, 42a	7 B-E	3a		
Fn 5	a, 53a, 54a, 55a	7 F-O	1a, 3a, 41a	12 L-O	1a
Fn 6	a, 56a, 57a, 1a	9 FG	1a, 41a	12 PQ	26a, 27a, 28a
Fn 7	a, 58a, 59a, 60a, 61a, 62a, 1a, 63a, 64a	9 HI	41a	14 BG	1a, 11a
		9 J-O	1a, 41a	14 HI	1a
Fn 8	a, 65a	9 PQ	23a, 24a, 25a, 26a	14 JK	1a, 11a
Fn 9	a, 1a	10 BC	1a, 3a, 4a, 5a, 6a, 7a	14 LM	1a
Fn 10	a, 36a, 57a, 64a, 1a, 67a	10 DE	38a, 42a, 43a, 44a, 45a	14 NO	1a, 11a
Fn 11	a, 65a, 1a, 29a, 36a	10 FG	1a, 3a, 4a, 5a, 6a, 7a, 8a	15 DE	37a
Fn 12	a, 68a, 39a, 69a, 70a	10 HI	1a, 7a, 9a	15 FG	1a
Fn 13	a, 41a	10 JK	1a, 4a, 5a, 6a	15 HI	18a, 21a, 22a, 46a
Fn 14	a, 39a	10 LM	1a	15 JK	1a, 18a, 22a, 46a
Fn 15	a, 1a, 38a	11 BC	1a, 3a, 7a, 8a, 9a	15 LM	1a, 21a
Fn 16	a	11 FG	1a, 11a, 12a, 13a	15 NO	1a
Fn 17	a	11 HI	1a	15 PQ	28a
Fn 18	a, 3a, 1a	11 JK	1a, 11a, 12a, 13a	15 H-M	10a
Fn 19	a	11 LM	1a	16 NO	1a, 10a
Fn 20	a, 3a, 1a	11 NO	1a, 11a, 12a, 13a	17 B-G	1a, 13a
2 B-O	39a	12 BC	1a, 5a, 16a, 19a, 20a	17 HI	1a
3 EC	40a	12 DE	1a, 15a, 16a, 19a, 20a, 37a	17 JK	1a, 13a
3 DE	20a			17 LM	1a
3 H-O	14a, 1a			17 NO	1a, 13a
				19 B-G	10a
				19 HI	1a
				19 JM	10a
				19 NO	1a, 10a

Contributor, Singer, R. B.

References (1) Singer, N. B., and Hastings, A. B., "Acid-Base Data for Human Blood. Normal Standards and Some

(Continued on the next page)



# 99. EFFECTS OF RADIATION ON HEMATOPOIETIC TISSUES

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1-10 ABCDEF	8a	35-37 ABCDEF	9a	102-107 ABCDEF	1a
11-14 ABCDEF	10a	38-42 ABCDEF	3a	108-111 ABCDEF	7a
15-17 ABCDEF	1a	43 A-F	1a	112-114 ABCDEF	1a
18, 19 A-F	9a	44-49 ABCDEF	3a	115 A-F	6a
20, 21, 22 A-F	1a	50-60 ABCDEF	1a, 3a	116-135 ABCDEF	1a, 6a
23-27 ABCDEF	3a	61-97 ABCDEF	1a	136 A-F	2a
28-34 ABCDEF	1a	98-101 ABCDEF	7a		

Contributors. Ingram, M.

References: (1) Bloom, W. "The Histopathology of Irradiation." McGraw-Hill, NY, 1948. (2) Bonham, Donaldson, Foster, et al., *Growth* 12: 107, 1948. (3) Brecher, G., Endicott, K. M., Cump, H., and Brawner, H. P., *Blood* 2: 1259, 1948. (4) Denstad, T., *Acta Radiol. [Suppl. 52]*, 1943. (5) Hagen, C. W., Jacobson, L. O., Murray R., and Lear, F., *MDDC* 893, 1944\*. (6) McDonald, et al. "Neutron Effects on Animals." Williams & Wilkins, 1947. (7) Metcalf, Inda, et al., *UR* 123, 1950. (8) Murray, R., Pierce, M., Jacobson, L. O., *CH* 3873, 1948\*. (9) Kettleship, A., *Radiol.* 42: 64, 1949. (10) Rekers, P. E., *UR* 11, 1948\*.

\*For information regarding the availability of starred references, the Reference Services Branch, Technical Information Service, U. S. Atomic Energy Commission, Oak Ridge, Tennessee should be contacted.

# 100. CHANGES IN STORED PRESERVED BLOOD

Data Coordinates and Footnotes	Contributors and References
1-45 & Fn 1-14	Read from smoothed curves in 1
46	Read from smoothed curve in 2

References (1) Rapoport, S. J. *Clin. Invest.* 26 591 1947 (2) Strumia, M.M., and McGraw, J.J., "Blood and Plasma Transfusions," F.A. Davis Co., Phila., 1949.

# 101. EFFECTIVE BLOOD LEVELS OF THERAPEUTIC AGENTS. MAN

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
1 A-F	1a	16 A-E G	14a	31-37 ABCDE G	a
2 A-F & Fn 1	2a	17 A-F	15a	38 A-F	18a
3 A-F	3a	18 A-F	16a	39 A-F	19a
4 A-F	4a	19 A-F	17a	40 A-F	20a
5-7 ABCDEF	5a	20 A-E G & Fn 4	18a	41-43 ABCDE H & Fn 7	30a
8 A-F	6a	21 A-F	19a	44 A-F & Fn 8	31a
9 A-F	7a	22 A-E G	20a	45 A-E G	32a, 33a
10 A-F	8a	23 A-E H	21a	46 A-F	34a
11 A-F	9a, 10a	24 A-E H & Fn 5	22a	47-49 ABCDEF	35a, 36a
12 A-F	11a	25 A-E H	23a	50 A-F	37a
13 A-F	12a	26 A-F	24a	51-53 ABCD G	38a
14 A-E H & Fn 2	13a	27 A-E G & Fn 6	25a		
15 A-F & Fn 3	1a	28 A-E G	26a		
		29 A-E G	27a		
		30 A-E G	28a		

(Continued on the next page)



(Continued from preceding page)

Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References	Data Coordinates and Footnotes	Contributors and References
54 A-E H & Fn 9	39a	80, 81 AB DE H & Fn 14	53a	98 A-D G	
55 A-E G	40a	82 A-F	54a	99 A-E G	62a, 63a
56 A-E G	41a, 42a	83-85 ABCDE G & Fn 15	55a	100 A-D F	64a, 65a
57, 58 A-E G	43a	86 A-F	56a	101 A-D G	66a
59 A-E G	44a	87 A-E G	57a	102 A-D F	62a, 63a
60 A-F & Fn 10	45a	88, 89 A-F	58a	103 A-E G	67a
61 A-F	46a	90 A-F	59a	104 A-D F	66a, 68a
62, 63 A-E G	47a	91-93 ABCDEF & Fn 16	60a	105 A-D F	68a
64-67 ABCDE G	48a	94 A-F	61a	106 A-D F	69a
68 A-E H	49a	95 A-F	a	107 A-E G	70a
69 A-E G & Fn 11	48a, 50a, 51a	96-97 A-E G		108-112 ABCDE H	71a
70 A-E G	52a			Fn 17	
71-73 ABCD G				113 A-E G	72a
74-79 ABCDE G 1 & Fn 12, 13				114 A-F	73a
				115 A-F	74a

Contributor: (a) Koppány, T.

- References: (1) Fisher, R. S., N. England J. Med. 240: 395, 1949. (2) Levy, G. A., Biochem. J. 34: 76, 1940. (3) Buckmaster, G. A. and Gardner, J. A. Proc. Roy. Soc. B. 29: 555, 1907. (4) Robbins, B. H., "Cyclopropane Anesthesia," Williams & Wilkins, Baltimore, 1940. (5) Ellerbrook, L. D. and Van Caasbeek, C. B., J. Am. Med. Assoc. 122: 996, 1943. (6) Jetter, W. W., Clinics (Phila.) 1: 1487, 1943. (7) Camus, J. and Niclaus, M., J. Physiol. path. 10: 76, 1908. (8) Niclaus, M. and Yovanovitch, A., Compt. rend. Soc. biol. 93: 1654, 1937. (9) Robbins, B. H., J. Pharm. Exp. Therap. 53: 251, 1935. (10) Potter, R., et al., Surgery 10: 761, 1941. (11) Kibler, D. V., Anesthesiology 8: 293, 1947. (12) Crowell, W. S., Anesth. and Analg. 4: 24, 1925. (13) Gardner, H. L., Levine, H., and Bodansky, M., Am. J. Obst. Gyn. 40: 438, 1940. (14) Jailer, J. W. and Goldbaum, L. R., J. Lab. Clin. Med. 31: 1347, 1946. (15) Sebenius, W. J., Canad. J. Res. 23: 48, 1945. (16) Domanski, T., J. Biol. Chem. 119: 89, 1937. (17) Cameron, A. T. and Gutherie, and Ligon, E. W., J. Exp. Med. 73: 361, 1947. (18) Kimbly, L. W., J. Invest. Derm. 2: 335, 1939. (19) Way, E. L., Cimble, A. J., and Rubin, B., J. Pharm. Exp. Therap. 92: 314, 1949. (20) Miller, J. H. and Whitehead, R. W., Rocky Mountain Med. J. 1, 1942, 1943. (21) J. Pharm. Exp. Therap. 92: 314, 1949. (22) Sampson, J. V., Albertson, C. C., Anesthesiology 8: 239, 1947. (23) Salter, W. T., Sciarini, L. J., and Boyle, M. N., Am. J. Med. 4: 377, 1948. (24) Robertson, R. W., and Peterson, M. C., Anesthesiology 8: 239, 1947. (25) Wegria, R., and Munnich, W. J., Clin. Invest. 18: 569, 1939. (26) McGavack, T. S., and Silverman, I. J., Am. Heart J. 28: 142, 1944. (27) Berger, F. M., J. Pharm. Exp. Therap. 95: 245, 1949. (28) Rose, B., J. Allergy 12: 442, 1941. (29) Moore, C. V., Arrowsmith, W. R., and Heisler, A., J. Allergy 19: 251, 1948. (30) Rose, B., J. Allergy 12: 442, 1941. (31) Cattell, R. H., Dreken, I. J., Schutler, S. R., and Caldwell, E. H., Med. Assoc. State Alabama 2: 803, 1948. (32) Christensen, B. and Larkin, C. V., Arrowsmith, W. R., Parker, R. T., Lister, L. M., Bauer, R. E., Hall, H. E., and Woodward, T. E., J. Am. Med. Assoc. 143: 9, 1950. (33) Rantz, L. A., and Kisby, W. M., J. Clin. Invest. 28: 792, 1944. (34) Ory, E. J., Am. Med. Assoc. 143: 9, 1950. (35) Lepper, M. H., Dowling, H. P., Brickhouse, R. L., C., and Finland, M., J. Lab. Clin. Med. 30: 817, 1945. (36) Seod, J. C., and Wilson, C. E., Science 110: 707, 1945. (37) Sussman, M. L., Janowitz, H. D., and Blum, L., Proc. Soc. Exp. Biol. Med. 233: 581, 1945. (38) King, F. H., Schnelzer, S. S., Israel, H. L., Filpken, H. P., Am. J. Med. Sci. 215: 252, 1948. (39) Priest, W. S., and O'Neill, J. B., Am. J. Med. 4: 363, 1948. (40) King, E. G., Lewis, C. N., Welch, H. Clark, E. A., Johnson, J. J., Lyons, J. B., Am. J. Med. 4: 363, 1948. (41) Schaffer, A. T., and Lewis, R. A., Bull. Johns Hopkins Hosp. 78: 270, 1946. (42) Heart, E. P., J. Pharm. Exp. Therap. 72: 157, 1943. (43) Otto, G. F., Maren, T. H., and Brown, H. W., Am. J. Hyg. 46: 1193, 1947. (44) Boyd, M. F., "Malariaology," vol. 2, W. B. Saunders Co., Phila., 1949. (45) Henning, H. B., and Kampmeier, R. H., Arch. Derm. Syph. 45: 783, 1942. (46) Hawking, F., and Macleach, E. H., Am. J. Syph. 30: 147, 1946. (47) Spink, W., Arch. Derm. Syph. 45: 783, 1942. (48) Fulton, D. J., and Goodwin, T. W., J. Pharm. Exp. Therap. 72: 157, 1943. (49) Mattis, P. A., Korn, E. S., and Laiter, A. R., Am. J. Med. Sci. 215: 252, 1948. (50) Siegel, J., Goldstein, D. H., and Goldwater, L., Arch. Derm. Syph. 45: 783, 1942. (51) Hawking, F., and Macleach, E. H., Am. J. Syph. 30: 147, 1946. (52) Denko, C. W., Health 25: 157, 1946. (53) Hertz, R., Tullner, W. W., Westfall, B. K., Morrow, A. G., Emge, M. K., Smith, E. L., Samuel, L. T., Davenport, H., J. Clin. Endocrinol. 9: 593, 1949. (54) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (55) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (56) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (57) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (58) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (59) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (60) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (61) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (62) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (63) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (64) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (65) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (66) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (67) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (68) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (69) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (70) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (71) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (72) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (73) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (74) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (75) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (76) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (77) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (78) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (79) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (80) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (81) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (82) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (83) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (84) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (85) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (86) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (87) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (88) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (89) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (90) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (91) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (92) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (93) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (94) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (95) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (96) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (97) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (98) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (99) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948. (100) Archer, H. E., Chapman, L., Rhodes, E., and Warren, P. W., J. Clin. Endocrinol. 9: 911, 1948.

# INDEX



## Index

- [illegible]

CHROMATIN PATTERN, in bone marrow cells, 77, 79, 81, 83, 85, in leukocytes, 55, 56

CHROMOMERE, platelets, 58

CITRIC ACID, 91

CITRULLINE, 99, 102

CIVET, 45

CLOT RETRACTION, 19

CLOTTING TIME, 16, 17

COAGULATION AGENTS, 97

COAGULATION THEORIES, Owren, P. A., 11, Quick, A. J., 12, Seegers, W. H., 13, Stefani, M., 14, Tocantins, L. M., 15

COATL, 45

COCAPOXYLASE, 116

COD, 7, 8

COENZYME A, 116

COENZYME CONTENT, 116

COLORADO, 40

COMPLEMENT COMPONENTS, human plasma, 97, 98

CONDUCTIVITY, electrical, human serum, 7

CONGER CONGER, 8, 44, 88, 103

CONJUGATED ESTROGENS, 143

CONNOCHAEUS GNOU, 45

CONTACT, in blood coagulation, 11-15

CONVERSION FACTOR, plasma prothrombin, 14

COPPER, 97, 119

CORTICOSTEROIDS, 112

COW, 21, 43, 45-47, 87, 92, 93, 98, 100, 108-113, 119

CRAFFISH, 89

CREATINE, 101, 102

CREATININE, 101, 102

CRUSTACEA, 118

CYCLOPROPANE, 138

CYCLOTRON NEUTRONS, effect on peripheral blood, 126, 127, 134

CYNOPTHECUS, 45

CYPRINUS CARPIO (See Carp)

CYSTINE, 99, 102

CYSTOSE, 99, 102

CYTOLASM, in bone marrow cells, 76, 78, 80, 82, 84, chemical contents, 57-63, 86, histochemical properties, 86, morphology, 55, 56, 76, 78, 80, 82, 84, stain reactions, 57-63, 68

DANES, 32

DASYATIS CENTROURUS, 21, 44

DASYBRANCHUS, 89

DASYURUS VIVERRINUS, 45

DEER, 45

DEGENERATED CELL, 70

DEHYDROGENASE, glucose-6-phosphate, 115, lactic, 115

DEHYDROPEPTIDASE, 115

DEMOL, 138

DENMARK, 26, 39

142

DIDELPHIS MARSUPIALIS (See Opossum)

DIDETHYLAMINOETHANOL, 142

DIGITOXIN, 139

DIMETHYLAMINOETHYL BENZHYDRYL ETHER HYDROCHLORIDE, beta-, 139

DINGO, 45

DIPHOSPHOGLYCERIC ACID, 91

DIPHENTERIA, antitoxin, 97, 98, toxin, 97

DISINTEGRATED CELLS, 70-73

DIVINYL OXIDE, 138

DOG, 5-10, 21, 23, 42, 45, 49, 53, 74, 75, 87-93, 96, 99-101, 103-113, 117-119, 125, 129

DOGFISH, 8, 21, 44, 103-105

DORIS SUBERCOLATA, 118

DOVE, 8

DPN, 90, 116

DROMEDARY, 10

DRUGS, effective levels in blood, 138-143,

Interaction with albumin, 97

DUCK, 43, 88, 90, 91, 103-107

DYES, interaction with albumin, 97

ECHINIS NAUCRATES, 44

ECHINODERMATA, 118

ECHINUS ESCULENTUS, 118

EL, 8, 44, 88, 107

EEL POUT, 44

EGYPT, 28, 32

ELASMOBRANCH(S), 9 (See also specific elasmobranchs)

ELECTRICAL PROPERTIES, erythrocytes, 7

ELECTROLYTES, in blood, 117, 118 (See also specific electrolytes)

ELECTROPHORETIC MOBILITY, erythrocytes, 7, 37

ELEPHANT, 45, 103-106

ELAPHAS MAXIMUS, 45, 103-105

ELK, 45

FQUS, 45

ERGOTHIONEINE, 102, 108

ERYTHROCEBUS PATAS, 45

ERYTHROBLAST(S), 87, 89, 70, 80-83

ERYTHROCYTE(S), agglutination, 23, 31, 33-36, anions in, 123, vs. anti-Lu, 35, vs. anti-P, 35, vs. anti-Su, 23, 31, 33, 34, arterial, 120, cations in, 123, changes in when preserved and stored, 135, 137, counts, 37, 42, 44, 45, diameters, 37, 42, dimensions, 137, electrical charge, 7, 37, electrophoretic mobility, 7, fragility, 10, 37, hemoglobin content, 38, 47, life span, 37, mass, 37, morphology, 82, oxygen consumption, 48, packed volume (See Hematocrit), pH, 37, production rate, 37, sedimentation, 9, solids in, 87, specific gravity, 5, 37, spherocytic index, 37, surface area, 37, thickness, 37, venous, 120, volume, 37, 38, 41, 42, 47, water in, 87

ESKIMOS, 30

ESOX LUCIUS, 7, 6, 44

ESTRIOL, 97

ESTROGEN, as estradiol, 112

ESTRONE(S), conjugated, 143

ESTRONE SULFATE, 143

ETHYL ALCOHOL, 138

ETHYL CHLORIDE, 138

ETHYL ETHER, 138

ETHYLENE, 138

ETHYLSTIBAMINE, 141

EUGLOBULIN(S), 85, 97, 98, antibody, 87, 98, beta<sub>2</sub>-lipid-poor, 97, 98

EUPHRASIS SEXCINCTUS, 45

EWE, 119

FAD, 90, 116

FARM ANIMALS (See also specific animals), leukocyte counts, 54, platelet counts, 23

FAT, neutral, 92

FATTY ACIDS, 93, 97

FELIS, 45 (See also Cat)

FERRA CELL, 70

FERRIC SODIUM CITRATE, 139

FERRUS AMMONIUM SULFATE, 140

FETUSES), 46, 47, 88, 89 at fractions of term, 45, 47

FIBRIN, clot formation, 97, in coagulation, 11-15

FIBRINOGEN, 94-98, in coagulation, 11-15, properties and reactions, 97, physical properties, 98

FILIPINOS, 30

FINNS, 26, 32

FISH, 7, 8, 44, 88, 91 (See also specific fish, as Bullhead, Dogfish)

FLAVINE ADENINE DINUCLEOTIDE, 90, 118

FLUORINE, 118

FOLIC ACID, 102, 111, 143

FOLLICLE STIMULATING HORMONE, 112

FOWL, 9 (See also specific fowl)

FOX, 45, 88

FREZZING POINT, serum, 7

FRENCH, 32

FRUCTIONAL RATIO, plasma proteins, 98

FROM, 8, 43, 48, 86, 90, 103-105, 111, 118, 119

FRUCTOSE, 89, metabolism of, 49

FSH, 112

FUMARATE, metabolism of, 49

PUMARIC ACID, 91

PUNCTIONS, of blood cells, 57-65

GADUS MORRHUA, 7, 8

GALACTOSE, metabolism of, 49

GAMMAGLOBULINS, 95; antibody, 97, physical properties, 98

GAMMA RAYS, effects on peripheral blood, 125, from radium, 128, 127

GASES, blood, 120-122 (See also specific gases)

GASTROPHILUS INTESTINALIS, larva 88, larva and pupa, 49

GAYAL, 45

GENEALOGY, of formed elements, 57, 58, 69

GENETIC DETERMINANTS (See Genotypes) GENOTYPES, in blood factor systems, 25, 28, 31, 33-36

GERMANIN, BAYER 205, 143

GERMANY, 26, 32, 39

GIBBON, 45

GIRAFFA CAMELOPARDALIS, 45

GIRAFFE, 45

GLOBULIN(S), 11, 13, 15, 21, 29, 95-98; accelerator, 21, alpha<sub>2</sub>-bilirubin, 97, 98, alpha<sub>2</sub>, 95, antithrombin, 11, 15, 97, 98, beta<sub>2</sub>, 95, beta<sub>2</sub>, 98, cold-insoluble, 88, plasma accelerator, 13, 15, serum accelerator, 13, 15, total in human plasma, 95

GLUCOSAMINE, 89, 102

GLUCOSE, 88, 89, 136, metabolism of, 49

GLUCOSE-6-PHOSPHATE DEHYDROGENASE, 115

GLUCURONIDASE, beta-, 115

GLUTAMIC ACID, 89, 102

GLUTAMINE, 101, 102

GLUTARIC ACID, alpha-keto-, 91

GLUTATHIONE, 102, 108

GLYCINE, 99, 102

GLYCOGEN, 89

GLYCOLYSIS, 137

GLYCOPROTEIN(S), alpha<sub>2</sub>-, 88

GLYCOLALASE, 115

GNU, 45

GOAT, 3, 6, 8, 9, 10, 43, 45, 46, 56, 87, 92, 93, 96, 99, 100, 103, 105, 106, 108, 109, 125

GONADOTROPHINS, 112

GOOSE, 8, 43, 48, 90, 103-107

GOOSEFISH, 44

GRANULOCYTES, 70

GRANULOCYTES (See also specific granulocytes) in granulocytes, 87-89, morphology, 55

GRASSHOPPER, 89

GREENLAND, 26, 32

GROUPS, blood, 26-36

GROWTH HORMONE, 112

GUINEA PIG, 7-10, 21, 23, 42, 43, 53, 88, 89-92, 98, 103-106, 110-113, 116, 117, 118, 126, 128

GURNARD, 8

HAGFISH, 44

HAMSTER, 45, 117

HARE, 45

HAWAII, 28, 39

HBO, 120

HEDGEHOG, 117

HELARCTOS MALAYANUS, 45

HEMATOCRIT, 37, 38, 41-44, 48, human,  
birth to maturity, 34, fetus, newborn, 45,  
human, during labor, pregnancy, 41  
HEMATOPOIETIC TISSUES, effects of  
radiation on, 129-135  
HEMOCTOBLAST, 67, 70, 75 (See also  
myeloblasts)

HEMOGLOBIN COEFFICIENT, erythrocyte,  
37

HEMOGLOBIN CONCENTRATION, -in blood,  
at altitude, 40, 41, 42, 43, 44, 45, 46, 47, 48,  
newborn, 46, pregnancy, labor, 30, fetal,  
41, -in erythrocyte, 37-43, 46, 47, 48,  
at altitude, 40, birth to maturity, 38, fetal,  
partum, 41, -in plasma, changes in  
stored blood, 137

HEMOHISTOBLAST, 70, 75, 77 (See also  
reticulum cell)

HEMOLYSIS, in fragility test, 10 in stored  
blood, 137

HEMOPOIETIC (See Hematopoietic)

HEMOPHILIC, blood clotting, 97  
HEN, 4, 112, 119

HEPARIN RETARDED CLOT TIME,  
17

HEPARIN TOLERANCE vs. clotting time,  
37

HEREDITY, 27, 28, 32

HERPESIES INCHUMON, 45

HEXOKINASE, 117

HEXOKONATE, 89

HINDUS, 26, 32

HIPPOTAMUS AMPHIBIUS, 45

HISTAMINE, 102, 139

HISTIDINE, 95, 102

HISTIOCYTES, 63, 70

HISTOCHEMICAL PROPERTIES, blood cell  
types, 86

HOG, 45

HOC-DEER, 45

HOLLAND, 30

HOMARIUS VULGARIS, 118

HONEYBEE, 118

HORMONES, 97, 112, 143 (See also specific  
hormone)

HORME (See also Mare), 5-10, 23, 47, 45,  
54, 81-87, 89, 99-101, 102, 104, 105,  
108-111, 116-118

HYALIN SYSTEM (groups), 28, 29

HYALINA STRIATA (groups), 28, 29

HYALOMERE, 36

HYDROCHLORUS HYDROCHLORUS, 45

HYDROXYPROLINE, 102

HYDROXYLASE, 102

HYPERTENSIVE AGENTS, 139

HYRAX, 45

HYSTRIC CRISTATA, 45

INDIA, 39, 40

INDIANS, American, 26, 30, 32 Argentine  
Toba, 26, Asiatic, 30, Mexican, 30,  
Ute, 30

INDICAN, 102

INDONESIANS, 30

INOSITOL, 110, 143

INSULIN, 112

INTERMEDIUM, 112

IODINE, 118

IODOLOGOLBULIN, 97, 98

IONS (-) -total, in human arterial blood,  
121 -total, 121

ISOGLUTAMINE, 97, 98

ISO-ELECTRIC PH, 98

ISOLEUCINE, 98, 102

ISOPHOSPHATE, 118

ITALIANS, 26, 32

JAPANESE, 28, 30, 32

JUXTANUCLEAR BODIES, 88

KININ (groups) 33

KANDAROO, 45, 102-108

KARYOBLAST, 70

KELL, -cellular system, 33, -negative  
phenotypes, 31, -positive phenotypes, 33

KETONIC ACIDS, alpha-, 91

LABILE FACTOR, 12

LABILIZING ACTION, 12, 14

LABOR (childbirth), 41

LACTATE, 49, 118

LACTATION, 49

LACTIC ACID, 49, 81

LACTIC DEHYDROGENASE, 113

LACTOGENIC HORMONE, 112

LACTOSE, 49

LAMB, 10, 112

LAMPREY, 44

LANGER, 45

LE SYSTEM (groups), 34

LEAD, 119

LECITHIN, 20, 92

LEMLU, 10, 45

LEONTOCEBUS ROSALIA, 41

LEPIDOSTEUS OSSEUS, 118

LEPIDOTRIS, 45

LEUCINE, 99, 102

LEUCYLGLYCYLGLYCINE, 97

LEUKEMIA, 61

LEUKOBLAST, 64, 70, 76, 77

LEUKOCYTE(S), 50-56, 70, 76, 79, 89, 90,  
119 (See also specific leukocytes)

LEWIS SYSTEM (groups), 34

LIV, 112

LION, 45

LIPASE, 115

LIPIDS, 15, 80, 92, 97

LIPROTEIN, alpha-, beta-, 87, 98

LLAMA, 10

LOBOCYTE, 70

LOPHUS PISCATORIS, 44

LORIS TARDIGRADUS, 44

LOUISIANA, 28

LOXODONTA AFRICANA, 45

LT, 112

LU SYSTEM (groups), 35

LUTEAL PHASE, 112

LUTEINIZING HORMONE, 112

LUTEOTROPIN, 112

LUTHERAN SYSTEM (groups), phenotypes,  
35

LUTRA LUTRA

LYMPH NODES, 129, 130-133

LYMPHOBLAST(S), 69, 70, 76, 79, 86

LYMPHOCTE(S), 50-54, 58, 61, 62, 68-75,  
78, 79, 89, 90, 92, 93, 94, 95, 96, 97, 98, 99,  
100, 101, 102, 103, 104, 105, 106, 107, 108, 109,  
110, 111, 112, 113, 114, 115, 116, 117, 118,  
119, 120, 121, 122, 123, 124, 125, 126, 127, 128,  
129, 130, 131, 132, 133, 134, 135, 136, 137, 138,  
139, 140, 141, 142, 143, 144, 145, 146, 147, 148,  
149, 150, 151, 152, 153, 154, 155, 156, 157, 158,  
159, 160, 161, 162, 163, 164, 165, 166, 167, 168,  
169, 170, 171, 172, 173, 174, 175, 176, 177, 178,  
179, 180, 181, 182, 183, 184, 185, 186, 187, 188,  
189, 190, 191, 192, 193, 194, 195, 196, 197, 198,  
199, 200, 201, 202, 203, 204, 205, 206, 207, 208,  
209, 210, 211, 212, 213, 214, 215, 216, 217, 218,  
219, 220, 221, 222, 223, 224, 225, 226, 227, 228,  
229, 230, 231, 232, 233, 234, 235, 236, 237, 238,  
239, 240, 241, 242, 243, 244, 245, 246, 247, 248,  
249, 250, 251, 252, 253, 254, 255, 256, 257, 258,  
259, 260, 261, 262, 263, 264, 265, 266, 267, 268,  
269, 270, 271, 272, 273, 274, 275, 276, 277, 278,  
279, 280, 281, 282, 283, 284, 285, 286, 287, 288,  
289, 290, 291, 292, 293, 294, 295, 296, 297, 298,  
299, 300, 301, 302, 303, 304, 305, 306, 307, 308,  
309, 310, 311, 312, 313, 314, 315, 316, 317, 318,  
319, 320, 321, 322, 323, 324, 325, 326, 327, 328,  
329, 330, 331, 332, 333, 334, 335, 336, 337, 338,  
339, 340, 341, 342, 343, 344, 345, 346, 347, 348,  
349, 350, 351, 352, 353, 354, 355, 356, 357, 358,  
359, 360, 361, 362, 363, 364, 365, 366, 367, 368,  
369, 370, 371, 372, 373, 374, 375, 376, 377, 378,  
379, 380, 381, 382, 383, 384, 385, 386, 387, 388,  
389, 390, 391, 392, 393, 394, 395, 396, 397, 398,  
399, 400, 401, 402, 403, 404, 405, 406, 407, 408,  
409, 410, 411, 412, 413, 414, 415, 416, 417, 418,  
419, 420, 421, 422, 423, 424, 425, 426, 427, 428,  
429, 430, 431, 432, 433, 434, 435, 436, 437, 438,  
439, 440, 441, 442, 443, 444, 445, 446, 447, 448,  
449, 450, 451, 452, 453, 454, 455, 456, 457, 458,  
459, 460, 461, 462, 463, 464, 465, 466, 467, 468,  
469, 470, 471, 472, 473, 474, 475, 476, 477, 478,  
479, 480, 481, 482, 483, 484, 485, 486, 487, 488,  
489, 490, 491, 492, 493, 494, 495, 496, 497, 498,  
499, 500, 501, 502, 503, 504, 505, 506, 507, 508,  
509, 510, 511, 512, 513, 514, 515, 516, 517, 518,  
519, 520, 521, 522, 523, 524, 525, 526, 527, 528,  
529, 530, 531, 532, 533, 534, 535, 536, 537, 538,  
539, 540, 541, 542, 543, 544, 545, 546, 547, 548,  
549, 550, 551, 552, 553, 554, 555, 556, 557, 558,  
559, 560, 561, 562, 563, 564, 565, 566, 567, 568,  
569, 570, 571, 572, 573, 574, 575, 576, 577, 578,  
579, 580, 581, 582, 583, 584, 585, 586, 587, 588,  
589, 590, 591, 592, 593, 594, 595, 596, 597, 598,  
599, 600, 601, 602, 603, 604, 605, 606, 607, 608,  
609, 610, 611, 612, 613, 614, 615, 616, 617, 618,  
619, 620, 621, 622, 623, 624, 625, 626, 627, 628,  
629, 630, 631, 632, 633, 634, 635, 636, 637, 638,  
639, 640, 641, 642, 643, 644, 645, 646, 647, 648,  
649, 650, 651, 652, 653, 654, 655, 656, 657, 658,  
659, 660, 661, 662, 663, 664, 665, 666, 667, 668,  
669, 670, 671, 672, 673, 674, 675, 676, 677, 678,  
679, 680, 681, 682, 683, 684, 685, 686, 687, 688,  
689, 690, 691, 692, 693, 694, 695, 696, 697, 698,  
699, 700, 701, 702, 703, 704, 705, 706, 707, 708,  
709, 710, 711, 712, 713, 714, 715, 716, 717, 718,  
719, 720, 721, 722, 723, 724, 725, 726, 727, 728,  
729, 730, 731, 732, 733, 734, 735, 736, 737, 738,  
739, 740, 741, 742, 743, 744, 745, 746, 747, 748,  
749, 750, 751, 752, 753, 754, 755, 756, 757, 758,  
759, 760, 761, 762, 763, 764, 765, 766, 767, 768,  
769, 770, 771, 772, 773, 774, 775, 776, 777, 778,  
779, 780, 781, 782, 783, 784, 785, 786, 787, 788,  
789, 790, 791, 792, 793, 794, 795, 796, 797, 798,  
799, 800, 801, 802, 803, 804, 805, 806, 807, 808,  
809, 810, 811, 812, 813, 814, 815, 816, 817, 818,  
819, 820, 821, 822, 823, 824, 825, 826, 827, 828,  
829, 830, 831, 832, 833, 834, 835, 836, 837, 838,  
839, 840, 841, 842, 843, 844, 845, 846, 847, 848,  
849, 850, 851, 852, 853, 854, 855, 856, 857, 858,  
859, 860, 861, 862, 863, 864, 865, 866, 867, 868,  
869, 870, 871, 872, 873, 874, 875, 876, 877, 878,  
879, 880, 881, 882, 883, 884, 885, 886, 887, 888,  
889, 890, 891, 892, 893, 894, 895, 896, 897, 898,  
899, 900, 901, 902, 903, 904, 905, 906, 907, 908,  
909, 910, 911, 912, 913, 914, 915, 916, 917, 918,  
919, 920, 921, 922, 923, 924, 925, 926, 927, 928,  
929, 930, 931, 932, 933, 934, 935, 936, 937, 938,  
939, 940, 941, 942, 943, 944, 945, 946, 947, 948,  
949, 950, 951, 952, 953, 954, 955, 956, 957, 958,  
959, 960, 961, 962, 963, 964, 965, 966, 967, 968,  
969, 970, 971, 972, 973, 974, 975, 976, 977, 978,  
979, 980, 981, 982, 983, 984, 985, 986, 987, 988,  
989, 990, 991, 992, 993, 994, 995, 996, 997, 998,  
999, 1000

MARMOTA CALIGATA, 45

MARRAS, effects of X-rays on, 129-135

MART CELLS, 60, 84, 86

MASAMA SATORI, 45

MEGAKARYOBLAST, 70, 80, 81

MEGAKARYOCYTE(S), 63, 67-70, 71, 74,  
75, 81, 86

MEGALOCYTOBLAST, 68, 69

MEGALOCYTE, 68, 69

MEGALOCARYOBLAST, 70

MEGAPTERA NOVACANALIA, 45

MELLIOWTA CAPENSIS, 45

MENHADEN, 44

MERCAPTALAN, 97, 98

MESENCYME, 47, 68, 77 (See also  
pelliculum cell)

METACHROMATIC GRANULES, 55

METAKARYOBLAST, 70

METAKARYOCYTE, 70

METAKARYOCTE(S), 68-75, 76, 79, 86,  
genology of, 68, 69, histochemical  
properties of, 66, morphology of, 76, 79,  
values, 71-75

METACRYLIC, 70-75, 82, 83

METHANOGLOBIN, 49, 115

METHIONINE, 99, 102

METHYL QUININE, 101

METOPRYL, 128

MEXICO, 40

MINERALS, 117-119 (See also specific  
minerals)

MINK, 48

MITOCHONDRIA, 86

MNS BLOOD FACTORS, 32

MNS SYSTEM (groups), 31

MOLE, 45

MOLLUSCA, 85, 118

MONGOOSE, 45

MONKEY, 1, 23, 43, 45, 89, 93, 95, 103-106,  
111-113, 117, 118

MONOCYTE, 68-70, 80, 81

MONOCYTE(S), 50-54, 56, 62, 67-70, 72-75,  
genology of, 62, count 50-54,  
properties and functions, 62, 86 stain re-  
actions of, 62 values 72-75

MONOSODIUM ANTIMONY THIOGLYCOL-  
LATE, 141

MORPHOLOGY, of blood and bone marrow  
cells, 55, 76-85 (See also specific  
leukocytes)

MOUNTAIN MOSCHIFERUS, 45

MOUNTAINOUS REGIONS, 122

MOUSE, 5, 7, 10, 23, 42, 45, 55, 89, 93, 96,  
98-100, 102-106, 109-113, 118, 119, 126,  
130, 131

MUCOPROTEIN ALPHA<sub>2</sub>, 97, 98

MUS MUSCULUS, 45

MUSTELA 45

MYANESIN 139

MYELOBLAST, 68-77, 86 genology of,  
86, 89 histochemical properties of, 86,  
morphology of, 79, 81 values, 70-73

MYELOCTE(S), 67-77, 86 genology of,  
67-69, histochemical properties of, 86,  
morphology of, 79, 81 values, 71-75

MYELOBLAST CELL(S), 68, 70

MYRMECOPHAGA TRIDACTYLA, 45

MYRINE GLUTINOSA, 44

NARCOTICS, aliphatic, 138

properties and functions, 57, 86, stain reactions of, 57; values, 71, 73-75  
 NEWBORN ANIMALS, 46, 47, 85  
 NICOTINAMIDE, 102  
 NICOTINIC ACID, 102, 110, 143  
 NITROGEN COMPOUNDS, 101, 102 (See also specific compounds)  
 NITROGEN GAS, 120  
 NITROUS OXIDE, 138  
 NOR-EPIPHENEPHRI, 112  
 NORLEUCINE, 102  
 NORMOBLASTS, 85, 70-75, 80-83, 85, genealogy, 85, histochemical properties, 85, values, 71-75  
 NORMOCYTE, 70  
 NORWAY, 39  
 NUCLEAR MEMBRANE, 55, 56, 77, 79, 81, 83, 85  
 NUCLEI, 77, 79, 81, 83, 85, 86  
 NUCLEOTIDES, 80, 101  
 NYCTALUS NOCTULA, 45

OCELOT, 45  
 ODOCOILEUS VIRGINIANUS, 45  
 OPIUM, 45, 88, 103-106  
 ORANG UTAN, 45  
 OREGON, 39  
 ORNITHINE, 102  
 ORYCTOLAGUS CUNICULUS, 45  
 OSMOTIC PRESSURE, colloidal, in serum, 6  
 OSMOTIC REGULATION, blood volume, 97  
 OTOCYON MEGALOTIS, 45  
 OTTER, 45  
 OUBAIN, 138  
 OVIS ARIES (See Sheep)  
 OVULATION, hen, 112  
 OWL, 85  
 OX, 8-8, 10, 88, 103-106, 110, 111, 113, 118, 119  
 OXOPHENARSINE HYDROCHLORIDE, 142  
 OXYGEN, content and pressure, 120-124, consumption, 48  
 OXYTICIN (See Pilocin)

P SYSTEM (groups), 38  
 PABA, 102, 111, 143  
 PACKED VOLUME, erythrocytes (See Hematocrit)  
 PAGUMA LARVATA, 45  
 PALUDRINE, 141  
 PAMAQUINE NAPHTHOATE, 141  
 PAN TROGLODITES, 45  
 PANTOTHENIC ACID, 102, 110, 143 (See also Coenzyme A)  
 PAPIO DOGUERA, 45  
 PAPUANS, 39  
 PARA-AMINOBENZOID ACID, 102, 111, 143  
 PARA-CHLOROPHENOL, 143  
 PARACHROMATIN, 77, 79, 81, 83, 85, pattern, 55, 56  
 PARALDEHYDE, 112  
 PARATHORMONE, 102  
 PARENTS vs. CHILDREN, ABO groups, 27, subgroups, 28, MN factors, 32  
 PATERKITY, disputed, application of blood group heredity, 27, 28, 32  
 PATHOGENS, antibodies for, 97  
 PATHOLOGIC LARGE LYMPHOCYTE, 70  
 PATHOLOGICAL LYMPHOCYTE, 78, 79  
 PENCILLIN, 140  
 PENTAQUINE MONOPHOSPHATE, 141  
 PHENOBARBITAL, 138  
 PENTOSE, 89  
 PEPTIDASE, 47, 98  
 PERIPHERAL BLOOD, effects of radiation on, 125-128  
 PEROXIDASE, histochemical tests for, 86  
 PERU, 39, 40  
 PETAURISTA PETAURISTA, 45  
 PETROMYX MARINUS, 44  
 pH, changes in preserved and stored blood, 138, erythrocyte, 37, isoelectric, 98, plasma, 124  
 PHAGOCYTE, 69  
 PHASCOLOMUS LELINUS, 45

PHENOBARBITAL, 138  
 PHENOLSULPHATASE, 113  
 PHENOTYPE(S), in ABO system, 25, in K system, 33, in Le system, 34, in Lu system, 35, in MNS system, 31, in Rh system, 29, in P system, 36  
 PHENYLALENE, 100, 102  
 PHLODYAS, 85  
 PHOCA VITULINA, 45  
 PHOCAENA PHOCAENA, 45  
 PHOSPHATASE, acid, 86, 115, alkaline, 86, 97, 115  
 PISCES, 8 (See also specific fish)  
 PIT VIPER, 88  
 PITOCIN, 112  
 PITRESSIN, 112  
 PLAICE, 7, 8  
 PLASMA, 5-7, 13-15, 21, 25, 36, 87-101, 103-107, 123, 124, 138-143  
 PLASMA CELL, 85, 97, 10 (See also Plasmacyte)  
 PLASMA PROTEIN(S), 84 (See also specific proteins), properties, 88  
 PLASMABLAST(S), 70, 84, 85  
 PLASMACTOBLAST(S), 69, 71, 72, 74, 75, properties and functions, 85, 88  
 PLASMACTOBLAST, 69  
 PLASMACTOBLAST, 70  
 PLASMACTOBLASTIC LYMPHOCYTE, 70  
 PLASMIN, 97, 98  
 PLASMINOGEN, 97, 98  
 PLATELET(S), 87, 88, 70, properties and functions, 64, 86, -accelerator(s), 13, 14, -activator, 14, -counts, 22, 23, 38, -disintegration, 11-15, morphology, 56  
 PLEUROMYX AURITUS, 45  
 PLEUROMYXES PLATESSA, 7, 8  
 PMS, 113  
 P-PHENOTYPES, 36  
 PO<sub>4</sub> PHOSPHOROUS, 102  
 POLECAT, 45  
 POLYMORPHONUCLEAR, 70  
 POLYPEPTIDES, 101  
 POLYPEPTIDIC THEORY, 69  
 POLYSACCHARIDES, 89  
 PONDO SATYRUS, 45  
 POPULATIONS, distribution of blood groups in, 28, 30, 32  
 PORCUPINE, 45  
 PORPOISE, 45  
 POSTPARTUM, human, 41, 52, 112  
 POTASSIUM, 117, 138, -acetate, 129, -bicarbonate, 139  
 PREGNANCY, human, 41, 52, 73, 89, 112  
 PREGNANT MARE'S SERUM, 112  
 PREMAGALOKARYOCYTE, 70  
 PREMONOCYTE, 70  
 PREMYELOCYTE, 70  
 PRESBYTIS PYRUS, 45  
 PRESERVED BLOOD, changes in, 135, 137  
 PRESSURE, colloidal osmotic, 6  
 PRIMITIVE FREE CELL, 88  
 PRIONOTUS, 44, 48  
 PROACCELERIN, 11  
 PROCAINE HYDROCHLORIDE, 138  
 PROCAVIA CAPENSIS, 45

PROCONVERTIN, 11  
 PROCYON LOTOR, 45  
 PRODUCTION RATE, erythrocytes, 37  
 PROERYTHROBLASTS, 71-75, 80, 81, 86  
 PROFERINOLYSIS, 119  
 PROGESTERONE, 112  
 PROGRANULOCYTE(S), 70-72, 76, 77, 88  
 PROKARYOCYTE(S), 70  
 PROLACTIN, 112  
 PROLINE, 100, 102  
 PROLYMPHOCYTE(S), 70, 78, 79  
 PROMEGAKARYOCYTE(S), 70, 80, 81  
 PROMEGALOBAST(S), 68, 70, 82, 83  
 PROMONOCYTE, 70  
 PROMYELOCYTE(S), 68, 70, 76, 77 (See also Myelocyte, early)  
 PRONORMOBLAST(S), 68, 70, 80, 81  
 PROPERTIES (See specific properties)  
 PROPLASMACYTE, 84, 85  
 PROPRUBRACYTE, 70, 80, 81, pernicious anemia type, 82, 83  
 PROTEIN(S), 94, 95, 97, 102, 118, alpha-, 80, beta-, 97, 99  
 PROTEINODON RUFOGROSSES, 45  
 PROTHROMBIN, 11-15, 21, 87, 88, -conversion factor, 14, -time, 20  
 PROTHROMBINOGEN, 12  
 PROTHROMBOPLASTIN, 11  
 PSEUDOGLOBULIN(S), 95  
 PSEUDOPREGNANCY, rabbit, 112  
 PTEROPUS DASYMALLUS, 45  
 PTEROYLGLUTAMIC ACID, 102, 111, 143  
 PUERTO RICANS, 30  
 PUFFER, 44  
 PYCATHRIX ADURATA, 45  
 PYRIBENZAMINE, 139  
 PYRIDINE NUCLEOTIDE(S), 80, 116  
 PYRIDOXIN, 102, 111  
 PYRUVATE, 118  
 PYRUVIC ACID, 91  
 QUINACRINE, 141, -hydrochloride, 141  
 QUINIDINE SULFATE, 139  
 QUININE, salts, 141  
 RABBIT, 5-10, 20, 21, 23, 42, 43-49, 53, 87-93, 96, 99, 103-113, 115, 117-119, 127, 131-133  
 RACCOON, 45, 103-108  
 RADIATION, effects on blood, 128-135, from radium, 126, 127  
 RAJA, 44  
 RANA (See Frog)  
 RAT, 5, 7-10, 21, 23, 42, 43-47, 53, 87-93, 96, 99-101, 103-113, 115-118, 126, 133, 134  
 RATUFA INDICA, 45  
 RAY, 6  
 RED BLOOD CELL (See Erythrocyte)  
 REDUCTASE, methemoglobin, 115  
 REFRACTIVE INDEX, serum, 7  
 RELAXANTS, skeletal muscle, 138  
 RELAXIN, 112  
 REPTILES, 8, 91 (See also specific reptiles)  
 RESTING WANDERING CELL(S), 70  
 RETICULAR CELL(S), non storing, 87, 89, 76, 77, phagocytic, 67  
 RETICULAR PLASMA CELL(S), 70, 84, 85  
 RETICULAR-ENDOTHELIAL CELL(S), 74, 77 (See also Reticulum cell)  
 RETICULO-ENDOTHELIAL SYSTEM, 69  
 RETICULO-ENDOTHELIAL, 68  
 RETICULO-LYMPHOCYTE(S), 68  
 RETICULO-LYMPHOCYTES, 70, -count, 38, 42  
 RETICULUM CELL(S), 69, 69, 71, 72, 74, 75, 76, 77, 86  
 RHABDOCYTE, 70  
 RHESUS MONKEY (See Monkey)  
 Rh-System (groups), 29, 30  
 RHINOCEROS, 45  
 RHOMBUS MENES, 8  
 RIB EXTRUSIONS, differential counts in marrow, 74, 75  
 RIBOPHAGIN, 102, 111, 143, -bound, 90, 118  
 RIBONUCLEIC ACID, 90  
 RIBONUCLEOPROTEIN, 86  
 ROD NUCLEAR CELL, 70

- ROOSTER, 112  
ROUSSETTUS LEACHII 45  
RUBRIBLAST(S) (See also Proerythroblasts),  
70, 80, 81, pernicious anemia type, 82,  
83 (See also Promegaloblast)  
RUBICAPRA 10, 20-83  
RUPICAPRA RUBICAPRA, 45  
RUSSIA, 96  
RUSSIAN Viper VENOM 20  
RUSSIANS, 32
- SAMURI SCIUREUS, 45  
SALMON, 118, 128, 135  
SALMONELINIS FONTINALIS, 43, 44  
SAMBAR 45  
SATURATION PERCENTAGE, oxygen, 132,  
132  
SCIRUS, 45  
SCORBER SCORBER, 8, 43, 44  
SCORPION FISH, 8  
SCYLLOPHILUS CANICILLIS, 8  
SCYLLUM (See specific elasmobranchs)  
SEA LEVEL, 39, 40, 52, 121  
SEA ROBIN, 44, 48  
SEAL 45  
SEACAL, 138  
SEDIMENTATION CONSTANTS, proteins, 98  
SEDIMENTATION RATE, 9  
SEGMENTED CELLS, 50, 51, 56, 70-75  
SERINE CELL, 70  
SERINE, 102  
SEROLOGICAL AGENTS, 97  
SERUM, 5-8, 87-89, 91-93, 95, 101, 107, 113,  
115, 117-119, 128-143 -accelerator, 13-15,  
-albumin, 98  
SHARK SUCKER, 44  
SHEEP (See also Lamb) 5-10, 23, 43, 45-47,  
116, 118  
SILICON, 119  
SILVER, 119  
SIPUNCULUS, 89  
SKATE, 44  
SKELETAL MUSCLE RELAXANTS, 139  
SLOW, 45  
SLOW, bleeding time of, 18  
SMEAR CELL, 70  
SMUDGE, 20  
SNAIL, 43, 48, 90, 103, 105, 111  
SODIUM, 117, 136, -bromide, 138, -iodide,  
140 -pentothal, 138, -sulfate, 139  
SOLUBLE values of in blood constituents, 87  
SOLUBLETHAN, 142  
SOUTH AFRICA 40  
SOW pregnant 112  
SPECIFIC GRAVITY 5 37  
SPHEROIDIC INDEX erythrocyte 98  
SPHEROIDIC ASCULATUS 37  
SPHONGOMYELIN 92  
SPINAL CORD MACULATUS 44  
SPLEEN 130-135  
SQUALUS ACANTHIAS (See Dogfish)  
SQUIDREL, 45
- STAB CELL, 70  
STARCH, 70  
STAFF CELL, 70  
STAIN REACTIONS, 57-68  
STAIN CELL, 70, 77 (See also  
reticulum cell)  
STERNUM, marrow of differential count in,  
71, 73 effects of radiation on, 129  
STERILIZATION, 142  
STINGRAY, 21, 44  
STORED BLOOD, 136, 137  
STREPTOKINASE, 97  
STREPTOMYCIN, 140  
STRIGON, 44  
SUCCINIC ACID, 91  
SULFA COMPOUNDS, 142  
SULFATE, 118 -compounds, 108  
SULFADRIAL GROUPS 97  
SULFONAMIDES, 142  
SULFUR compounds, 108 (See also specific  
compounds)  
SUNUS INDICES 45  
SURAMIN SODIUM, 143  
SURFACE AREA erythrocyte, 37  
SUS SCROFA, 45  
SWEDEN, 26, 32  
SWINE, 84, 96 116  
SWITZERLAND, 40
- TACHYGLOSSUS ACULEATA, 45  
TALPA ELROPAEA, 43  
TAMARIN, 45  
TAPIR 45  
TAPIRUS INDICES, 45  
TAUTOG, 44  
TAUTOGA ONITIS 44  
TEMPERATURE vs clotting time, 16  
TENCH, 7 8  
TENSURE, 45  
TENSION blood gases, 121 122  
TERMINOLOGY 70  
TERRAMYCIN HYDROCHLORIDE 141  
TESTOSTERONE, 112  
TH 102, 112, 143  
THALACTOS MARITIMUS, 45  
THERAPEUTIC AGENTS, 130-143  
THERMINE 102 111 -bound 116  
THIOCYANATE, 108  
THIOPIANET SODIUM, 138  
2-THIOLACIL, 140  
THROMBIA 140  
THROMBIN 100, 102  
THROMBIN 11-15, 87  
THROMBOCYTES (See Platelets)  
THROMBOPLASTIC 70  
THROMBOPLASTIN, 11-15, 20 87  
THROMBOPLASTOGEN, 12, 14  
THROMBOPLASTINASE, 12  
THROMBOTONIN, 12  
THYLACOMYS LAGOTIS, 45  
THYROID-ACTIVE PROTEIN, 97, 98  
THYROID ACTIVITY, 102 112, 143  
THYROID HORMONE, 97, 98, 112  
THYROTROPIC HORMONE, 97, 98, 112  
TIGER, 45
- TIN, 119  
TINCA VULGARIS, 7, 8  
TISSUE(S), in blood coagulation, 11-15,  
hematopoietic, 129-133  
TISSUE EOSINOPHILS, 39  
TISSUE MAST CELL(S), 40, 84, 85  
TOAD, 117  
TOCOPHEROL (See also Vitamin E), 143  
TORPEDO MARMORATA, 8  
TOXIN, diphtheria, 87  
TRAGILLUS JAVANICUS, 45  
TRICHOMETHANOL, 138  
TRICHOSIA VILPECELLA, 45  
TRIGLA LUCERNA, 8  
TRIPLELLEAMIDE 139  
TROUT, 43, 44  
TRYPARSAMIDE, 143  
TRYPTOPHAN 100, 102  
TSH, 97, 98, 112  
2-THIOCLARINE CHLORIDE, 139  
TUMOR CELL, 70  
TURK BRITANNIA FORM, 70  
TURBOT, 8  
TURKEY, 26, 45, 103-109, 111  
TURKLE, 6, 8, 21, 43, 48, 90, 91, 103-107,  
109, 116  
TYPHOID "O" AGGLUTININS 97  
TYROSINE, 100, 102
- UNCLASSIFIED CELLS, 71, 72, 74 75  
UNNA'S PLASMA CELL, 70  
UREA, 101, 103  
URIC ACID, 101, 102  
URETHANE, 142  
URUS 45  
UNITED STATES, 25, 30, 32
- VALINE 100, 102  
VASOCONSTRICTOR(S), 11-15  
VASOPRESSIN, 112  
VENTERATES, 116 125-135 (See also  
specific vertebrates)  
VESPERTILLO MURINUS 45  
VINETHANE, 138  
VINYL ETHER, 138  
VISCOSITY, 6  
VITAMIN(S), A, 109, 143 B<sub>12</sub>, 109, B<sub>6</sub>,  
110, 143 -carriers, 97 -content 109-111  
VOLUME erythrocyte, 37, 44, 47 packed  
(See Hematocrit; regulation of osmotic, -97  
VULPES VULPES 45
- WALLABY, 45  
WATER, 87  
WAXMOTH, 89  
WEASEL, 45  
WHALE 45  
WHITE BLOOD CELLS (See Leukocyte see  
also specific leukocytes)  
WOLF, 45  
WOMBAT, 45  
X-RAYS, 125-135  
ZINC, 119  
ZOARCES ANGIILLARIS, 44









